



# International Journal of Fisheries and Aquatic Studies

E-ISSN: 2347-5129

P-ISSN: 2394-0506

(ICV-Poland) Impact Value: 5.62

(GIF) Impact Factor: 0.549

IJFAS 2019; 7(5): 442-445

© 2019 IJFAS

www.fisheriesjournal.com

Received: 03-07-2019

Accepted: 07-08-2019

**Idowu TA**

Department of Fisheries,  
Modibbo Adama University of  
Technology, Yola, Nigeria

**Denham SA**

Department of Fisheries,  
Modibbo Adama University of  
Technology, Yola, Nigeria

**Adedeji HA**

Department of Fisheries,  
Modibbo Adama University of  
Technology, Yola, Nigeria

## Growth performance and hematological indices of *Clarias gariepinus* (Burchel, 1822) fingerlings fed varying levels of *Telfairia occidentalis* leaf meal additives

**Idowu TA, Denham SA and Adedeji HA**

### Abstract

As aquaculture production become more intensive, the sustainability of the industry depends upon the availability of suitable and economical feeds, which are organically derived. Essential information is needed especially on organically derived feed additives that can enhance diet digestibility and immune responses. This study evaluates the performance of *Clarias gariepinus* fingerlings fed varying levels of *Telfairia occidentalis* leaf meal additives. The plant leaves were shade dried to constant weight and milled to powder form. Five diets; T<sub>1</sub>, T<sub>2</sub>, T<sub>3</sub>, T<sub>4</sub> and T<sub>5</sub> containing 0g (control), 0.5%, 1.0%, 1.5% and 2.0% *Telfairia occidentalis* leaf meal additives respectively were formulated. The diets were fed to *C. gariepinus* fingerlings for eight weeks. The mean weight gain were significantly different ( $p < 0.05$ ) with diet T<sub>3</sub> having the highest value,  $25.93 \pm 2.11$ g and T<sub>5</sub> the least value,  $17.93 \pm 1.81$ g. Diet T<sub>1</sub>, T<sub>3</sub>, T<sub>4</sub>, and T<sub>5</sub> had 85% survival and diet T<sub>2</sub> had the least survival, 75% however, with no significant difference ( $p > 0.05$ ). The haematology profiles of the experimental fish indicated significant differences ( $p > 0.05$ ) in all the parameters examined. The erythrocytes and leukocytes were highest for diet T<sub>3</sub> with  $3.17 \pm 0.27$  and  $285.10 \pm 2.10$  respectively. The proximate analysis of the carcass of *C. gariepinus* indicated diet T<sub>1</sub> having higher crude protein content (63.59%) than other treatments however with no significant difference ( $p > 0.05$ ). *Telfairia occidentalis* leaf meal can be used as plant additives in the diet of *Clarias gariepinus*.

**Keywords:** Aquaculture, growth performance, haematological profiles, *Clarias gariepinus*, fluted pumpkin, *Telfairia occidentalis*

### Introduction

Aquaculture is the fastest growing food producing sector, compared to other food commodities [1]. FAO [2] projected the total annual demand for fish and fishery products to expand by over 50 million metric tons from year 2015; and explained that 73% of supply is expected to come from aquaculture. Likewise, the demand for animal protein in Nigeria is increasing with increase in human population and aquaculture is an important option to meet this demand [3]. As aquaculture production become more intensive in Nigeria, fish feed has been identified to be a significant factor in increasing its productivity and profitability [4, 5]. Information on type, quality, quantity, seasonality and cost of fish feeds is important in determining the appropriate production strategy in aquaculture [6]. However, essential information is needed especially on organically derived feed additives that can enhance diet digestibility and immune responses of culture fish. There are a rich abundance of plants reputed in traditional medicine to possess medicinal values, protective and therapeutic properties [7, 8]. Traditional medicinal plants are a therapeutic resource used by the population of the continent specifically for health care, which may also serve as starting materials for drugs [9].

Public health issues are considered as those of direct importance to both producers and consumers of fish and include broader issues of food production, processing and delivery systems [10]. As aquaculture assumes an expanding role in meeting the increasing consumers' demands and the intensified use of antibiotics and other synthetic drugs to treat and control disease outbreaks. The safety of aquaculture products for consumption has continued to generate anxiety among consumers, posing public health concern globally [11]. Thus, actions are needed to explore better alternatives in other to replace or reduce the use of antibiotics

**Correspondence**

**Idowu TA**

Department of Fisheries,  
Modibbo Adama University of  
Technology, Yola, Nigeria

(synthetic drugs) in aquaculture production to enhance quality and safety of products. Herbal biomedicine has been indicated as promising alternatives [12, 13]. This present study aimed at examining the performance of African mud catfish (*Clarias gariepinus*) fingerlings fed varying levels of fluted pumpkin (*Telfairia occidentalis*) leaf meal additives.

## Materials and Methods

### Preparation of experimental diets

The experiment was carried out at the Department of Fisheries, Modibbo Adama University of Technology, Yola (9.35 00 °N 12.50 11 °E). The feed ingredients and *Telfairia occidentalis* leaves were obtained from Jimeta-Yola. The feedstuffs were given appropriate processing, milled separately using a domestic hammer milling machine, package and stored for use. The fresh leaves of *T. occidentalis* were air dried under shade and milled to a fine powder using mortar and pestle to form *Telfairia occidentalis* leaf meal (TOLM).

The feed ingredients were measured and mixed to obtain a homogeneous mass. TOLM was added at 0% (control), 0.5%, 1.0%, 1.5% and 2.0% and mixed thoroughly with cassava starch as a binder. The resultant dough was pelleted, air-dried under shade to constant weight and stored in the laboratory for further use. The percentage composition of ingredients in experimental basal feed is shown in Table 1. The proximate composition of the experimental basal feed was carried out according to the methods of Association of Official Analytical Chemist Methods [14].

### Feeding trials

One hundred and fifty (150) fingerlings of *Clarias gariepinus* procured from a reputable fish farm in Yola were acclimatized for 7 days and randomly distributed into plastic tanks of 25 litres volume. The fish were distributed at 10 fish/tank representing five treatments in triplicates; 0g (control), 0.5%, 1.0%, 1.5% and 2.0% of TOLM and designated as T<sub>1</sub>, T<sub>2</sub>, T<sub>3</sub>, T<sub>4</sub> and T<sub>5</sub> respectively. Each tank was three quarter (¾) filled with water in a mini-flow through system set up. Fish were fed at 5% of their body weight in two equal meals, 08:00 – 9:00 hours and 17:00 – 18:00hours). The water quality of the system were monitored and kept at acceptable limit during the period of the feeding experiment.

Fish were weighted weekly and feeding rates adjusted accordingly. The experiment lasted for eight weeks. Weekly weight of fish and feed fed were used to assess growth response of fish to diets [15].

### Haematological examination

Five fish were selected randomly from each tank and blood samples were collected for haematological analysis as described in Idowu *et al.* [17].

### Carcass analysis

Five fish were randomly chosen from each treatment and the whole fish body proximate analysis was carried out according to AOAC [14].

### Statistical analysis

Data generated in the study were subjected to analysis of variance (ANOVA) and Duncans LSD at  $p < 0.05$ .

**Table 1:** Composition of the 40% crude protein experimental basal diet

Ingredients	Composition (% dry matter)
Fish meal	32.00
Soybean	33.00
Maize	30.00
Vitamin premixes	1.0
Bone meal	0.5
Lysine	0.5
Methionine	0.5
Palm oil	1.0
Cassava starch	1.0
Common salt	0.5
Total	100.0
Calculate crude protein	39.94

## Results

The proximate compositions of experimental diets; T<sub>1</sub>, T<sub>2</sub>, T<sub>3</sub>, T<sub>4</sub> and T<sub>5</sub> are represented in Table 2. Water quality parameters during the feeding experiment, are indicated Table 3. Table 4 presents growth performance of *Clarias gariepinus* fed experimental diets. The haematological indices of *Clarias gariepinus* fed the experimental diets is presented in Table 5. The proximate analysis of the carcass of *C. gariepinus* fed experimental diets are indicated in Table 6.

**Table 2:** Proximate composition of experimental diets ( $p > 0.05$ )

Diet	Crude protein (%)	Crude lipid (%)	Crude fibre (%)	Ash (%)	Moisture (%)	NFE (%)
T <sub>1</sub>	40.03±0.05	2.01±0.10	9.43±0.11	6.28±0.13	5.33±0.27	36.77±0.29
T <sub>2</sub>	40.05±0.00	2.03±0.15	9.53±0.00	6.12±1.00	5.35±0.45	36.82±0.00
T <sub>3</sub>	40.60±0.00	2.01±0.01	9.52±0.12	6.18±0.04	5.20±0.10	36.70±0.00
T <sub>4</sub>	40.03±0.03	2.00±1.00	9.61±0.11	6.10±0.00	5.23±0.23	37.03±0.33
T <sub>5</sub>	38.79±0.00	2.10±1.00	9.77±0.54	6.36±2.84	5.23±0.00	37.37±0.17

NFE – Nitrogen Free Extract

**Table 3:** Water quality parameters during the experimental period

Parameters	Minimum	Maximum
Temperature (°C)	27.60	29.55
Dissolved oxygen (mg/l)	4.25	5.00
pH	6.60	6.90

**Table 4:** Growth performance of *Clarias gariepinus* fed experimental diets

Parameters	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>
Mean initial weight of fish (g)	3.59±0.62 <sup>a</sup>	3.63±0.36 <sup>a</sup>	3.96±0.47 <sup>a</sup>	3.87±0.55 <sup>a</sup>	3.71±0.60 <sup>a</sup>
Mean final weight of fish (g)	27.52±3.14 <sup>ab</sup>	26.38±2.90 <sup>b</sup>	29.89±2.66 <sup>a</sup>	25.01±2.71 <sup>b</sup>	21.64±2.45 <sup>c</sup>
Mean weight gain (g)	23.93±2.12 <sup>b</sup>	22.75±1.86 <sup>b</sup>	25.93±2.11 <sup>a</sup>	21.14±1.96 <sup>c</sup>	17.93±1.81 <sup>d</sup>
Relative weight gain	566.57±20.19 <sup>a</sup>	478.73±17.09 <sup>b</sup>	554.79±18.45 <sup>a</sup>	446.25±20.27 <sup>b</sup>	283.28±30.23 <sup>d</sup>
Specific growth rate (%/Day)	1.57±0.24 <sup>a</sup>	1.49±0.16 <sup>b</sup>	1.56±0.18 <sup>a</sup>	1.45±0.14 <sup>b</sup>	1.36±0.14 <sup>c</sup>
Initial number of fish stocked	20	20	20	20	20
Final number of fish stocked	17	15	17	17	17
Survival (%)	85	75	85	85	85

Values in the same row with different superscripts are significantly different ( $p < 0.05$ )

**Table 5:** Haematology of *Clarias gariepinus* fed experimental diets ( $p > 0.05$ )

Parameters	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>
Erythrocytes	3.03±0.65 <sup>b</sup>	2.93±0.25 <sup>c</sup>	3.15±0.27 <sup>a</sup>	3.08±0.23 <sup>b</sup>	2.95±0.31 <sup>c</sup>
Leukocytes	283.65±2.45 <sup>ab</sup>	280.00±0.40 <sup>b</sup>	285.10±2.10 <sup>a</sup>	279.90±0.30 <sup>b</sup>	279.30±1.90 <sup>b</sup>
PCV (%)	41.40±1.10 <sup>a</sup>	38.60±1.90 <sup>b</sup>	41.40±1.70 <sup>a</sup>	39.70±0.50 <sup>b</sup>	36.35±3.75 <sup>c</sup>
MCV (fl)	136.40±0.70 <sup>a</sup>	131.45±5.35 <sup>b</sup>	131.90±5.90 <sup>ab</sup>	129.75±8.25 <sup>b</sup>	123.25±0.25 <sup>c</sup>
MCHC (g/dL)	28.95±0.45 <sup>b</sup>	31.10±1.70 <sup>a</sup>	28.25±2.35 <sup>b</sup>	29.35±0.75 <sup>b</sup>	32.50±0.90 <sup>a</sup>
Hb (g/l)	12.00±0.50 <sup>a</sup>	12.05±1.25 <sup>a</sup>	11.75±1.45 <sup>b</sup>	11.65±0.45 <sup>c</sup>	11.78±1.48 <sup>b</sup>
MCH (pg)	39.50±0.80 <sup>a</sup>	41.00±3.90 <sup>a</sup>	37.20±1.40 <sup>b</sup>	38.00±1.40 <sup>b</sup>	40.05±1.05 <sup>a</sup>
Platelets	97.50±18.50 <sup>a</sup>	84.50±4.50 <sup>b</sup>	63.50±12.50 <sup>c</sup>	62.00±4.00 <sup>c</sup>	66.50±6.50 <sup>c</sup>

Values in the same row with different superscripts are significantly different ( $p < 0.05$ )

PCV – Pack cell volume; MCV – Mean cell volume; MCHC – Mean cell hemoglobin concentration; Hb – Haemoglobin; MCH – Mean cell hemoglobin

**Table 6:** Proximate analyses of the carcass of *C. gariepinus* fed the experimented diets ( $p > 0.05$ )

Parameters	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>
Crude protein (%)	63.59	63.10	63.19	62.11	61.83
Crude lipid (%)	13.63	13.10	13.11	13.10	13.05
Crude Fibre (%)	1.97	1.80	1.84	2.00	2.08
Ash (%)	2.94	2.63	2.65	3.10	2.83
Moisture (%)	1.92	1.93	2.01	1.93	1.97
Ether extract (%)	17.25	17.45	17.20	17.66	18.04

## Discussion

The physicochemical parameters of the culture condition for the experimental fish were within the tolerable limits for African catfish [18]. In the present study there was a general increase in weight gain in all treatments, thus indicating that the fish were able to convert feed protein to extra muscle. In fish nutrition, weight gain is usually considered as the most important measurement of productivity of diets [4, 17]. The increase in weight gain reported in all the treatments indicated that the fish responded positively to all the diets and the protein content of the experimental diets enhanced weight gain, growth and dietary energy supply of the fish. However, the observed decrease in the growth of the experimental fish fed diet of TOLM additives may be attributed to the anti-nutritional factors present in leaves of *Telfairia occidentalis*. Fasuyi and Nonyerem [19] reported the presence of phytates and Alkanoids in *Telfairia occidentalis*. The decrease in growth rate with increase inclusion level might be attributed to the presence of these anti-nutritional factors in *Telfairia occidentalis*. These anti nutrients may have affected the digestibility and palatability - taste and even smell of the feed [15].

Blood is a major index of physiological, immunopathological, and nutritional status of an organism. And as reported in Haruna and Adikwu [20], Adeyemo [21] and Sogbesan *et al.* [6], diet compositions among other factors affect the blood biochemical compositions of fish and result to changes in their haematology profiles. In the present study, there were significant differences in all the parameters examined for haematology profiles of the experimental fish fed TOLM

meal diets. The reduction in the values of haematological indices such as packed cell volume, haemoglobin concentration, and erythrocytes could be as a result of nutritionally deficient diet or presence of toxic substance in the diet of fish [22, 23]. The above explanation could be responsible for the haematological values observed in the present study. The change in the blood constituent components could be used to adjudge the health status of an organism [24, 25]. Leukocytes are important blood constituents for defense against unwanted foreign bodies in an organism. Their amount has implication in immune responses and the ability of the animal to defend itself against infections [26, 27]. In this study, the highest Leukocytes recorded for diet T<sub>3</sub> with a corresponding highest value for erythrocytes could be an indication that this inclusion level (1.0%) of fluted pumpkin (*Telfairia occidentalis*) leaf meal in catfish (*Clarias gariepinus*) diet have better stimulatory effects on the fish leucocyte populations as compare to the other inclusions levels (diets).

The proximate values of the carcass of *C. gariepinus* fed fluted pumpkin (*Telfairia occidentalis*) leaf meal indicated the potency of *Telfairia occidentalis* leaf meal as additives from plant origin in the diet of *Clarias gariepinus* for better food fish. However, the carcass of the experimental fish crude protein values reduced with increase in inclusion levels of TOLM, although with no significant difference ( $p > 0.05$ ). The presence of high fiber levels in fish diets can cause intestinal irritation, lower digestibility and overall decreased nutrient utilization [28, 19].

## Conclusion

The results obtained from this study indicated that *Telfairia occidentalis* leaf meal can be used as plant additives in the diet of *Clarias gariepinus*. *Telfairia occidentalis* leaf meal at 1% inclusion could be the preferable level in the diet of *Clarias gariepinus*.

## References

- Adedeji OB, Okocha RC. Constraint to aquaculture development in Nigeria and way forward. *Journal of Applied Sciences Research*. 2011; 7(7):1133-1140.
- FAO Food and Agricultural Organization of the United Nations. The state of world fisheries and aquaculture. FAO Fisheries Department, Rome Italy, 2014, 30.
- Sogbesan OA, Mohanta KN, Sahoo PK, Mitra F, Jayasankar P. Application of solid state fermentation technology in aquaculture. Technical Paper of Central Institute of Freshwater Aquaculture (CIFA), India, 2012, 118-128.
- Sogbesan OA, Ugwumba AA, Madu CT. Nutritive potential and nutrient utilization of garden snail (*Limnicolaria aurora*; Jay 1937 Gastropoda, limnicolaria) meat meal in the diet of *Clarias gariepinus* (Burchell, 1982) fingerlings. *African Journal of Biotechnology* 2006; 5(20):1999-2003.
- Akinrotimi OA, Abu OMG, Aranyo AA. Environmental Friendly Aquaculture Key to Sustainable Fish Farming Development in Nigeria. *Continental Journal Fisheries and Aquatic Science*. 2011; 5(2):17-31.
- Sogbesan OA, Madu CT, Ugwumba AA. Culture and utilization of tadpole as animal protein supplement in the diet of *Heterobranchus longifilis* fingerlings. *Journal of Fisheries and Aquatic Sciences*. 2007; 2(5):353-360.
- Kubmarawa D, Ajoku GA, Enwerem NM, Okorie DA. Preliminary phytochemical screening of 50 medicinal plants from Nigeria. *African Journal of Biotechnology*. 2007; 6(14):1690-1696.
- Mustapha BA, Kubmarawa D, Shagal MH, Ardo BP. Preliminary phytochemical screening of medicinal plants found in the vicinity of quarry site in Demsa, Adamawa state, Nigeria. *American Chemical Science Journal*. 2016; 11(2):1-7.
- Sofowora A. Medicinal plants and traditional medicine in Africa. 2nd Edition, Spectrum Books Ltd., Ibadan, Nigeria, 1993, 289. ISBN-13: 9782462195.
- Idowu TA. A review on pharmacognosis applications in good aquaculture practices. Seminar PhD. Department of Fisheries, Modibbo Adama University of Technology, Yola, Nigeria, 2015; 41.
- INFOFISH Handbook on organic aquaculture. CFC/FAO/INFOFISH Project on Organic Aquaculture in Myanmar, Thailand and Malaysia. INFOFISH, Lumpur, Malaysia, 2011, 35.
- Syahidah A, Saad CR, Daud HM, Abdelhadi YM. Status and potential of herbal applications in aquaculture: A review. *Iranian Journal of Fisheries Sciences*. 2014; 14(1):27-44
- Ugoala E, Ndukwe GI, Ayo RG, Mustapha B. Natural products and aquaculture development. *Journal of Pharmacy and Biological Science*. 2014; 9(2):70-82.
- AOAC Association of Official Analytical Chemists. 18<sup>th</sup> ed. Official methods of analysis. Washington D.C, 2000.
- Sogbesan OA. Culture and utilization of some non-conventional animal protein feedstuffs in the diet of *Heterobranchus longifilis* (Valenciennes, 1840). Thesis PhD. Department of Zoology, Faculty of Science, University of Ibadan, Nigeria, 2007, 252.
- Kori-Siakpere O, Ake JEG, Idoge E. Haematological characteristics of the African snakehead, *Parachanna obscura*. *African Journal Biotechnology*. 2005; 4(6):527-530.
- Idowu TA, Ishaku AH, Adedeji HA, Olaniyi IJ, Sogbesan OA. Effect of varying levels of fermented sorghum (*Sorghum bicolor*) on growth and haematology profiles of *Clarias gariepinus* (Burchell 1822) fingerlings. *Journal of Agriculture and Environment*. 2016; 12(1):89-96.
- Olurin KB, Olojo AA, Olukoya OA. Growth of African catfish *Clarias gariepinus* fingerlings fed different levels of cassava. *World Journal of Zoology*. 2006; 1(1):54-56.
- Fasuyi AO, Nonyerem AD. Biochemical nutritional and haematological implications of *Telfairia occidentalis* leaf meal as protein supplement in broiler starter diets. *African Journal of Biotechnology*. 2007; 6:1055-1063.
- Haruna AB, Adikwu IT. Haematological responses to non-familiar diets. A study of the African catfish (*Clarias gariepinus*). *Journal of Aquaculture in Tropics*. 2001; 10:183-191.
- Adeyemo OK. Haematological and histopathological effects of cassava mill effluent in *Clarias gariepinus*. *African Journal of Biomedical Resources*. 2005; 8:179-183.
- Tacon AGJ. Nutritional fish pathology morphological signs of nutrient deficiency and toxicity in farmed fish. FAO Tech. Paper No. 330. Rome, FAO, 1992, 75.
- Osugwe DI, Obiekezie AI, Onuoha GC. Some haematological changes in hybrid catfish (*Heterobranchus longifilis* x *Clarias gariepinus*) fed different dietary levels of raw and boiled jackbean (*Canavalia ensiformis*) seed meal. *African Journal of Biotechnology*. 2005; 4(9):1017-1021.
- Babatude GM, Fajimi OA, Oyejide OA. Rubber seed oil versus palm oil in the broiler chicken diet: Effect on performance nutrient, digestibility, haematology and carcass characteristics. *Animal Feed Science Technology*. 1992; 35:133-146
- Oyawoye EO, Ogunkunle M. Physiological and biochemical effects of raw jack beans on broilers. *Proceedings of Annual Conference of Nigerian Society of Animal Production*. 1998; 23:141-142.
- Douglass JW, Janes KW. In: Schalm's Veterinary Haematology. John Wiley and Sons. Blackwell publishing Ltd, 2010, 1232.
- Dhayanithi NB, Kumar TTA, Balasubramanian T, Tissera K. A study on the effect of using mangrove leaf extracts as a feed additive in the progress of bacterial infections in marine ornamental fish. *Journal of Coastal Life Medicine*. 2013; 1(3):217-224.
- Fasuyi AO. Nutritional potentials of some tropical vegetable leaf meals: Chemical characterization and functional properties. *African Journal of Biotechnology*. 2006; 5:49-53.