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Phytoplankton diversity and trophic state of natural water reservoirs in Azagny National Park (Côte d'Ivoire)

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Abstract

Azagny National Park, located 40 kilometres from the town of Grand-Lahou, is home to numerous animal and plant species, including microalgae. The aim of this study was to determine the phytoplankton diversity and trophic status of the water reservoirs in the Azagny National Park. The physico-chemical parameters of the water were measured at stations ST1, ST2 and ST4, from March 2019 to February 2020. Nutrient salts were measured using a spectrophotometer. Phytoplankton sampling was carried out using a plankton net with a mesh size of 20 µm. A total of 134 phytoplankton taxa were inventoried, divided into 5 phyla, 8 classes, 14 orders and 32 families. The waters of the Azagny National Park could be considered rich in taxa, with the majority being Heterokontophyta (48 taxa) and Euglenophyta (39 taxa), followed by Chlorophyta (27 taxa), Cyanoprokaryota (18 taxa) and Dinophyta (2 taxa). Sorensen's index or coefficient of similarity gave values ranging from 0.89 to 0.74, showing that the Park's water reservoirs have taxa distributed in the same way in the different stations. The study also revealed that the park's reservoirs are warm and acidic, with low levels of dissolved oxygen and high levels of nutrient salts (nitrite and phosphate). The A and B index values of 9.81 and 1.62 respectively show that the park's reservoirs are tending towards a state of eutrophy.

Keywords: Diversity, phytoplankton, physico-chemistry, eutrophy, water reservoirs, Azagny

1. Introduction

The degradation of aquatic ecosystems can cause the disappearance or rarefaction of certain species and also lead to the proliferation of certain aquatic organisms or species. In Côte d'Ivoire, these disturbances have been observed in several rivers, lakes and reservoirs, and have had a major impact on aquatic biodiversity (Sweeney *et al.*, 2004) ^[1]. Coastal areas are places of human concentration where the imperatives linked to the development of agriculture and urbanisation threaten the balance of small river ecosystems. Despite studies of aquatic fauna in these regions (Ouattara *et al.*, 2000; Ouattara *et al.*, 2007) ^[2, 3], and those of certain authors in the Adzopé reservoir (Kouassi, 2013; Adon, 2013) ^[4, 5], in the Fresco lagoon (Konan, 2014) ^[6], in the Bandama river (Lozo, 2016) ^[7] and in the Grand-Lahou lagoon complex (Komoé, 2010) ^[8], data on phytoplankton remain poorly documented. Studies on phytoplankton in Azagny National Park are non-existent. In addition, the Park is subject to various types of anthropogenic pressure that can disrupt its ecological functioning. This lack of knowledge of phytoplankton is a limitation in the management of water reservoirs and the enhancement of this biodiversity. Therefore, this study aims to understand the phytoplankton diversity of the Azagny National Park as well as the trophic status of the waters in order to contribute to the conservation of its biodiversity and guarantee effective and sustainable management of resources.

2. Materials and methods

2.1 Study environment

Azagny National Park is located 130 km from Abidjan, in the prefecture of Grand-Lahou at the mouth of the Bandama River (OIPR, 1981) ^[9]. The Park covers an area of 19,400 ha and

stretches between two administrative departments (Grand-Lahou and Jacqueville) (Komoé *et al.*, 2008) [10]. The Park is part of the Upper Guinea phytogeographic region, which extends from Togo to Senegal (Atanle *et al.*, 2012) [11]. Four stations (Figure 1) were selected for this study, taking into account their distribution throughout the territory and their accessibility: ST1, ST2, ST3 and ST4.

2.2 Physico-chemical parameters

The physico-chemical parameters of the water (Temp, pH, O2, COND) were measured using a HANNA multiparameter model HI 83200 at the four stations from March 2019 to February 2020. The nutrient salts were analysed in the

laboratory and the various concentrations determined using a spectrophotometer. The principle of measurement is based on Beer-Lambert's law, which indicates the proportionality of the optical density with the thickness of the solution (Sample analysed) and the concentration of the chemical element sought.

For the statistical test, the Kruskal-Wallis test was used, which is a non-parametric alternative to the first-order ANOVA (Between groups). It is used to compare at least three samples, and to test the null hypothesis (Significant test at $p < 0.05$) that the different samples to be compared come from the same distribution or from distributions with the same median.

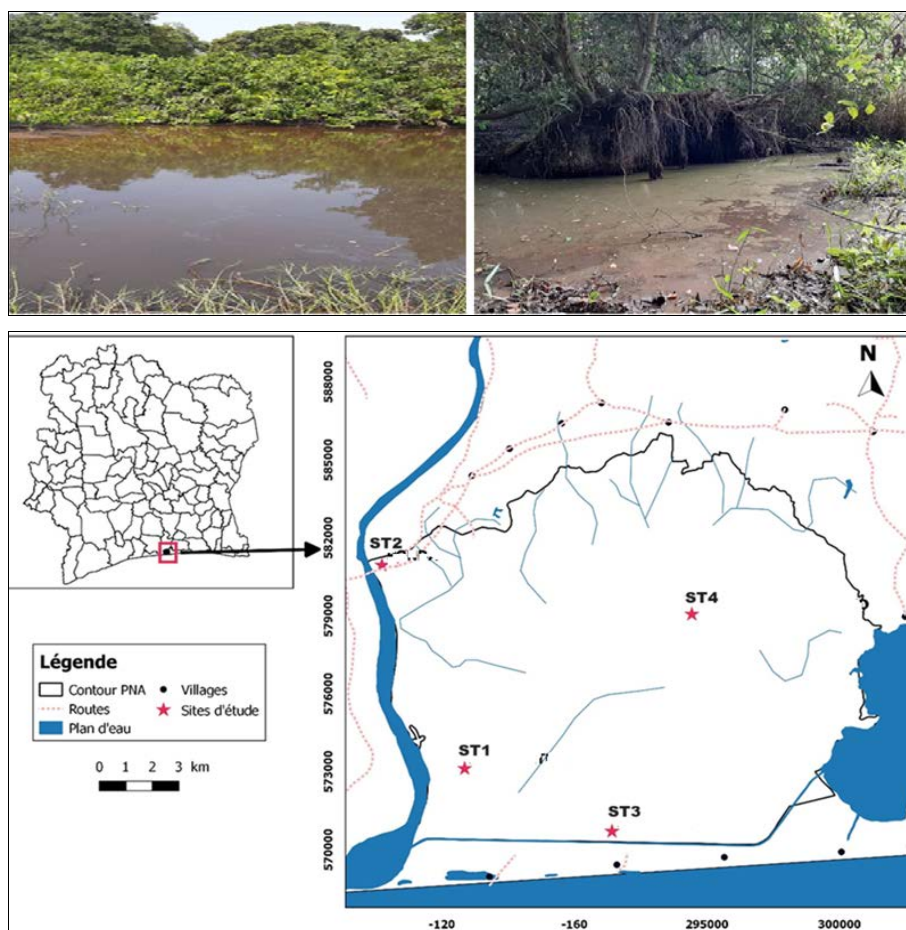


Fig 1: Map of Azagny National Park with sampling stations

2.3. Phytoplankton and trophic status

Phytoplankton were sampled using a 20 µm plankton net at each station. Water samples fixed with 5% formalin were mounted between slide and coverslip and examined under a light microscope with a 40x objective. The different species were identified using various documents (Da, 1992; Ouattara, 2000; Komoé, 2010; Adon, 2013 and Lozo, 2016) [12, 13, 8, 5, 7]. Assessment of the trophic status of the stations was based on examination of the phytoplankton population, for which the absence of a certain number of organisms is as significant as the presence, development or regression of certain species. Three indices (A, B and C) were therefore calculated:

$$A = \frac{\text{Cyanophycées} + \text{Chlorococcales} + \text{Centrales} + \text{Euglenophycées}}{\text{Desmidiacées}}$$

A = composite index

1. $0.3 < A < 1$ = oligotrophic environment

2. $1 < A < 2.5$ = mesotrophic environment
3. $2.5 < A < 5$ = eutrophic environment

A second index (B) based on the quotient of the relative number of Chlorococcales and Desmidiaceae species present in lakes has generally been used to recognize the type of trophy.

$$B = \frac{\text{Chlorococcales}}{\text{Desmidiacées}}$$

- $B < 1$ = oligotrophic environment
- $B > 1$ = eutrophic environment

A third index (C) derived from the Diatoms, translates the ratio of the number of species of the Centrales to that of the Pennales:

$$C = \frac{\text{Centrales}}{\text{Pennaes}}$$

C: 0 - 0.2 = oligotrophic environment

C: 0.2 - 3 = eutrophic environment

3. Results and discussion

3.1 Physico-chemical parameters

Parameters such as temperature (T°), dissolved oxygen (O₂), dissolved solids content (TDS), conductivity, depth and transparency varied significantly from one station to another ($p < 0.05$), whereas other parameters such as pH, nitrate content and phosphorus content did not vary ($p > 0.05$). The water in the park's reservoirs is slightly warm (27.68 °C) and acidic, with low levels of dissolved oxygen (3.52 mg/l) and high levels of nutrient salts (Nitrite and phosphate). This could be explained by a number of factors, such as the degree of isolation, wind and plant cover (Canopy), the decomposition of macrophytes and the faeces of animals that drink from the water.

3.2 Phytoplankton diversity

The algal flora of all the environments studied comprises 134 phytoplankton taxa divided into 5 phyla, 8 classes, 14 orders and 32 families (Table I). With a majority of Heterokontophyta (48 taxa or 36%) and Euglenophyta (39 taxa or 29%), followed by Chlorophyta (27 taxa or 20%), Cyanoprokaryota (18 taxa or 13%) and Dinophyta (2 taxa or 2%), Azagny National Park could be considered rich in taxa (Figure 2). This richness could be due to the fact that the waters are stagnant, which favours biological processes such as the complete reproduction and development cycles of algae (Ouattara, 2007) [14]. The taxonomy of the phytoplankton community in the park's reservoirs is dominated by Chlorophyta and Euglenophyta, which are characteristic of environments rich in nutritive salts and organic matter resulting from human activity and natural decomposition of tree trunks, dead leaves and animal excrement. The Sorensen index or similarity coefficient gives values ranging from 0.89 to 0.74. These values mean that the Park's water reservoirs have species that are distributed identically in all the stations.

Table 1: Composition and distribution of phytoplankton taxa sampled in the reservoirs of the Azagny National Park from June 2020 to September 2020

TAXA	ST1	ST2	ST3	ST4
Cyanoprokaryota Anagnostidis komárek				
Cyanophyceae Sachs				
Chroococcales Wettstein and Westerheim				
Chroococcaceae Nails				
<i>Aphanocapsa elachista</i> gs West		x		x
<i>Aphanocapsa incerta</i> Lemmermann	x		x	
<i>Chroococcus dispersus</i> (Keissler) Lemmermann	x		x	
<i>Chroococcus turgidus</i> (Kützing) Nägeli		x	x	
<i>Chroococcus</i> sp.	x		x	x
<i>Coelomonon pusillum</i> (Van goor) Komárek	x	x		
<i>Merismopedia glauca</i> (Ehrenberg) Nägeli		x		
<i>Microcystis aeruginosa</i> (Kützing) Kützing	x	x		x
<i>Microcystis incerta</i> Lemmermann	x		x	
Nostocales (Borzi) Geitler				
Nostocaceae Dumortier				
<i>Anabaena affinis</i> Lemmermann	x		x	
<i>Anabaena mucosa</i> Komarkov-legnerova and Eloranta				x
<i>Anabaena spiroides</i> Klebahn	x	x	x	x
<i>Anabaena</i> sp.	x		x	
<i>Cylindrospermopsis raciborskii</i> (Woloszynska) seenayya & Subba raju				x
Elenkin Oscillator				
Oscillatoriaceae (Gray) Bory of St. Vincent				
<i>Oscillatoria princeps</i> Gomont	x	x	x	x
Spirulinaceae				
<i>Spirulina meneghiniana</i> fo. <i>Fontinalis</i> schwabe				x
<i>Spirulina princeps</i> West & West	x		x	
<i>Spirulina</i> sp.	x		x	x
Heterokontophyta Van den Hoek and al.				
Bacillariophyceae Haeckel				
Schütt power stations				
Aulacoseiraceae Crawford				
<i>Aulacoseira granulata</i> var. <i>Angustissima</i> (o. Müller) Simonsen	x	x	x	x
<i>Aulacoseira granulata</i> var. <i>Angustissima</i> fo. <i>Spiralis</i> (o. Müller) Simonsen			x	x
Stephanodiscaceae Glezer and Makarova				
<i>Cyclotella meneghiniana</i> Kützing	x			x
Pennaes schütt				
Amphipleuraceae Grunow				
<i>Frustulia saxonica</i> Rabenhorst	x		x	x
<i>Frustulia rhomboides</i> (Ehrenberg) De Toni	x		x	x
<i>Frustulia</i> sp.	x			
<i>Nitzschia acicularis</i> (Kützing) W.smith	x			
<i>Nitzschia amphibia</i> Grunow			x	

<i>Nitzschia longissima</i> (Brébisson) Ralfs	x		x	
<i>Nitzschia perversa</i> Grunow	x			
<i>Nitzschia sigma</i> (Kützing) W.smith				x
<i>Nitzschia sigmoidea</i> (Nitzsch) W.smith		x	x	x
Achnanthidiaceae dg Mann				
<i>Achnanthidium minutissimum</i> (Kützing) Czarnecki		x		
Cocconeidaceae Kützing				
<i>Cocconeis ankobraensis</i> Foged		x		
<i>Cocconeis placentula</i> Ehrenberg			x	
Diadesmidaceae Mann				
<i>Diadesmis contenta</i> (Grunow) dgMann		x	x	
Diploneidinae				
<i>Diploneis smithii</i> (Brébisson) Cleve			x	x
<i>Diploneis</i> sp.			x	
Eunotiaceae Kützing				
<i>Eunotia</i> sp.		x		
Fragilariaceae Kützing				
<i>Fragilaria acus</i> (Kützing) Lange-bertalot				x
<i>Fragilaria crotensis</i> Kitton	x	x	x	x
<i>Fragilaria ulna</i> (Nitzsch) Lange-bertalot	x	x	x	x
Gomphonemataceae Kützing				
<i>Gomphonema brasiliense</i> Grunow				x
Naviculaceae Kützing				
<i>Gyrosigma acuminatum</i> (Kützing) Rabenhorst	x	x	x	x
<i>Gyrosigma attenuatum</i> (Kützing) Rabenhorst	x		x	
<i>Gyrosigma balticum</i> (Ehrenberg) Rabenhorst			x	
<i>Gyrosigma nodiferum</i> (Grunow) Reimer		x		
<i>Gyrosigma scalproides</i> (Rabenhorst) Cleve	x		x	
<i>Gyrosigma</i> sp.				x
<i>Navicula gastrum</i> (Ehrenberg) Kützing				x
<i>Navicula contenta</i> Grunow in van Heurck			x	
<i>Navicula subtilissima</i> Cleve		x		
<i>Navicula</i> sp.	x		x	x
Pinnulariaceae Mann in Round <i>et al.</i>				
<i>Pinnularia acrosphaeria</i> W.smith	x	x		
<i>Pinnularia brebissonii</i> (Kützing) Rabenhorst				x
<i>Pinnularia divergens</i> W. Smith	x		x	x
<i>Pinnularia maior</i> var. <i>Linearis</i> Cleve		x		
<i>Pinnularia microstauron</i> (Ehrenberg) Cleve				x
<i>Pinnularia</i> sp.	x	x		
Pleurosigmataceae Mereschkowsky				
<i>Pleurosigma diverstriatum</i> F.meister			x	x
<i>Pleurosigma salinarum</i> (Grunow) Cleve & Grunow			x	
<i>Pleurosigma strigosum</i> W. smith			x	
Rhopalodiaceae (Karsten) Topachevs'kyj & Oksiyuk				
<i>Epithemia turgida</i> (Ehrenberg) Kützing			x	x
<i>Epithemia zebra</i> (Ehrenberg) Kützing	x		x	
Stauroneidaceae Mann				
<i>Stauroneis anceps</i> Ehrenberg	x		x	
<i>Stauroneis brasiliensis</i> Metzeltin & Lange-bertalot			x	x
Coscinodiscophyceae Round & Rmcrawford				
Coscidiscals round & rmcrawford				
Coscinodiscaceae Kützing				
<i>Coscinodiscus</i> sp.				x
Synurophyceae Cavalier-smith				
Synurales Andersen				
Synuraceae Lemmermann				
<i>Mallomonas matvienkoeae</i> (Matvienko) Asmund & Kristiansen	x			
CHLOROPHYTA Cavalier-smith				
Chlorophyceae Wille in warming				
Sphaeropleales Luerksen				
Ankistrodesmaceae Korshikov				
<i>Ankistrodesmus fusiformis</i> Corda	x			
<i>Ankistrodesmus gracillis</i> (Reinsch) Korshikov	x			x
Chlorococcales Pascher				
Hydrodictyaceae (Gray) Dumortier				
<i>Pediastrum biradiatum</i> Meyen	x		x	
<i>Pediastrum duplex</i> var. <i>Gracillimum</i> West & gs West	x	x	x	x

<i>Pediastrum simplex</i> Meyen	x	x	x	
Neochloridaceae Ettl and Komárek				
<i>Tetraëdron incus</i> (Teiling) Gmsmith	x			
Scenedesmaceae Oltmanns				
<i>Crucigeniella rectangularis</i> (Nägeli) Komárek				x
<i>Desmodesmus quadricaudatus</i> (Turpin) Hegewald	x	x	x	
<i>Scenedesmus apiculatus</i> (West & Gswest) Chodat			x	
<i>Scenedesmus acuminatus</i> (Lagerheim) Chodat			x	x
<i>Scenedesmus bernardii</i> Chodat		x	x	
<i>Scenedesmus opoliensis</i> pg Richter			x	
Volvocaceae Ehrenberg				
<i>Eudorina elegans</i> Ehrenberg			x	x
<i>Pandorina morum</i> (Müller) Bory	x		x	
Conjugatophyceae Engler				
Desmidiaceae Bessey				
Closteriaceae Bessey				
<i>Closterium kuetzingii</i> Brébisson	x		x	x
<i>Closterium lineatum</i> Wolle	x	x		
Desmidiaceae Kützing ex Ralfs				
<i>Cosmarium connatum</i> Brébisson ex Ralfs				x
<i>Cosmarium reniforme</i> (Ralfs) W. archer	x			
<i>Pleurotaenium eugeneum</i> (w. B. Turner) West & West			x	x
<i>Staurastrum asperatum</i> Behre		x		
<i>Staurastrum Leptocladum</i> Nordstedt	x		x	
<i>Staurastrum tetracerum</i> Ralfs			x	
<i>Staurastrum</i> sp.		x	x	
<i>Staurodesmus convergens</i> (Ralfs) Lillier				
<i>Teilingia granulata</i> (Roy and Bisset) Bourrelly	x			
Mesotaeniaceae Rabenhorst				
<i>Gonatozygon brebissonii</i> De Bary		x	x	x
Zygnemataceae Kützing				
<i>Spirogyra</i> sp.				x
Euglenophyta Pascher				
Euglenophyceae Schoenichen				
Euglenales Engler				
Euglenaceae Stein				
<i>Euglena proxima</i> Dangeard		x		
<i>Euglena spirogyra</i> Ehrenberg		x	x	
<i>Euglena</i> sp.	x	x		x
<i>Lepocinclis acus</i> (Müller) Marin and Melkonian		x		x
<i>Lepocinclis oxyuris</i> (Schmarda) Marin and Melkonian		x		x
<i>Lepocinclis ovum</i> Ehrenberg	x	x	x	x
<i>Lepocinclis salina</i> Fefritsch	x		x	x
<i>Lepocinclis texta</i> (Dujardin) Lemmermann			x	x
<i>Phacus acuminatus</i> A.stokes			x	x
<i>Phacus cucicauda</i> Svirenko		x		
<i>Phacus indicus</i> Skvortsov			x	
<i>Phacus longicauda</i> (Ehrenberg) Dujardin	x	x		x
<i>Phacus orbicularis</i> Hübner	x	x		
<i>Phacus onyx</i> Pochmann	x	x	x	x
<i>Phacus ranula</i> Pochmann	x			
<i>Phacus suecicus</i> Lemermann		x	x	x
<i>Phacus tortus</i> (Lemermann) Skvortsov	x	x	x	
<i>Strombomonas acuminata</i> (Schmarda) Deflandre	x	x	x	
<i>Strombomonas fluviatilis</i> (Lemermann) Deflandre			x	
<i>Strombomonas treubii</i> (Wołoszyńska) Deflandre	x	x	x	
<i>Strombomonas verrucosa</i> (Daday) Deflandre	x			
<i>Trachelomonas abrupta</i> Svirenko	x	x	x	x
<i>Trachelomonas armata</i> (Ehrenberg) F.stein		x		x
<i>Trachelomonas bernardinensis</i> Vischer		x		
<i>Trachelomonas curta</i> Amcunha	x	x	x	
<i>Trachelomonas hispida</i> (Perty) Stein	x			x
<i>Trachelomonas lefevrei</i> Deflandre	x	x	x	x
<i>Trachelomonas oblonga</i> Lemermann	x		x	
<i>Trachelomonas planktonica</i> Svirenko		x		
<i>Trachelomonas radiosa</i> Fritsch	xx	x		x
<i>Trachelomonas scabra</i> Playfair	x		x	
<i>Trachelomonas similis</i> Stokes	x		x	x

<i>Trachelomonas superba</i> Swirenko (Emend.) Deflandre		x		x
<i>Trachelomonas volvocina</i> Ehrenberg	x	x	x	x
<i>Trachelomonas volvocinopsis</i> Swirenko				
<i>Trachelomonas</i> sp. 1	x	x		
<i>Trachelomonas</i> sp.2	x	x		x
<i>Trachelomonas</i> sp.3		x		x
<i>Trachelomonas</i> sp.4		x		x
DINOPHYTA Auct.				
Dinophyceae Pascher				
Haeckel's Peridinales				
Peridiniaceae Ehrenberg				
<i>Bagredinium crenulatum</i> Da, Zongo, Mascarell and Couté	x		x	
<i>Peridinium</i> sp.	x			
Total: 134	69	56	74	63

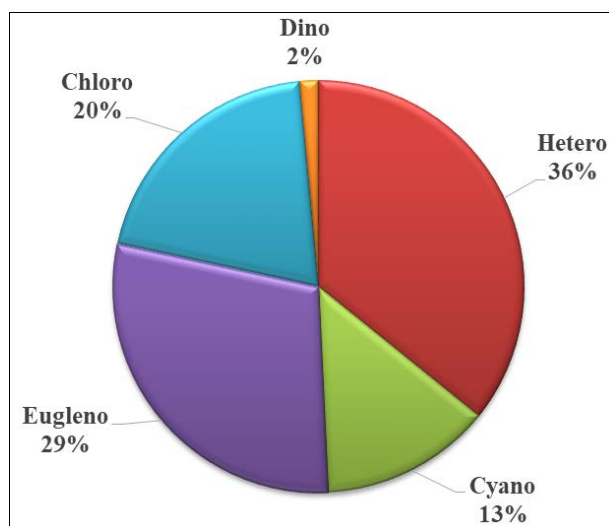


Fig 2: Proportion of the different phyla observed. Chloro: Chlorophyta; Cyano: Cyanoprokaryota; Dino: Dinophyta; Eugleno: Euglenophyta; Hetero: Heterokontophyta

3.3 Trophic status

The values of the A index and the B index, amounting to 9.81 and 1.62 respectively, indicate an eutrophic state of the water in the reservoirs. The C index, with a value of 0.11, indicates an Oligotrophic state. These results enabled us to assess the trophic status of the park's reservoirs, which tends more towards eutrophic conditions. At all the stations, of the three indices calculated, two (index A and index B) give values that show that the park's reservoirs are eutrophic and index C gives values for oligotrophic environments. Based on the values for physico-chemical parameters such as phosphate (0.32 mg/l and 0.47 mg/l), dissolved oxygen (2.00 mg/l and 6.56 mg/l), nitrate (0.18 mg/l and 0.62 mg/l) and transparency (0.07m and 0.48m), the water in the park tends to be eutrophic after comparing these values with those obtained by Mason (1991) [15]. Taking the biological aspect into account, we also note that the waters in the park are closer to the eutrophic state than to the oligotrophic or mesotrophic state. According to Brook (1965) [16], the relative number of species representing different taxonomic groups in a phytoplankton sample reflects the ecological conditions of the environment. Consequently, in eutrophic lakes, the number of species of Chlorococcales would be greater than the number of Desmidiaceae, and the opposite conditions in oligotrophic lakes (Rawson, 1956) [17]. In our study, the same observations were made (more Chlorococcales than Desmidiaceae).

4. Conclusion

This study, carried out in the reservoirs of the Azagny

National Park, revealed that 134 phytoplankton taxa were inventoried. Of the five phyla observed, two are more representative in terms of the number of taxa. These were the Heterokontophyta (35.82%) and the Euglenophyta (29.10%). This work has shown that the waters within the Park are negatively impacted by human activities in the surrounding area, on the one hand, and are influenced by natural phenomena such as the decomposition of organic matter, on the other. The physico-chemical parameters and the various indices calculated have enabled us to say that the park's water reservoirs are tending towards a state of eutrophy that is favourable to certain species of toxic cyanobacteria, which could be a danger to the emblematic and rare protected species.

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