



# International Journal of Fisheries and Aquatic Studies

E-ISSN: 2347-5129

P-ISSN: 2394-0506

(ICV-Poland) Impact Value: 5.62

(GIF) Impact Factor: 0.549

IJFAS 2022; 10(2): 125-132

© 2022 IJFAS

www.fisheriesjournal.com

Received: 15-01-2022

Accepted: 18-02-2022

## Akpan II

Department of Zoology, Akwa  
Ibom State University, Ikot  
Akpaden, Akwa Ibom State,  
Nigeria

## Abiaobo NO

Department of Zoology, Akwa  
Ibom State University, Ikot  
Akpaden, Akwa Ibom State,  
Nigeria

## George UU

Department of Fisheries and  
Aquaculture, Akwa Ibom State  
University, Obio Akpa Campus,  
Akwa Ibom State, Nigeria

## George IE

Department of Zoology, Akwa  
Ibom State University, Ikot  
Akpaden, Akwa Ibom State,  
Nigeria

## Composite baseline survey of ichthyofaunal assemblage in a segment of lower Imo river estuary

Akpan II, Abiaobo NO, George UU and George IE

### Abstract

A composite baseline survey of the Ichthyofaunal assemblage in a segment of the lower Imo River Estuary was carried out from March 2020 - February, 2021 at three sampling stations: Esene Creek (SI), Harbour Area (SII) and Uta Ewa Creek (SIII). Fish samples were collected from the catch of the artisanal and subsistence fishermen operating in the area. The assemblage revealed a rich and varied species, which were grouped into 17 families of 31 species. The contribution from the stations followed an increasing order from stations I to III: 324(8.20%), 350 (12.9%) and 2588 (79.33%). Station III had a higher density and varied significantly when compared to station II and I, respectively. Biotic statistics indices showed polydiversal conditions among the species. Results of the survey revealed that fish assemblage from the study area varied from species to species and from family to family, which implies that the fishery and fishery products of the area can contribute significantly in boosting foreign exchange if properly managed.

**Keywords:** Composite, ichthyofaunal, estuary, assemblage, lower Imo river

### 1. Introduction

The knowledge of Biodiversity is increasing on a daily basis, this is prompted by the frequent disturbances of the natural ecosystem (Jamu *et al.*, 2003 and Guo *et al.*, 2018) <sup>[29, 23]</sup>, which lead to the rapid extinction of some species of flora-fauna origin, which fish is inclusive. This menace is as a result of anthropogenic perturbations, which enhances the alterations of the natural ecosystem (Arthington *et al.*, 2016 and Fu *et al.*, 2003) <sup>[10, 20]</sup>. Diversity is a fundamental property of every living system, which is manifested at every level of the hierarchy, from molecules to ecosystems. Fish exhibits the greatest biodiversity amongst vertebrates, with over 22,000 species (Cosgrove and Rijsberman, 2000) <sup>[12]</sup>. The high diversity of fish is probably the main source of stability to various tropical fisheries and also gives a strong reason for its conservation (Onuoha *et al.*, 2010) <sup>[38]</sup>. Naturally, fauna assemblage, especially that of fish is a very valuable resource from the eco-economic viewpoint and as such its management and conservation is of great importance. Nowadays, fish diversity and its corresponding habitats management is a vital flash point and the ability to evaluate the effects of habitat distortion and several other impacts on fish population dynamics need a comprehensive survey of the fish population before and after change assessment (Dudgeon *et al.*, 2006) <sup>[15]</sup>. This could be achieved through a critical survey of its composition (King, 1992 and Essien-Ibok, 2020) <sup>[30, 18]</sup>.

Imo River Estuary is an extensive coastal area, which encompasses the Niger Delta Region, of Nigeria and is also noted for its great potential in fish production (Orji and Onyejiaka, 1990) <sup>[39]</sup>. Aside from agricultural practices, fishing forms the economic basis of the people. At the lower estuary of the river, there are sketchy fishing settlements, which is situated within an effluent discharge zone of some industries. Transportation, dredging, agricultural practices etc. are the major activities going on within the river. There is a shortage of information on a composite baseline survey of Ichthyofaunal assemblage in a segment of Lower Imo River Estuary. This study aims at surveying the Ichthyofaunal assemblage in a segment of the Lower Imo River estuary.

### 2. Materials and Methods

#### 2.1 Study Area

The study was conducted in a segment of Lower Imo River estuary (Figure 1), with three sampling sites at Esene Creek, Harbour Area and Uta Ewa Creek all located in Ikot Abasi,

### Corresponding Author:

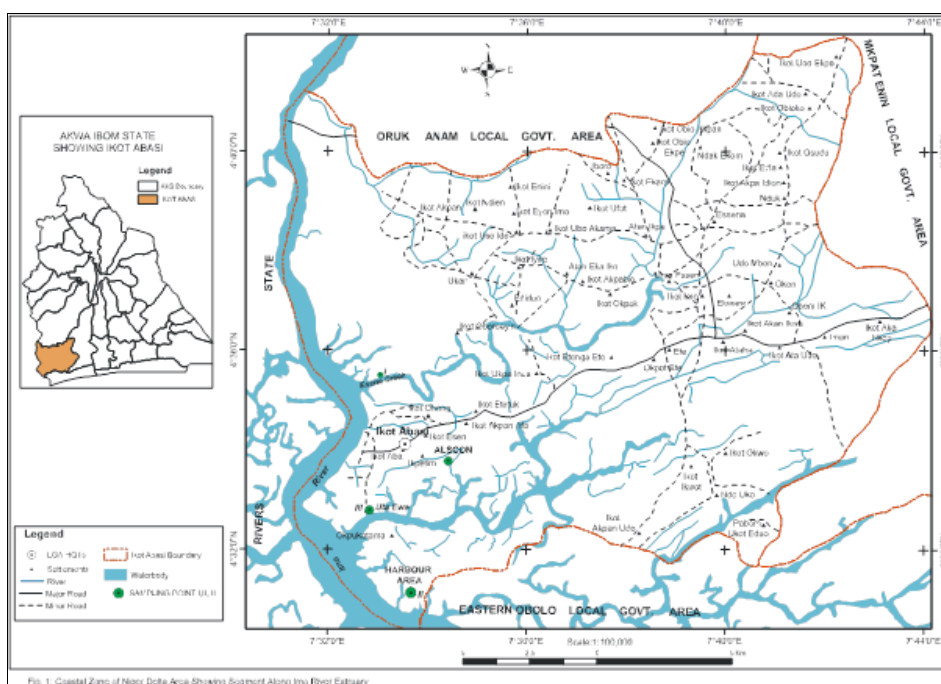
#### Akpan II

Department of Zoology, Akwa  
Ibom State University, Ikot  
Akpaden, Akwa Ibom State,  
Nigeria

**Niger Delta Region, Nigeria.**

It lies in the Lower Imo River Basin and extends from latitudes 5.0°00'N – 6.0°00'N and longitudes 6.0°00'E – 7.0°00'E. The river has a main channel total length of about 240km<sup>2</sup> between its source and where it discharges into the Atlantic Ocean. The source is dendritic and flows through

coastal plain alluvium and mangrove swamp and empties into the Atlantic Ocean at the Bight of Bonny, forming a natural boundary between Akwa Ibom and River State. A detailed description of the study area is contained in (Akoma, 2008, Akpan, 2006, Petter *et al.*, 1994)<sup>[5, 7, 40]</sup>.



**Fig 1:** Map of the study area showing sampling locations

**2.2 Sample Collection**

Fish samples were collected for a period of one year (March, 2020 - February, 2021) from commercial landings of artisanal and subsistence fisheries landings. Sampling was done twice a month. Fish samples were selected and purchased from the fisherfolks. The samples purchased were preserved in an ice-chest and transported to the laboratory for sorting and identification. Oral interviews were conducted with f fisherfolks and inhabitants of the area to gain insight into the fisheries of the area. The samples were identified to the species level with the aid of identification keys as provided by Edwards *et al.*, (2001)<sup>[16]</sup>, Fisher *et al.*, (1981)<sup>[19]</sup>, Schneider (1990)<sup>[41]</sup>, Idodo Umeh (2003)<sup>[28]</sup> and Holden and Reed, (1972)<sup>[26]</sup>. Samples were weighed to the nearest 0.1g using a spring balance and total length using a meter rule.

**2.3 Statistical Analysis**

Basic simple statistics of range, length, weight, standard deviations and percentage composition were computed. The index of preponderance (%IP) was computed as follows:

$$IP = \frac{\% N \% W \times 10}{\sum (\% N \% Wt)}$$

Where, N = No. of fish species, Wt = Weight of species. Fishes with IP valued less than 0.5 were regarded as being a relatively insignificant contribution, while those with IP values greater than 0.5 were regarded as being of significant contribution. Analysis of variance was done with the aid of a statistical computer package (SPSS, version 20.0).

**3. Result**

The Ichthyofaunal composition in a segment of the Lower

Imo River estuary is characterized by a rich and varied fish species assemblage. Table 1-3 indicates the species composition, size ranges, abundance and index of the preponderance of fish species collected at the sampling stations. Tables 4-6, showed an analysis of the variance of the species collected on a station, monthly and seasonal basis. Species richness of the collected samples on the family basis is represented in figures 2-4. A total of 3262 fish species weighing 4,057.8kg belonging to 17 families of 31 species were collected during the survey period.

The families comprised one or more species which differ in number from each other. The morphometric characters of the collected species revealed differences in length and body weight which is natural. Station III recorded the highest number of fish species totaling 2588 (79.33%), while station I had the least number of 324 (9.93%). *Liza grandisquamis* was the most abundant species as it recorded a total of 267 individual species in all the sampling stations.

From the index of preponderance, the most frequently occurring species in station III were *chrysichthys auratus* (71%), *Pseudotolithus elongatus* (0.40%) and *Chrysichthys nigrodigitatus* (1.10%). Station II recorded *Chrysichthys nigrodigitatus* (1.53%) *Pseudotolithus typus* (1.46%) and *Pseudotolithus elongatus* (0.44%) as the most frequently occurring species. *Chrysichthys auratus* (1.00%), *Chrysichthys nigrodigitatus* (0.64%) and *Pseudotolithus elongatus* (0.65%) were recorded as the most frequently occurring species in station 1. The least occurring fish species recorded was *Eleotris vitata* (0.001%) for station III, *Ethmalosa fimbriata* (0.001%) for station II and *Eleotris vitata* (0.02%) for the station I.

Analysis of variance of fish density at different stations, showed a higher density ( $r = 230^{0.08} P < 0.05$ ) at station III

and this varied significantly with the density at station I and station II respectively. Analysis of variance of the fish density on a monthly basis was high in the month of June ( $r = 152.00^a, p < 0.05$ ). The analysis of fish density on a seasonal

basis showed that the density was high at the wet season ( $r = 308.9^a, P < 0.05$ ), but significantly different in the dry season ( $r = 74.3^b, P < 0.05$ ).

**Table 1:** Species Composition, Size Range, Abundance and Index of Preponderance of Species in station 1 during the Period of Survey (March, 2020 to February, 2021).

Family/Specie	N	TL (cm) Min-Max	± SD	TW (g) Min-Max	± SD	% IP	% N	% W
Carangidae								
<i>Caranx hippos</i>	32	8.08-18.21	11.29 ± 1.50	17.28-58.18	29.40 ± 6.30	0.24	0.24	9.90
<i>Trachinotus teraia</i>	08	16.10-17.10	16.60 ± 0.50	195.40-211.40	203.40 ± 8.00	0.41	2.50	16.62
<i>Trachinotus goreensis</i>	20	4.75-8.05	6.02 v 0.70	6.18-9.45	8.02 ± 0.60	0.04	6.17	0.70
Eleotridae								
<i>Eleotris senegalensis</i>	13	23.18-25.10	24.14 ± 0.96	31.81-34.18	32.99 ± 1.20	0.11	4.01	2.70
<i>Eleotris vittata</i>	12	6.01-8.01	7.01 ± 0.54	5.41-6.71	6.06 ± 0.65	0.02	3.70	0.50
Gobiidae								
<i>Bathygobius soporator</i>	13	23.18-25.10	24.14 ± 0.96	31.81-34.18	32.99 ± 1.20	0.11	4.01	2.70
Clupeidae								
<i>Ilisha Africana</i>	29	10.39-21.38	16.9 ± 1.26	24.17 -47.38	32.60 ± 2.37	0.24	8.95	2.70
<i>Odoxothrissa mento</i>	04	7.28-8.00	7.64 ± 0.36	9.25-10.18	9.72 ± 0.47	0.010	1.23	0.80
<i>Ethmalosa fimbriata</i>	16	7.12-10.63	9.53 ± 0.54	14.18-16.10	14.60 ± 0.32	0.10	5.00	1.20
Mugilidae								
<i>Liza grandisquamis</i>	22	15.10-22.17	19.25 ± 0.95	34.02-48.18	41.93 ± 2.09	0.23	6.80	3.43
<i>Mugil cephalus</i>								
Trichiuridae								
<i>Trichiurus lepturus</i>	07	11.81-13.28	12.55 ± 0.51	39.51-48.20	44.67 ± 1.53	0.10	2.80	8.50
Malapteruridae								
<i>Malapterurus electricus</i>	09	8.71-11.71	10.57 ± 0.51	39.51-48.20	44.67 ± 1.53	0.10	2.80	3.65
Bothidae								
<i>Citharichthys stampfi</i>	08	09.01-10.01	9.43 ± 0.56	23.03-25.01	24.54 ± 0.52	0.10	2.50	2.01
Sciaenidae								
<i>Pseudolithus senegalensis</i>	28	9.10-12.62	10.52 ± 0.42	25.38-36.20	31.03 ± 1.35	0.2	8.64	2.53
<i>Pseudolithus elongatus</i>	39	11.51-23.10	17.03 ± 1.20	45.02 -87.70	66.18 ± 4.56	0.65	12.04	5.41
<i>Pseudolithus typhus</i>	13	18.16-22.01	20.50 ± 1.19	83.18-98.15	89.98 ± 4.41	0.30	4.0	7.40
Claroteidae								
<i>Chrysichthys auratus</i>	23	15.18-18.19	16.55 ± 0.31	125.30-218.20	173.50 ± 9.74	1.00	7.10	14.20
<i>Bagrus docmack</i>	08	06.18-8.32	7.10 ± 0.18	36.23-40.28	38.23 ± 0.18	0.08	2.50	3.12
<i>Chrysichthys nigrodigitatus</i>	13	28.17-39.81	35.23 ± 2.18	194.20 -200.12	197.17 ± 1.11	0.64	4.00	16.10
Cichlidae								
<i>Tilapia zilli</i>								
Σ	342						100.25	100.2

Source: Field Study, 2021

**Table 2:** Species Composition, Size Range, Abundance and Index of Preponderance of Species in station 2 during the Period of Survey (March, 2020 to February, 2021).

Family/Specie	N	TL (cm) Min-Max	± SD	TW (g) Min-Max	± SD	% IP	% N	% W
Carangidae								
<i>Caranx hippos</i>	22	9.01-10.13	9.59 ± 0.29	19.18-23.48	22.31 ± 0.05	0.12	6.29	1.94
<i>Trachinotus teraia</i>	4	0.00-8.28	8.28 ± 0.00	12.23-12.27	12.25 ± 0.54	0.01	1.14	1.07
<i>Trachinotus goreensis</i>	14	5.81-8.04	6.93 ± 0.53	7.10 -8.08	7.46 v 0.22	0.03	4.00	0.65
Eleotridae								
<i>Eleotris senegalensis</i>	15	13.18-17.05	14.91 ± 1.01	35.38-40.35	38.51 ± 1.18	0.14	4.29	3.36
<i>Eleotris vittata</i>	12	6.01-8.01	7.01 ± 1.01	4.31-5.38	4.85 ± 0.54	0.01	3.43	0.42
Clupeidae								
<i>Ilisha Africana</i>	27	12.40-24.28	20.01 ± 1.10	25.18-130.10	39.93 ± 10.92	0.27	7.71	3.50
<i>Odoxothrissa mento</i>	3	0.00-6.00	6.00 ± 0.00	7.18-8.18	8.08 ± 1.02	0.006	0.90	0.70
<i>Pelloni leonensis</i>	32	9.10-11.72	10.52 ± 0.27	13.18 -19.20	16.06 ± 0.80	0.13	9.14	1.40
<i>Ethmalosa fimbriata</i>	2	0.00-5.18	5.18 ± 0.00	2.16-2.18	2.17 ± 1.00	0.001	0.57	0.19
Mugilidae								
<i>Liza grandisquamis</i>	34	18.18-25.11	21.86 ± 0.82	40.30-50.18	46.89 ± 1.14	0.40	9.71	4.10
<i>Mugil cephalus</i>	14	14.81-16.71	16.01 ± 0.60	49.18-58.18	54.89 ± 2.90	0.19	4.00	4.78
Ariidae								
<i>Arius gigas</i>	7	10.00-22.14	18.33 ± 2.80	37.20-80.18	59.21 ± 8.80	0.10	2	5.20
Cynoglossidae								
<i>Cynoglossus senegalensis</i>	8	19.18-26.01	22.79 ± 1.98	98.10-191.10	129.10 ± 1.00	0.25	2.29	11.25
Sciaenidae								
<i>Pseudolithus senegalensis</i>	18	11.08-20.19	13.31 ± 1.40	30.18-88.70	41.60 ± 9.51	0.18	5.14	3.62

<i>Pseudolithus elongatus</i>	25	17.13-28.10	21.02 ± 1.52	48.70-80.18	71.65 ± 4.93	0.44	7.14	6.24
<i>Pseudolithus typhus</i>	19	20.34-23.01	21.29 ± 0.60	93.18-100.31	97.51 ± 1.51	0.46	5.43	8.50
<b>Claroteidae</b>								
<i>Chrysichthys auratus</i>	31	17.18-20.18	18.55 ± 0.31	145.30-238.20	193.50 ± 9.74	1.50	8.86	16.90
<i>Bagrus docmack</i>	27	08.00-10.78	09.18 ± 0.96	48.20-53.32	50.32 ± 0.56	1.50	8.86	16.90
<i>Chrysichthys nigrodigitatus</i>	29	38.18-50.38	44.67 ± 1.80	232.10-328.10	212.68 13.81	1.53	8.29	18.53
<b>Cichlidae</b>								
<i>Tilapia zilli</i>	7	18.10-20.30	19.20 ± 1.10	31.10-40.34	35.72 ± 4.62	0.06	2.0	3.11
<b>Σ</b>	<b>350</b>						<b>100.04</b>	<b>100.11</b>

Source: Field Study, 2021

**Table 3:** Species Composition, Size Range, Abundance and Index of Preponderance of Species in station 3 during the Period of Survey (March, 2020 to February, 2021).

Family/Species	N	TL (cm) Min-Max	± SD	TW (g) Min-Max	± SD	% IP	% N	% W
<b>Carangidae</b>								
<i>Caranx hippos</i>	107	7.21-10.71	9.83 ± 0.3	18.19 -26.50	24.02 ± 0.30	0.06	4.13	1.43
<i>Trachinotus teraia</i>	93	7.90-11.21	9.30 ± 0.33	14.17-18.58	16.58 ± 0.58	0.04	4.00	0.98
<i>Trachinotus goreensis</i>	86	7.04-9.98	8.20 ± 0.32	6.42-9.08	7.15 ± 0.29	0.01	3.32	0.42
<b>Malapteruridae</b>								
<i>Malapterurus electricus</i>	68	0.00-13.20	13.20 ± 0.00	0.00-52.00	52.00 ± 0.00	0.08	2.63	3.08
<b>Eleotridae</b>								2.32
<i>Eleotris senegalensis</i>	90	14.18-17.25	15.81 ± 0.33	4.20 -40.64	34.95 ± 4.44	0.07	3.50	2.07
<i>Eleotris vittata</i>	10	8.9-9.10	8.96 ± 0.05	4.18-4.38	4.28 ± 0.10	0.001	0.40	2.05
<b>Gobiidae</b>								2.05
<i>Bathygobius soporator</i>	45	14.80-29.10	22.48 ± 3.12	31.38-38.10	34.51 ± 1.23	0.04	1.74	2.05
<b>Mochokidae</b>								7.73
<i>Synodontis omias</i>	10	4.60-48.10	26.35 ± 21.75	5.28-25.30	130.29 ± 125.01	0.03	0.40	7.73
<b>Schilbedae</b>								
<i>Schilbe mytus</i>	0.8	0.00-3.20	3.20 ± 0.00	0.00-5.31	5.31 ± 0.00	0.009	0.31	0.32
<b>Clupeidae</b>								4.79
<i>Illisha Africana</i>	198	18.18-31.13	23.81 ± 1.18	28.15-59.30	36.51 ± 3.16	0.2	7.65	2.17
<i>Odoxothrissa mento</i>	74	1.79-15.10	6.09 ± 1.72	10.18 -48.10	19.04 ± 4.72	0.03	2.90	1.13
<i>Pellonua leonensis</i>	46	4.41-6.81	5.50 ± 0.46	1.85-13.01	10.28 ± 2.16	0.08	2.32	3.44
<i>Ethmalosa fimbriata</i>	134	3.40-12.40	10.33 ± 0.83	13.29-20.58	16.05 ± 0.84	0.05	5.20	0.95
<b>Mugilidae</b>								6.57
<i>Liza grandisquamis</i>	211	18.91-30.20	24.15 ± 1.06	41.28-65.81	52.79 ± 2.06	0.3	8.20	3.13
<i>Mugil cephalus</i>	60	8.46-18.30	15.60 ± 1.48	49.81 - 64.28	58.04 ± 2.16	0.08	2.32	3.44
<b>Ariidae</b>								6.61
<i>Arius gigas</i>	05	0.00-18.78	18.78 ± 0.00	0.00-48.28	48.28 ± 0.00	0.006	0.20	2.86
<i>Arius heudeloti</i>	116	17.52-22.81	20.90 ± 0.65	45.82-70.18	63.24 ± 2.25	0.2	4.50	3.75
<b>Cynoglossidae</b>								5.80
<i>Cynoglossus senegalensis</i>	86	18.10-27.15	24.70 ± 1.15	78.28 -105.10	98.68 ± 3.55	0.02	3.32	5.86
<b>Trichiuridae</b>								
<i>Trichiurus lepturus</i>	71	10.98-16.18	14.30 ± 0.67	48.10-67.31	54.27	0.09	3.00	3.22
<b>Bothidae</b>								
<i>Citharichthys stampfi</i>	67	11.30-13.10	12.47 ± 0.30	19.05-38.05	26.54 ± 2.84	0.05	3.00	1.57
<b>Lutjanidae</b>								
<i>Lutjanus goreensis</i>	87	9.10-25.10	11.94 ± 1.89	17.38-31.80	20.34 ± 1.70	0.04	3.40	1.21
<b>Haemulidae</b>								
<i>Pomadasys jubelini</i>	36	8.18-12.01	10.34 ± 0.70	19.41-33.18	26.78 ± 2.65	00.02	1.40	1.59
<b>Sciaenidae</b>								
<i>Pseudolithus senegalensis</i>	126	9.20-15.50	12.93 ± 0.70	30.18-40.60	34.23 ± 1.15	0.10	4.90	2.03
<i>Pseudolithus elongatus</i>	178	17.21-98.12	31.00 ± 8.51	83.18-94.70	87.76 ± 1.38	0.40	6.90	5.21
<b>Claroteidae</b>								
<i>Chrysichthys auratus</i>	147	19.20-24.18	21.55 ± 0.48	158.15-252.10	208.70 ± 10.19	0.71	5.70	12.38
<i>Bagrus docmack</i>	74	14.18-15.38	14.54 ± 6.23	49.21-65.28	61.60 ± 2.50	0.11	2.90	3.65
<i>Chrysichthys nigrodigitatus</i>	173	25.10-53.50	48.31 ± 2.14	52.18-352.10	269.75	1.10	6.70	15.99
<b>Cichlidae</b>								
<i>Tilapia zilli</i>	13	22.00-26.40	24.20 ± 2.20	34.21-42.34	38.28 ± 4.07	0.01	0.50	2.27
<i>Tilapia goreensis</i>	15	0.00-12.10	12.10 ± 0.00	0.00-17.01	17.00 ± 0.00	0.006	0.60	1.01
<i>Tilapia mariae</i>	10	0.00-21.10	21.00 ± 0.00	0.00-28.20	28.20 ± 0.00	0.007	0.40	1.67
<b>Σ</b>	<b>2588</b>				<b>1685.99</b>		<b>101.52</b>	<b>97.92</b>

**Table 4:** Analysis of Variance for Fish Density at various Stations in Lower Imo River Estuary

Stations	Fish Density
III	230.08 <sup>a</sup>
I	32.33 <sup>b</sup>
II	34.38 <sup>b</sup>
LSD	52.54
P	0.05

\* Means with the same subscripts are not significantly different.

Source: Field Study, 2021.

**Table 5:** Analysis of Variance for Fish Density at Lower Imo River Estuary on a Monthly Basis

Months	$\bar{X}$
January	39.67 <sup>b</sup>
February	45.00 <sup>b</sup>
March	40.67 <sup>b</sup>
April	125.67 <sup>ab</sup>
May	117.67 <sup>ab</sup>
June	152.00 <sup>a</sup>
July	141.33 <sup>ab</sup>
August	123.33 <sup>ab</sup>
September	124.33 <sup>ab</sup>
October	104.33 <sup>ab</sup>
November	118.00 <sup>ab</sup>
December	57.00 <sup>ab</sup>
LSD	105.08
P	0.05

\*Means with the same subscripts are not significantly different.

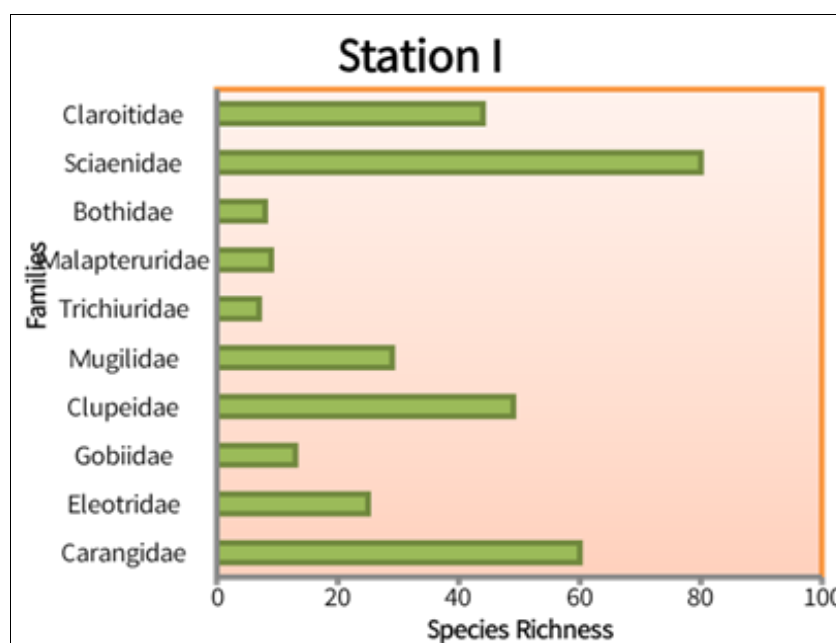
Source: Field Study, 2021.

**Table 6:** Analysis of Variance for Fish Density at Lower Imo River Estuary on Seasonal Basis

Season	x
Wet	308.9 <sup>a</sup>
Dry	74.3 <sup>b</sup>
LSD	604.77
P	0.05

\*Means with the same subscripts are not significantly different.

Source: Field Study, 2021.



**Fig 2:** Species Richness for Fish Families is Station I of Imo River Estuary

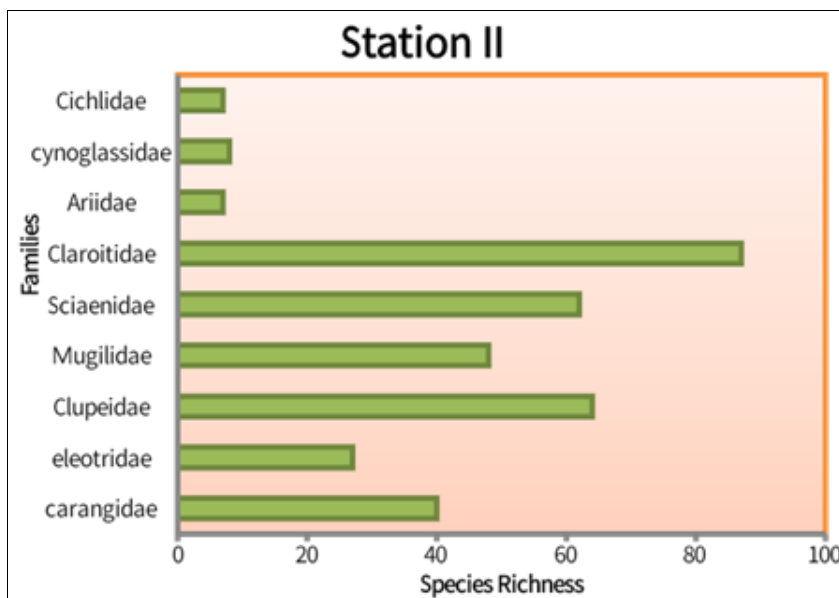


Fig 3: Species Richness for Fish Families is Station II of Imo River Estuary

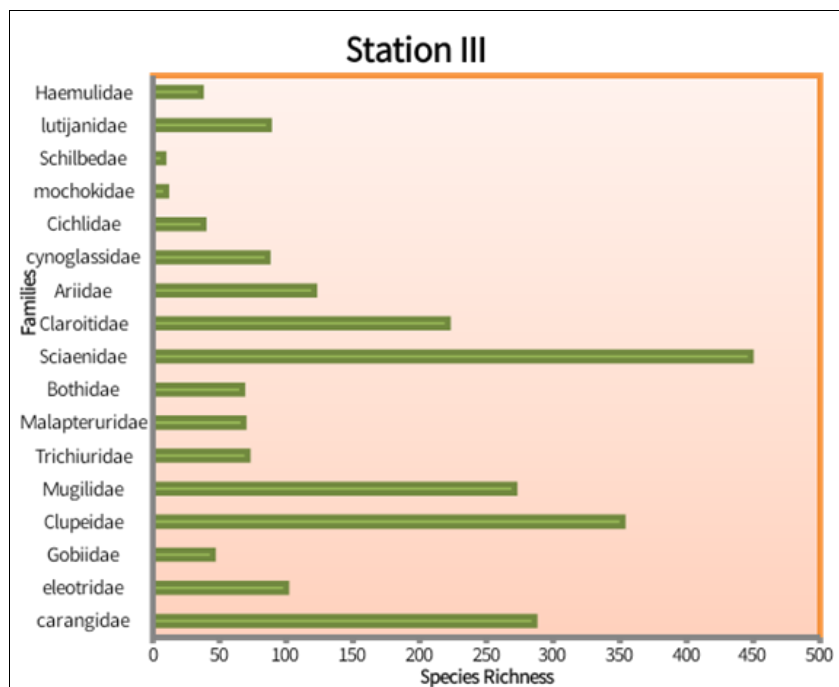


Fig 4: Species Richness for Fish Families is Station III of Imo River Estuary

**4. Discussion**

The composite baseline survey of Ichthyofaunal assemblage in a segment of Lower Imo River estuary, Niger Delta Region, Nigeria revealed an assemblage comprising of 17 families. The species distribution was found to be determined by the natural settings of the system coupled with other factors (Nwadiaro, 1989, Gaines *et al.*, 2009, Udoidiong and King, 2010 and David *et al.*, 2016) [36, 21, 42, 13]. The presence of a variety of dietary items and increased habitats could be linked to species richness in the system (Nwadiaro, 1989, Gaines *et al.*, 2009, Negi, *et al.*, 2013 and David *et al.*, 2016) [36, 21, 35, 13]. Also, its linkage to the Atlantic Ocean, may also account for its species richness and also the presence of some intrusive species (Essen, 1990) [17]. The fish species abundance in this study area could be linked to increased habitats availability and also due to the presence of various dietary materials. This agrees with the report of (Nwadiaro, 1989, Akpan, 2006 and Negi, *et al.*, 2013) [36, 7, 35] which the

opined that fish species were distributed by order and increased with stream size. Adaka *et al.*, 2010; 2014 and Bonjoru *et al.*, 2016 attributed the abundance of fish species with river order and ability to utilize various sources of energy [2, 3, 11].

The Lower Imo River estuary was observed to be housed by three types of fish species. Freshwater species, brackish water species and marine water species; Vakily (1992) [43] observed that some freshwater fishes moved into the estuarine water (brackish). This includes *chrysichthys* species. These fishes exhibited tolerance of the brackish water and are probably euryhaline in nature. The typical brackish water fishes at the estuary included *Ethmalosa fimbriata* and *Illisha Africana*. Welcome (1985; 1986) [44, 45], reported that the species living in the estuary comes from different sources, which include; the freshwater stenohaline species which enter the zone during the flood, marine stenohaline species which live within the zone permanently. It listed the euryhaline fishes of marine

origin to include the families; Clupeidae, Mugilidae, Lutjanidae, Sciaenidae, Arridae and Caranidae. All these families were found in the Lower Imo River estuary.

The preponderance of some families in terms of species diversity, number and weight could be explained by the presence of high food resources (Komolafe and Arawomo, 2008) [33], prolific breeding capabilities and strong adaptation to the prevailing environmental conditions in the aquatic system (Onuoha *et al.*, 2010) [38]. Dike and Agunwamba (2012) [14], reported that estuaries are highly productive zones which benefits fisheries by providing nursery grounds and also open large feeding groups to adult fishes. This assertion might account for the numerous species recorded during the study. The low species diversity as observed in some species might be linked to their low rate of breeding, fluctuations in the environmental conditions which could be less suitable for the species, migration and over-exploitation of the species (Adite *et al.*, 2006) [4]. Seasonal variation occurred in the number of specimens collected, with more specimens during the rainy season than in the dry season. The rainy season is considered to be the feeding period when fish utilize the expanded feeding areas, due to the inundation as a result of the surface-run offs. Also, more allochthonous food materials and nutrients are brought into the habitat.

King (1989) [31] found a positive correlation which corroborated with the assertion that feeding intensity increase with food availability. It has been reported that many tropical freshwater fishes have a broader trophic spectrum during the rains, high species preponderance/dominance during the rainy season (Welcomme, 1985, Akpan, 2006) [44, 7]. There were high species richness and diversity. As the stream order increases, diversity also increases. This is attributed to increase living space leading to an increased number of microhabitats (Udoion and King, 2010) [42].

The commercial species of importance in Lower Imo River include; *Chrysichthys auratus*, *Chrysichthys nigodigitatus*, *Bagrus docmack*. The fishery of the area could be said to be based on the family Bagridae and Sciaenidae. Fish species composition of the Lower Imo River estuary compares with that of other Nigerian aquatic system, (Abowei *et al.*, 2008, Allison and Okadi, 2009 and Akpan, 2013) [1, 9, 8].

The aquatic system is endowed with a rich aquatic bio-faunal assemblage of fish species, which varied from species to species, and families to families, which could boost foreign exchange and contribute to subsistence levels in terms of fishery and fishery products, if properly managed.

## 5. Conclusion

Presently, Lower Imo River Estuary is made up of major fisheries which sustain the livelihoods of its dependents, and as such, is a viable source of the fishery. The rapidly increasing population and other human factors, as was observed during the study, are potential negative impacts on the ecosystem, which will invariably affect the fishery. Therefore, a management plan must be established, which should be fully strategized with a view of preventing a collapse of the fishing and its attendant socio-economic benefits, especially owing to the fact that the fisheries operate basically on free and open-access. The results recorded in this research is an eye-opener to the need of further studies on this ecosystem and similar ecosystems.

## 6. References

1. Abowei JFN, Tawari CC, Hart AI, Garicks DU. Finfish

- species composition, abundance and distribution in the Lower Sombre River, Niger Delta, Nigeria. *Int. J Trop. Agric. Food System*. 2008;2(1):46-53.
2. Adaka GS, Nlewadim AA, Nwaka D, Anyanwu CN, Osuigwe DI. Evaluation of Ichthyofauna Composition of Oramiri-ukwa River Imo State. *Animal Production Research Advances*. 2010;6:286-290.
  3. Adaka GS, Udoh JP, Onyeukwu DC. Freshwater Fish Diversity of a Tropical Rainforest River in Southeast Nigeria. *Advances in Life Science and Technology*. 2014;24:16-23.
  4. Adite A, Winemiller KO, Fiogbe ED. Population Structure and Reproduction of the African Bony Tongue Silver Catfish in the So River-Floodplain System (West Africa): Implications for Management, Ecology of freshwater Fishes. 2006;15:30-39.
  5. Akoma OC. Phytoplankton and nutrient dynamics of a tropical estuarine system, Imo River Estuary, Nigeria. *Int. Multi-Disciplinary Journal*. 2008;2(2):253-258.
  6. Akoma OC, Osundu C. Phytoplankton and nutrient Dynamics of a tropical Estuarine system, Imo River Estuary, Nig. Afri. Res. 2008;2:253-264.
  7. Akpan II. Limnological study of Jaja Creek with special emphasis on species composition, trophic biology and heavy metals composition of some selected species. M.Sc. Thesis, Department of Zoology, University of Uyo, Uyo, 2006, 105-108.
  8. Akpan II. Species composition and abundance in Uta Ewa Creek, Niger Delta Region, Nigeria. *Journal of Agriculture and Veterinary Science, (IOSR)*. 2013;3(3):56-60.
  9. Allison ME, Okadi D, Akpan II. Species distribution and abundance in the Lower Nun River, Niger Delta, Nigeria. *J Fish. Inter*. 2009;4(4):13-18.
  10. Arthington AH, Dulvy NK, Gladstone W, Winfield IJ. Fish Conservation in Freshwater and Marine Realms: Status, Threats and Management. *Aquatic Conservation Marine and freshwater Ecosystems*. 2016;26(5):838-857.
  11. Bonjoru R, Abubakar KA, Hassan EI, Jerry TJ. Diversity and Abundance of Fish Species in some Selected Riverine Wetlands of Upper Benue River Basin, Nigeria. *Journal of Environmental Science, Toxicology and Food Technology*. 2019;13(8 Ser. II):14-18.
  12. Cosgrove WJ, Rijshberman FR. World water vision making water everybody's business. Earthscan, London, 2000, 108.
  13. David DL, Wahedi JA, Buba UN, Ali BD, Barau BW. A Study of Fish Diversity of Two Lacustrine Wetlands in the Upper Benue Basin. Nigeria. *Annual Research and Review in Biology*. 2016;7(5):318-328.
  14. Dike CC, Agunwamba JC. A Study on the Effects of Tide on Sedimentation in Estuaries of the Niger Delta, Nigeria. *Journal of Urban and Environmental Engineering. JSTOR*. 2012;6(2):86-93.
  15. Dugeon D, Arthington AH, Hessner MO, Kawabata ZI, Knowler DJ. Freshwater Biodiversity: Importance, Threats, Status and Conservation Challenges. *Bio. Rev*. 2006;81:163-182.
  16. Edwards AJ, Anthony CG, Abohweyere PO. A revision of Irvine's Marine Fishes of Tropical West Africa. Darwin Initiative Report 2, Ref. 2001;162(7):451-157.
  17. Essen AA. Review of the fisheries resources of Akwa Ibom State and the need for effective conservation. In transaction of the Nigeria Society for Biological

- Conservation. 1990;1:116-120.
18. Essien-Ibok MA, Isemin NL. Fish fauna of Akwa Ibom State Inalndn Water. *Biodiversity Int. J.* 2020;4(2):82-88.
  19. Fisher W, Bracher G, Scott WB. FAO Species identification sheet for fisheries purpose. Eastern Central Atlantic Fishing Areas. Rom/OFM, Ottawa. 1981;6(34):4-7.
  20. Fu C, Wu J, Chin J, Wu Q, Lei G. Fish Diversity of Two Lacustrine Wetlands of the Upper Benue Basin. Nigeria World Academy of Science, Engineering and Technology International Journal of Biological, Bimolecular Agricultural, Food and Biotechnological Engineering. 2003;10(5):294-298.
  21. Gaines SD, White C, Carr MH, Palumbi SR. Designing marine reserve networks for both conservation and fisheries management. *Proceedings of National Academy of Sciences.* 2009;107(1):18286-18293.
  22. Godwin SA, Anthony AN, James PU. Diversity and Distribution of Freshwater Fishes in Oguta Lake, Southeast Nigeria. *Advances in Life Science and Technology,* 2016, 46.
  23. Guo Q, Liu X, Ao X, Qin J, Wu X, Ouyang S. Fish Diversity in the Middle and Lower Reaches of the Ganjiang River of China: Threats and Conservation. *PlosINE,* 2018;13(11).
  24. Gupta I, Dhage S, Kumar R. Study of Variations in Water Quality of Mumbai Coast through Multivariate Analysis Techniques. *Indian Journal of Marine Sciences.* 2009;38(2):170-177.
  25. Hannah L. *Climate change Biology* (2<sup>nd</sup> edition), Academic Press, 2015, 470. ISBN 978-0-124-20218-4.
  26. Holden M, Reed W. *West African Freshwater fishes.* Longmand, 1972, 67p.
  27. Idodo-Umeh G. Pollution assessment of Olomoro water bodies using physical-chemical and biological indices: Ph.D. Thesis University of Benin, Benin City, Nigeria, 2002, 455.
  28. Idodo-Umeh G. *Freshwater Fishes of Nigeria* (Taxonomy, Ecological Notes, Diets and Utilization), Idodo-Umeh Publishers, Benin, Nigeria, 2003.
  29. Jamu DM, Ayinla OA. Potential for the Development of Aqua Culture in Africa NAGA. 2003;26(3):9-13.
  30. King RP. Biodiversity of freshwater fishes of the Cross River in the rainforest belt of Cameroon-Nigeria. *J Aquat. Sci,* 1992, 184-197.
  31. King RP. Distribution, abundance, size and feeding habits of *Brienomyrus brachyistus* (Gill, 1862) (Teleostei: Mormyridae) in a Nigerian Rainforest Stream. *Cybium.* 1989;13(1):25-36.
  32. King RP, Jonathan GE. *Aquatic environmental perturbations and monitoring African Experience,* USA, 2003, 166p.
  33. Komolafe OO, Arawonio GAO. Preliminary observations on fish species in a newly impounded Osinmo Reservoir. *Turkish J of fisheries and Aq Science.* 2008;8:289-282.
  34. Moses BS. The influence of flood regime on fish catch communities of the Cross River floodplain ecosystem, Nigeria. *Envir. Bio of Fish.* 1987;18(1):56-66.
  35. Negi RK, Sheetal M. Species Diversity, Abundance and Distribution of Fish Community and Conservation Status of Tons River of Ultrakhand State, India. *J Fish Aq. Sc.* 2013;8:617-626.
  36. Nwadiaro CS. Ichthyofauna of Lake Oguta, a shallow lake in South-Eastern Nigeria. *Arch. Hydrobiol.* 1989;115(3):463-475.
  37. Odo GE, Nwani CD, Eyo JE. The Fish Fauna of Anambra River Basin, Mogeroa: Species Abundance and Morphometry. *Revista de Biologia Tropical.* 2009;57(1-2);177-186.
  38. Onuoha GC, Ekpo IE, Chude LA, Isangedighi IA. Composite preliminary ichthyofaunal survey of Ntak Inyang Stream, Ikpa River, Niger Nig. *J Ag. Fool and Evt.* 2010;6(1 x 2):82-89.
  39. Orji RCA, Onyejiaka C. Preliminary investigation of fish species potentials of Imo River in Nigeria. *S. Aq. Sc.* 1990;5:83-91.
  40. Peters SW, Iwok ER, Uya OE. *Akwa Ibom: The land of promise* (Editors-Peters, Iwok SW, ER and Uya OE.) Gabumo publishing Co. Ltd, 1994, 377p.
  41. Schineider W. Field guide to the commercial marine resources of the Gulf of Guinea. FAO. Rome, 1990, 227p.
  42. Udoidiong OM, King RP. Ichthyofaunal Assemblages of some Nigerian Rainforest Streams. *Journal of Aquatic Sciences.* 2010;15:1-8.
  43. Valiky JM. Assessing and managing the marine fish resources of Sierra Leone, West Africa. *NAGA.* 1992;15(1):31-38.
  44. Welcome RL. *Rivers fisheries.* FAO Fish. Tech. Pap. 1985;262:330.
  45. Welcome RL. *Fish of the Niger system In: Ecology of River System: Davies, B. R. and K. F. Walker (eds) Dr. W. Junks Publishers, Dordrecht. Netherlands,* 1986, 25-45.