



# International Journal of Fisheries and Aquatic Studies

E-ISSN: 2347-5129

P-ISSN: 2394-0506

(ICV-Poland) Impact Value: 76.37

(GIF) Impact Factor: 0.549

IJFAS 2023; 11(4): 11-15

© 2023 IJFAS

[www.fisheriesjournal.com](http://www.fisheriesjournal.com)

Received: 20-05-2023

Accepted: 22-06-2023

**Mahendra Prasad Bhandari**

Rainbow Trout Fishery Research  
Station, Dhunche, Rasuwa,  
Nepal

**Abhimanyu Shrestha**

Sichuan Agricultural University,  
China

**Aakriti Regmi**

Rainbow Trout Fishery Research  
Station, Dhunche, Rasuwa,  
Nepal

## Effects of dietary probiotics and probiotics combined with enzymes on growth performance of rainbow trout (*Oncorhynchus mykiss*)

**Mahendra Prasad Bhandari, Abhimanyu Shrestha and Aakriti Regmi**

DOI: <https://doi.org/10.22271/fish.2023.v11.i4a.2818>

### Abstract

Rainbow trout (*Oncorhynchus mykiss*) is a popular aquaculture species, often faces health and growth challenges due to high stocking density. To address these issues, the present study focused on evaluating the impact of dietary supplementation on the growth performance of rainbow trout using probiotics as well as probiotics combined with enzymes. Four groups of trout fry, each with an average stocking size of  $1.423 \pm 0.002$  g,  $1.412 \pm 0.096$  g,  $1.451 \pm 0.136$  g and  $1.477 \pm 0.118$  g, were individually placed in experimental tanks names T<sub>1</sub>, T<sub>2</sub>, T<sub>3</sub>, and T<sub>4</sub>, respectively. T<sub>1</sub> received a supplement of 6 gm. of Dige pro and 2 gm. of gel per kg feed. T<sub>2</sub> received a supplement of 6 gm. Multizymes and 2 gm. of gel per kg feed and T<sub>3</sub> received a supplement of 3 gm. Dige pro, 3 gm. Multizyme and 2 gm. of gel per kg feed. At the end of the 90-day period, the findings showed that dietary probiotics of 6 gm. Multizyme-p and 2 gm. Gel per kg feed (T<sub>2</sub>) showed significantly improved growth performance in terms of weight gain and survival rate compared to both the control group and other treatments.

**Keywords:** Rainbow trout, commercial probiotics, dietary supplementation, growth performance

### 1. Introduction

Aquaculture is viewed as an important food security for a growing global human population and has rapidly developed due to intensified culture method<sup>[1]</sup>. As aquaculture practices has been shifted to an intensive culture system to meet the global demand of fish food consumption, disease outbreaks are being increasingly recognized as a significant constraint of aquaculture production and trade which ultimately affects the economic development of the sector in many countries<sup>[2]</sup>. Fish diseases are caused by a number of pathogens, such as parasites, bacteria and viruses. Of these, bacteria are the leading pathogen in commercial aquaculture. Antibiotics are the most widely used agents in the treatment of fish diseases. However, their use can result in potential environmental and food contamination, as well as in the development and/or dissemination of pathogens resistance<sup>[3]</sup>.

Rainbow trout, *Oncorhynchus mykiss* is widely cultured fish species around the world. Highly stocking density adversely affects the health, survival, growth performance as well as product quality of rainbow trout<sup>[4]</sup>. As a result, there is a growing emphasis on improving fish health and growth through novel approaches, such as the use of multi-strain probiotics and probiotics with enzymes<sup>[5]</sup>. This method provides a broader spectrum of beneficial microorganism impact and synergistic interactions that can boost fish and productivity<sup>[6,7]</sup>.

The development of probiotics as an alternative or complementary of chemotherapy and vaccination to prevent fish diseases has been shown to be of great interest. Probiotics can be utilized individually or in various combinations, taking forms such as multi-strain probiotics combined with plant extracts, probiotics mixed with yeast extracts and probiotics with enzymes<sup>[8]</sup>. The use of live microorganisms can enhance the disease resistance of fish and shrimp by suppressing the pathogens, improving growth and improve immune response<sup>[2]</sup>. In the aquaculture industry, there is an increasing emphasis on the use of plant-based protein as a replacement for fish meal, in order to promote sustainability. The plant-based proteins used as substitutes for fish meal in aquaculture may contain non-starch polysaccharide compound (NSP) that are not easily digested by fish.

**Corresponding Author:**

**Aakriti Regmi**

Rainbow Trout Fishery Research  
Station, Dhunche, Rasuwa,  
Nepal

These anti-nutritional compound have the potential to impede the digestion process<sup>[9]</sup>. In order to mitigate the deleterious effects of anti-nutritional factors and NSP in plant-based diets for fish in aquaculture, the supplementation of exogenous enzymes has been proven to be effective. This approach improves the utilization of nutrients by improving the digestibility of carbohydrates, protein, and minerals. As a result, fish growth and health improve<sup>[10]</sup>. In the present study, we investigated the impact of administering commercial probiotics, as well as probiotic combined with enzymes, on the growth and mortality rate of rainbow trout.

## 2. Materials and Methods

### 2.1 Fish rearing

The research was carried out at the Rainbow Trout Fishery Research Station in Dhunche, Rasuwa. The rainbow trout used for the research were obtained from the same research hatchery. Each raceway used in the study had dimensions of 120 × 75 × 55 cm and was filled with water to a depth of 30 cm. The average stocking size for each treatment is presented in Table (1). To ensure optimal conditions, each raceway was aerated with recirculating freshwater and maintained at a water temperature of 14.5±1.8 °C. The dissolved oxygen level was maintained between 8.2-8.5 mg/L, and the pH level was

kept at 8. The light period followed a schedule of 12 hours of light and 12 hours of darkness (12L: 12D).

A commercially available pellet diet with a protein content of 40%, fulfilling the nutritional requirements of rainbow trout, was used. The pellet diet underwent grinding in a blender to obtain a powdered form, followed by sieving through a 50-mesh sieve to eliminate larger particles. Probiotic supplements, including Dige Pro containing *lactobacillus*, *bifidobacterium* and *streptococcus*, as well as Multizyme containing probiotics and enzymes (xylanase, protease, amylase, lipase), were incorporated into the diets. The feed was meticulously mixed with probiotics and a 2 g gel to ensure proper binding with the pellets. Subsequently, the diets were air-dried for 24 hours. The experiment used a commercially available diet as the control group, while the treatment groups (T<sub>1</sub>, T<sub>2</sub>, and T<sub>3</sub>) received diets with different levels of probiotics and probiotics combined with enzymes for a duration of 90 days. The composition of each diet can be found in Table 1. The fish were fed twice daily, at 09:00 and 16:00 hours, with a feed rate equivalent to 3% of their body weight. All fish were individually weighed on a weekly basis, recording data on fish body weight and feed intake. Mortality rates were recorded on a daily basis. The experiment followed a complete randomized design with three replicates.

**Table 1:** Composition of ingredients in different diets for T<sub>1</sub>, T<sub>2</sub>, T<sub>3</sub>, and T<sub>4</sub>

Ingredients	T <sub>1</sub> (Kg)	T <sub>2</sub> (Kg)	T <sub>3</sub> (Kg)	T <sub>4</sub> (Kg) (Control)
Dried Anchovy	17.75	17.75	17.75	17.75
Soya Chokar	2.75	2.75	2.75	2.75
Wheat	2.75	2.75	2.75	2.75
Rice bran	1.25	1.25	1.25	1.25
Mineral	0.25	0.25	0.25	0.25
Vitamin	0.25	0.25	0.25	0.25
Probiotics with gel	(6 gm Dige-pro + 2 gm Gel)/ kg feed	(6 gm Multizyme + 2 gm Gel)/ kg feed	(3 gm Dige pro + 3 gm Multizyme+ 2 gm Gel)/kg feed	0

**Table 2:** Average stocking sizes (in grams) for different treatment

Treatment	Stocking size (gram)
T <sub>1</sub>	1.423±0.002
T <sub>2</sub>	1.412±0.096
T <sub>3</sub>	1.451±0.136
T <sub>4</sub>	1.477±0.118

### 2.2 Growth performances

During the study, various growth factors were assessed, which included final weight, daily weight gain, specific growth rate (SGR) and survival rate.

### 2.3 Statistical analysis

One-way analysis of variance (ANOVA, R-stat) was used to determine whether significant variation between the treatments existed. Difference between means were determined and compared by LSD test. All tests used a significance level of  $p < 0.05$ . Data are reported as means ± standard errors.

## 3. Results and Discussion

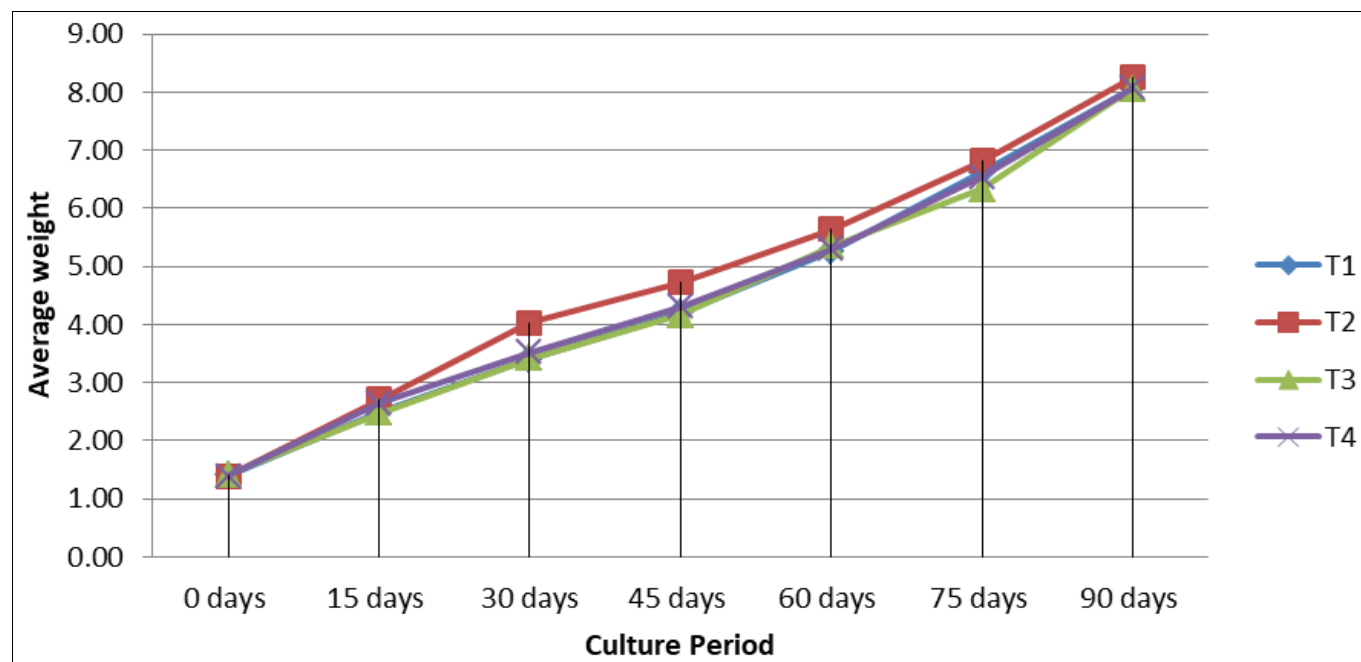
### 3.1 Fish growth performance and Survival rates

During a 90-day period, the impact of dietary probiotics and

probiotic plus enzymes on the growth performance of rainbow trout was examined. Table 3 presents the total stocked weight, total harvested weight, survivability, daily weight gain rate, and specific growth rate of fish among different treatment groups. The results revealed that the final weight, daily weight gain, and specific growth rate were significantly higher in the T<sub>2</sub> treatment group when compared to both the control group and other treatment groups. Additionally, the survival rates were determined as 77.77%, 71.42%, 66.66%, and 71.11% for T<sub>1</sub>, T<sub>2</sub>, T<sub>3</sub>, and the control group, respectively. Similarly, mean harvest weight was significantly higher in T<sub>2</sub> as compared with remaining treatment T<sub>1</sub>, T<sub>2</sub> and T<sub>4</sub> ( $p < 0.05$ ). The highest mean harvested weight was observed in treatment T<sub>2</sub> (8.26±0.07 g) followed by treatment T<sub>1</sub> (8.07±0.06 g) and T<sub>4</sub> (8.07±0.13g) which were not significantly different with each other ( $p > 0.05$ ). Conversely, the lowest mean harvest weight was recorded in treatment T<sub>3</sub> (8.06±0.06 g). The daily growth rate (DGR) was seen highest in T<sub>2</sub> (0.092 g/day) which was significantly different with treatment T<sub>1</sub> (0.090 g/day), T<sub>3</sub> (0.090 g/day) and T<sub>4</sub> (0.090 g/day) ( $p < 0.05$ ). Similarly the highest specific growth rate (SGR) was seen in T<sub>2</sub> (7.62), which is significantly different with the other treatment T<sub>1</sub> (7.41), T<sub>3</sub> (7.38) and T<sub>4</sub> (7.43) ( $p < 0.05$ ).

**Table 3:** Mean Value of growth parameter of rainbow trout during the experimental period of 90 days

Growth and Production Parameter	Unit	Treatment			
		T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>
Total Stocked Number	No	315	315	315	315
Total stocked wt.	(g)	441	437.85	447.30	437.85
Mean Stock wt.	(g/fish)	1.40±0.15	1.39±0.16	1.42±0.12	1.39±0.13
Total harvested wt.	G	1977.5	1857.75	1692.60	1808.43
Mean harvested wt.	g/fish	8.07±0.06 <sup>b</sup>	8.26±0.07 <sup>a</sup>	8.06±0.06 <sup>b</sup>	8.07±0.13 <sup>b</sup>
Daily weight gain	(g/fish/day)	0.090 <sup>b</sup>	0.092 <sup>a</sup>	0.090 <sup>b</sup>	0.090 <sup>b</sup>
Survival rate	(%)	77.77	71.42	66.66	71.11
Specific growth rate	%	7.41 <sup>b</sup>	7.62 <sup>a</sup>	7.38 <sup>b</sup>	7.43 <sup>b</sup>

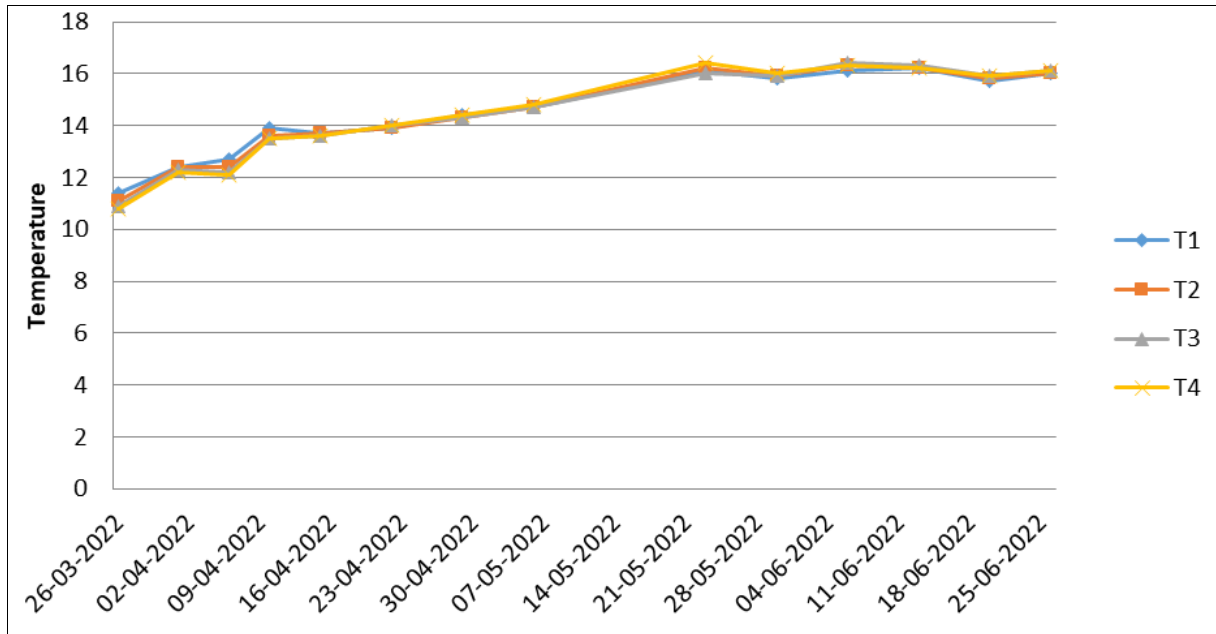
**Fig 1:** Growth parameter of rainbow trout during the experimental period of 90 days

In this study, Rainbow trout were fed a diet enriched with either a single probiotic, a combination of several probiotics, or a probiotic blend comprising enzymes. The current study found that including both probiotics and exogenous enzymes in the food considerably increased the growth rate of T<sub>2</sub>, as shown by increases in specific growth rate and daily weight gain across the culture period. Adeoye *et al.*, (2016) <sup>[11]</sup> and Ghodrati *et al.*, (2021) <sup>[12]</sup> supported these findings by demonstrating similar positive effects of commercial probiotics mixed with enzymes such as phytase, xylase, protease, amylase, cellulase, and lipase on tilapia and Siberian Sturgeon growth. The improved growth performance could be related to the probiotics' ability to create enzymes that aid in fiber digestion. These enzymes collaborate with the fish's own digestive enzymes as well as external enzymes to boost the availability of appropriate chemicals for probiotic effect <sup>[13]</sup>. The presence of non-starch polysaccharide compounds (NSP) in the gut can increase viscosity, causing detrimental effects on digesta viscosity, the mucus layer, and gut morphology. These changes adversely affect the digestion rate and nutrient absorption, leading to impaired function of internal organs and reduced growth performance. The presence of enzymes, on the other hands, serves to mitigate the negative impacts of anti-nutritional factors and NSP, thereby minimizing these adverse effects <sup>[14]</sup>. In addition, these enzymes positively

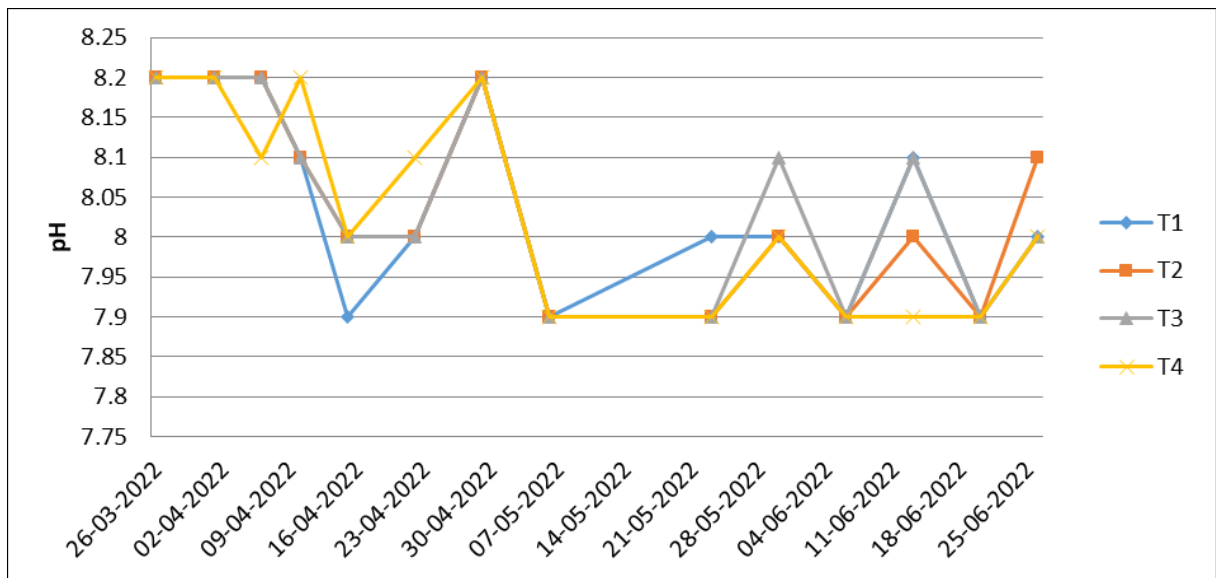
impact the gut flora by enhancing digestibility, allowing for better nutrient absorption and assimilation, resulting in increased fish growth performance <sup>[15]</sup>. Multizyme is a probiotic and enzyme blend that includes enzymes such as amylase, xylanase, phytase, lipase, protease, and cellulase. Enzyme supplementation has the ability to reduce the effects of anti-nutritional components while increasing the uptake of nutrients <sup>[16, 17]</sup>. Considering that aquatic species lack enzymes like xylanase, phytase including xylanase in their diet improves nutrient utilization by breaking down non-starch polysaccharides, resulting in higher feed efficiency <sup>[18, 19]</sup>. Furthermore, the use of phytase in aquafeed might boost nutritional value by hydrolyzing indigestible organic phosphorus into free form. This process supplies necessary nutrients for the growth of intestinal microbiota, enhancing overall gut health and consumption of nutrients in aquatic organisms <sup>[20]</sup>.

### 3.2 Water quality parameter

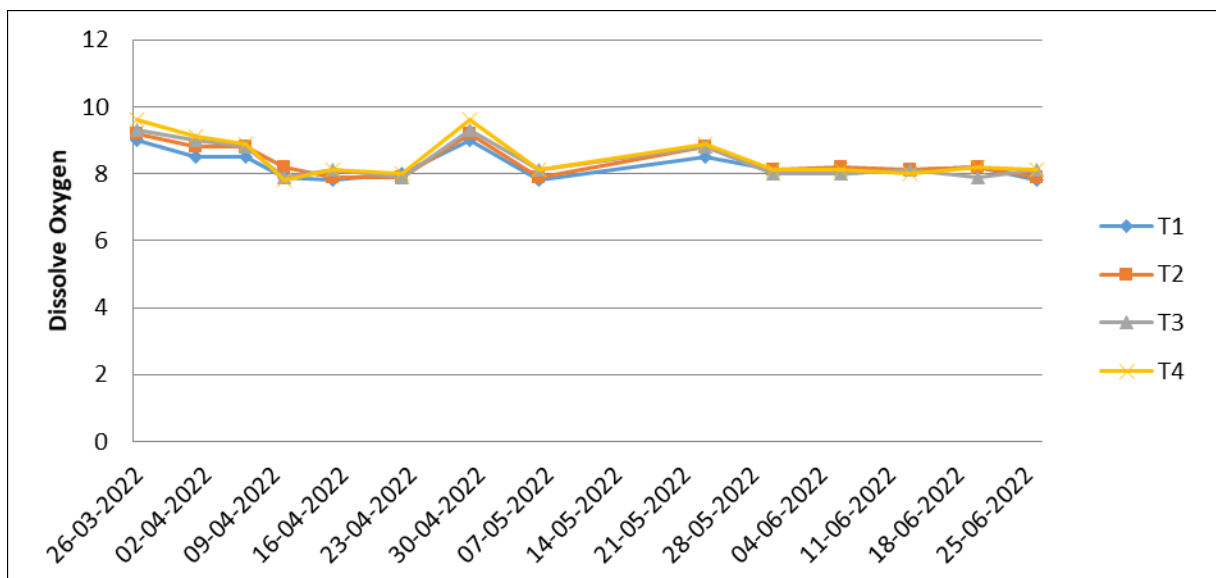
In aquaculture, the health of the fish is significantly impacted by the water quality parameters. During the period of experiment, critical water quality parameters like average temperature, dissolve oxygen and pH are within the acceptable range which are presented in following figures.



**Fig 2:** Water Temperature and its variation during Culture Period.



**Fig 3:** pH and its variation during Culture Period



**Fig 4:** Dissolve Oxygen and its variation during Cultural Period



#### 4. Conclusion

The current study's findings imply that introducing Multizyme, a probiotic and enzyme blend, at a concentration of 6 gm/kg of feed in a regular commercial diet can boost growth rates as well as enhance aquaculture health. It suggests that including probiotics in fish feeding strategies could provide further benefits in aquaculture. More research is needed, however, to fully understand the direct effects of probiotics and the precise processes underlying the reported enhancement in growth performance among the treated fish. Future research should look into hematological parameters, digestive enzyme activity, intestinal anatomy, and the chemical content of the fish.

#### References

1. Abedi SZ, Yeganeh S, Moradian F, Ouraji H. The influence of probiotic (isolated based on phytase activity) on growth performance, body composition, and digestibility of rainbow trout, *Oncorhynchus mykiss*. Journal of the World Aquaculture Society. 2022, Jun 13;53(5):1006-1030. <https://doi.org/10.1111/jwas.12906>
2. Pillinger M, Weber B, Standen B, Schmid MC, Kesselring JC. Multi-strain probiotics show increased protection of intestinal epithelial cells against pathogens in rainbow trout (*Oncorhynchus mykiss*). Aquaculture. 2022 Nov;560:738487. <https://doi.org/10.1016/j.aquaculture.2022.738487>
3. Docando F, Nuñez-Ortiz N, Serra C, Arense P, Enes P, Oliva-Teles A, et al. Mucosal and systemic immune effects of *Bacillus subtilis* in rainbow trout (*Oncorhynchus mykiss*). Fish & Shellfish Immunology. 2022 May;124:142-155. <https://doi.org/10.1016/j.fsi.2022.03.040>
4. Park Y, Lee S, Hong J, Kim D, Moniruzzaman M, Bai SC. Use of probiotics to enhance growth, stimulate immunity and confer disease resistance to *Aeromonas salmonicida* in rainbow trout (*Oncorhynchus mykiss*). Aquaculture Research. 2017;48(6):2672-2682.
5. Rasmussen JA, Villumsen KR, Ernst M, Hansen M, Forberg T, Gopalakrishnan S, et al. A multi-omics approach unravels metagenomic and metabolic alterations of a probiotic and synbiotic additive in rainbow trout (*Oncorhynchus mykiss*). Microbiome. 2022 Jan, 30, 10(1). <https://doi.org/10.1186/s40168-021-01221-8>
6. Sahu MK, Swarnakumar NS, Sivakumar K, Thangaradjou T, Kannan L. Probiotics in aquaculture: importance and future perspectives. Indian Journal of Microbiology. 2008;48(3):299-308. <https://doi.org/10.1007/s12088-008-0024-3>
7. Zhao C, Men X, Dang Y, Zhou Y, Ren Y. Probiotics Mediate Intestinal Microbiome and Microbiota-Derived Metabolites Regulating the Growth and Immunity of Rainbow Trout (*Oncorhynchus mykiss*). Microbiology Spectrum. 2023, 11(2). <https://doi.org/10.1128/spectrum.03980-22>
8. Rohani MF, Islam SM, Hossain MK, Ferdous Z, Siddik MA, Nuruzzaman M, et al. Probiotics, prebiotics and synbiotics improved the functionality of aqua feed: Upgrading growth, reproduction, immunity and disease resistance in fish. Fish & Shellfish Immunology. 2022 Jan;120:569-589. <https://doi.org/10.1016/j.fsi.2021.12.037>
9. Sinha AK, Kumar V, Makkar HP, De Boeck G, Becker K. Non-starch polysaccharides and their role in fish nutrition: A review. Food Chemistry. 2011;127(4):1409-1426. <https://doi.org/10.1016/j.foodchem.2011.02.042>
10. Cheng W, Chiu CS, Guu YK, Tsai ST, Liu CH. Expression of recombinant phytase of *Bacillus subtilis*E20 in *Escherichia coli* HMS 174 and improving the growth performance of white shrimp, *Litopenaeus vannamei*, juveniles by using phytase-pretreated soybean meal-containing diet. Aquaculture Nutrition. 2012 Apr 3;19(2):117-127. <https://doi.org/10.1111/j.1365-2095.2012.00946.x>
11. Adeoye AA, Yomla R, Jaramillo-Torres A, Rodiles A, Merrifield DL, Davies SJ. Combined effects of exogenous enzymes and probiotic on Nile tilapia (*Oreochromis niloticus*) growth, intestinal morphology and microbiome. Aquaculture. 2016 Oct;463:61-70. <https://doi.org/10.1016/j.aquaculture.2016.05.028>
12. Ghodrati M, Rajabi Islami H, Hosseini Shekarabi SP, Shenavar Masouleh A, Shamsaie Mehrgan M. Combined effects of enzymes and probiotics on hemato-biochemical parameters and immunological responses of juvenile Siberian sturgeon (*Acipenser baerii*). Fish & Shellfish Immunology. 2021 May;112:116-124. <https://doi.org/10.1016/j.fsi.2021.03.003>
13. Roy T, Mondal S, Ray AK. Phytase-producing bacteria in the digestive tracts of some freshwater fish. Aquaculture Research. 2009 Feb;40(3):344-353. <https://doi.org/10.1111/j.1365-2109.2008.02100.x>
14. Saputra F, Shiu YL, Chen YC, Puspitasari AW, Danata RH, Liu CH, Hu SY. Dietary supplementation with xylanase-expressing *B. amyloliquefaciens* R8 improves growth performance and enhances immunity against *Aeromonas hydrophila* in Nile tilapia (*Oreochromis niloticus*). Fish & Shellfish Immunology. 2016;58:397-405. <https://doi.org/10.1016/j.fsi.2016.09.046>
15. Bedford M, Cowieson A. Exogenous enzymes and their effects on intestinal microbiology. Animal Feed Science and Technology. 2012 April;173(1-2):76-85. <https://doi.org/10.1016/j.anifeedsci.2011.12.018>
16. Farhangi M, Carter CG. Effect of enzyme supplementation to dehulled lupin-based diets on growth, feed efficiency, nutrient digestibility and carcass composition of rainbow trout, *Oncorhynchus mykiss* (Walbaum). Aquaculture Research. 2007 Sept;38(12):1274-1282. <https://doi.org/10.1111/j.1365-2109.2007.01789.x>
17. Lin S, Mai K, Tan B. Effects of exogenous enzyme supplementation in diets on growth and feed utilization in tilapia, *Oreochromis niloticus* x *O. aureus*. Aquaculture Research. 2007 Nov;38(15):1645-1653. <https://doi.org/10.1111/j.1365-2109.2007.01825.x>
18. Jiang TT, Feng L, Liu Y, Jiang WD, Jiang J, Li SH, et al. Effects of exogenous xylanase supplementation in plant protein-enriched diets on growth performance, intestinal enzyme activities and microflora of juvenile Jian carp (*Cyprinus carpio*var. Jian). Aquaculture Nutrition. 2014 May 13;20(6):632-645. <https://doi.org/10.1111/anu.12125>
19. El-Ashry M. Effect of Dietary Xylanase on Growth Performance, Digestive Enzymes and Physiological Responses of Nile Tilapia, *Oreochromis niloticus* Fingerlings Fed Plant-Based Diets. Annals of Agricultural Science, Moshtohor. 2021;59(2):71-80. <https://doi.org/10.21608/assjm.2021.183649>
20. Heyer CME, Schmucker S, Burbach K, Weiss E, Eklund M, Aumiller T, et al. Phytate degradation, intestinal microbiota, microbial metabolites and immune values are changed in growing pigs fed diets with varying calcium-phosphorus concentrations and fermentable substrates. Journal of Animal Physiology and Animal Nutrition. 2019 April. <https://doi.org/10.1111/jpn.13088>