



E-ISSN: 2347-5129

P-ISSN: 2394-0506

(ICV-Poland) Impact Value: 5.62

(GIF) Impact Factor: 0.549

IJFAS 2019; 7(2): 60-63

© 2019 IJFAS

[www.fisheriesjournal.com](http://www.fisheriesjournal.com)

Received: 11-01-2019

Accepted: 16-03-2019

**Ashaq Hussain Bhat**

Department of Zoology, GDC  
Women Anantnag, Jammu and  
Kashmir, India

**Samiea Hussein**

Department of Zoology, JS  
University Shikohabad, Uttar  
Pradesh, India

## Assessment of some seasonal variables on some blood parameters of a local fish '*Schizothorax plagiostomus*' in 'River lidder' in Kashmir valley (India)

**Ashaq Hussain Bhat and Samiea Hussein**

### Abstract

Present work was carried out in order to find out some seasonal variables on some blood parameters of *Schizothorax plagiostomus* caught from the world-famous River lidder (Pahalgam Kashmir, India). Effects of the independent variables e.g., sex, weight and length values were determined seasonally. Analysis revealed that highest number of leukocytes was found in spring and lowest number was found in winter. Hemoglobin and Haematocrit values were highest during the months of summer and lowest during winter. It was also seen that Males were having greater values for hemoglobin and Haematocrit than females, whereas total leukocyte count in females was higher than males. It was also found that there was a positive correlation between Length/weight and Hemoglobin and Haematocrit values where as a negative correlation between length or weight and Total leukocyte counts.

**Keywords:** Schizothorax, leukocytes, haematocrit values, correlation

### Introduction

Due to great importance of blood in the diagnosis of diseases in medicine, hematological and serological investigations are now employed on an increasing scale for diagnostic purposes in fish pathology. Hematology concerns with the cells present in the blood and serology deals with the constituents in the fluid part of the blood, such as proteins, minerals, carbohydrates, pigments, enzymes, hormones, immunoglobulin's etc. Hematological investigations on fish have confirmed that the variations in the blood give indications of health conditions (Soivio, A, and A Oikari. 1976) [33].

Marked variations in hematological data, caused by the environment would necessitate an exact establishment of normal values of various environmental conditions and in different seasons of the year. In fisheries it is important to diagnose the illness of a fish by evaluating hematological data, particularly blood parameters (Ramesh M, *et al*, 2008) [32].

To picturize the actual hematological data is quite different because of marked internal and external variations sampling techniques, laboratory techniques, seasonal variations, habitat size, sex, population density, oxygen content, pH, water, ammonia, feeding etc. (Wilhelm Filho *et al*, 2001) [16]. Research on blood of Cyprinus Carpio had shown that blood parameters were affected by seasonal variations (Denton and Yousuf, 1975) [14]. In addition to this age, sex, weight and length of fish also affect the blood parameters of fish (Ezzat A *et al*, 1982) [20].

### Materials and Methods.

In the present study 10 male and female specimens of *S. plagiostomus* were collected from River lidder with the help of local fisherman. Sex, weight and length of the specimens was male and females, 180-3100gms and 22-33 cms respectively. Many techniques were utilized for the collection of blood e.g., Heart punctures, severing of caudal peduncle (Blaxhall and Daisley, 1973; Hatting 1975) [8].

Blow was given on the head of the fish and the needle was inserted at an angle of 45° in to the heart and the blood was collected in to the glass tubes already containing EDTA (Ethylene diamine tetra acetic acid). Blood analysis was carried out very quickly after sampling. Different leukocyte counting was performed by transporting blood samples diluted with turck's solution with a leukocyte pipette on to a neubar's counting chamber and examined for leukocytes (Blaxhall and Daisley, 1973) [8].

**Corresponding Author:**

**Ashaq Hussain Bhat**

Department of Zoology, GDC  
Women Anantnag, Jammu and  
Kashmir, India

Leishman staining was used to determine the percent leukocyte (Goel *et al*, 2011) [21]. The hemoglobin was determined according to the method of Haldane. Haematocrit values were determined by the microhaematocrit (Jewet *et al*, 1991) [22].

The blood without EDTA was transferred in to Haematocrit pipettes and centrifuged at 10,000×g rpm for 10 minutes and the ratio of the blood components in plasma was determined. The slides were examined under a binocular microscope with 100x objectives to see the percentage of neutrophils, eosinophils, lymphocytes and monocyte.

## Results

Water quality plays a vital role in determining any hematological parameter of the fish. Water quality criteria like Temperature of River liddar between January 2018 and December 2019 was different in different seasons. Total leukocyte count, eosinophil, neutrophil and monocyte percentages were significantly ( $p<0.05$ ) higher for both sexes in spring -summer seasons than those of other seasons. Lymphocyte ratio also increased ( $p<0.05$ ) in autumn-winter season. The amount of hemoglobin and Haematocrit value increased significantly ( $p<0.05$ ) in spring-summer season.

**Table 1:** Seasonal variations of total leukocytes and its types in males of *S. plagiostomus*.

Season/Temp.	No. of fishes	Total leukocyte count(x10/mm <sup>3</sup> )	Neutrophil%	Eosinophil%	Lymphocyte%	Monocyte%
Spring/16°C	10	13.00±0.20	26.10±0.50	1.45±0.1	61.40±1.50	12.20±0.30
Summer/28°C	10	12.60±0.50	25.40±0.40	1.02±0.2	64.70±1.20	9.50±0.50
Autumn/20°C	10	11.40±0.50	24.40±0.20	0.950±0.5	68.50±0.50	6.50±0.20
Winter/5°C	10	9.50±0.50	21.32±0.70	0.92±0.3	63.20±0.30	8.60±0.70

**Table 2:** Seasonal variations of total leukocytes and its types in females of *S. plagiostomus*.

Season/Temp.	No. of fishes	Total leukocyte count(×10/mm <sup>3</sup> )	Neutrophils%	Eosinophils%	Lymphocyte%	Monocyte%
Spring/16°C	10	18.00±0.50	33.00±0.70	1.05±0.02	53.05±0.30	11.50±0.17
Summer/28°C	10	16.50±0.30	29.50±0.50	0.90±0.50	61.02±1.20	12.70±1.09
Autumn/20°C	10	13.70±0.70	23.50±0.30	0.40±0.30	67.00±0.30	10.05±0.50
Winter/5°C	10	10.50±0.20	21.30±0.70	0.48±0.50	73.05±0.10	10.50±1.51

**Table 3:** Male dependent seasonal variations in Hemoglobin and Haematocrit values.

Season/Temp.	No. of fishes	Hemoglobin Density(g/dl)	Haematocrit values (%)
Spring/16°C	10	9.50± 0.05	33.80± 0.30
Summer/28°C	10	11.20± 0.20	37.65± 0.45
Autumn/20°C	10	7.30± 0.70	31.50 ±0.37
Winter/5°C	10	7.09 ± 0.30	25.30 ± 0.70

**Table 4:** Female dependent seasonal variations in Hemoglobin and Haematocrit values.

Season/Temp.	No. of fishes	Hemoglobin Density(g/dl)	Haematocrit values (%)
Spring/16°C	10	6.50± 0.08	28 ± 0.50
Summer/28°C	10	8.20± 0.05	28± 0.20
Autumn/20°C	10	5.90 ± 0.09	22± 0.10
Winter/5°C	10	5.50 ± 0.08	17.80± 0.20

**Table 5:** Comparison of Blood parameters between male and female *S. plagiostomus* between March 2018 to Mar.2019.

Blood parameters	Males	Females
Total leukocyte count	13.52± 1.50	11.50 ± 0.50
<b>Leukocyte types (%)</b>		
Neutrophils	25.20± 0.50	29.50± 0.30
Eosinophils	0.70 ± 0.01	0.40 ± 0.01
Lymphocytes	0.50± 0.80	55.20± 1.50
Monocyte	8.50 ± 1.00	10.50± 0.80
Hemoglobin Density(g/dl)	7.20 ± 0.50	6.02 ± 0.50
Haematocrit values (%)	28.50± 0.70	22.50± 0.23

**Table 6:** Length-weight group dependent frequencies of hematological parameters of *S. plagiostomus*.

Parameter	Parameter			
	10	10	10	10
No. of fishes	10	10	10	10
Length(cm)	17-23	24-30	31-33	29-33
Weight(gm)	180-210	180-280	282-320	300-320
Hemoglobin Density(g/dl)	6.07 ± 0.12	6.18± 1.50	7.12 ± 0.32	9.50 ± 0.3
Haematocrit values (%)	19.55± 0.50	20.32± 1.62	23.55± 1.32	25.80± 1.32
TLC(×10/mm <sup>3</sup> )	25.42± 1.30	22.60± 0.80	22.70± 1.30	21.80± 0.50
Neutrophils (%)	10.60± 1.50	11.50± 0.20	14.50± 1.30	11.30± 0.20
Eosinophils (%)	0.60 ± 1.31	0.49 ± 0.05	0.55 ± 0.30	0.55 ± 0.70
Lymphocytes (%)	83.80± 0.50	78.50± 1.32	82.30 ± 1.32	77.30± 0.30
Monocyte (%)	09.50± 0.50	11.30± 0.50	11.20± 0.70	12.50± 0.40

The leukocyte count, neutrophils and monocyte percentages were compared regarding sex difference, they were found to be higher in females than males but the level of hemoglobin and Haematocrit values and lymphocyte ratios were found to be higher in males than females. The amount of hemoglobin, Haematocrit value, neutrophil and monocyte ratio also increased in length and weight but lymphocyte ratio decreased significantly.

### Discussion

Although less work has been done on the hematology of *S. plagiostomus* there are few reports of normal blood values and published values are severely limited by low fish numbers. Annual changes in blood parameters showed a clear increase in spring-summer period and a decrease in autumn-winter period. The leukocyte numbers and Neutrophil ratios for female specimen were found to be higher compared to male specimen during the reproduction period (March-August). The same trend occurred with the values of hemoglobin, haematocrit, neutrophil and monocyte percentages. Our results are in accordance with other results of (Denton and yousuf, 1975; Ezzat, A. A, *et al.* 1974; Beelen, R. *et al.*, 1998; Azizoglu and Cengizler, 1996)<sup>[14, 18, 4]</sup>. When sex was taken into consideration, the amount of hemoglobin, Haematocrit and lymphocyte values for male fish were significantly higher than for female fish. Monocyte ratio, neutrophil and leukocyte values were higher in females at reproduction periods than in males. So, our results are in accordance with the results of Ezzat *et al.* 1974; Murray 1984; Terao *et al.* 1984; Lumsden 2011<sup>[18, 28, 34, 27]</sup>. Length to weight independent variables were determined to cause an increase in hemoglobin, hematocrit, neutrophil and Monocyte values but the ratio of lymphocytes decreased to a moderate level. The length weight variables on blood parameters (lymphocyte formation) are an essential component of immune system in the early stages of growth period.

To conclude, the large variation in hematological parameters obtained emphasizes the need of more extensive study. The changes in micro-environment and the macro-environment have a direct effect on the blood components of fish which makes the comparison difficult to be made. Evaluation of hematological analyses will enhance the culture of fish by facilitating early detection of infectious diseases and identification of sub lethal conditions affecting production performance this will contribute to more specific, timely and effective disease treatment in the future. Therefore, we are of this opinion that there should be separate data collection and comparison from healthy and unhealthy fish to obtain reliable hematological data. These results of our study may be used as reference values of blood parameters for other fishes.

### References

1. Alcorn SW, Murray AL, Pascho RJ. Effects of rearing temperature on immune functions in sockeye salmon (*Oncorhynchus nerka*). *Fish & Shellfish Immunology* 2002;12:303-334.
2. Alexander N, *et al.* Hematological characteristics of albacore, *Thunnus alalunga* (Bonnaterre), and skipjack, *Katsuwonus pelamis*. *Jour. Fish Biol* 2020;16:383-395.
3. Alvarez-Pellitero, *et al.* Some blood parameters in sea bass, *Dicentrarchus labrax*, infected by bacteria, virus and parasites. *Jour. Fish Biol* 2019;31:259-261.
4. Beelen R, van der Heijden T, Booms R, *et al.* Blood values of young Brazilian catfish *Pseudoplatystoma corruscans* (Agassiz, 1829). *Acta Sci* 1998;20:147-150.
5. Bennet MF, *et al.* Effects of cold shock on the distribution of leucocytes in goldfish, *Carassius auratus*. *Jour. Comp. Physiol* 2019;98:213-216.
6. Boyar HC. Blood cell types and differential cell counts in Atlantic herring, *Clupea harengus harengus*. *Copeia* 2019;2:463-465.
7. Burrows F, *et al.* Blood leucocytes of the turbot, *Scophthalmus maximus* (L.). *Aquaculture* 2020;67:214-210.
8. Blaxhall PC, Daisley KW. Routine haematological methods for use with fish blood. *Journal of Fish Biology* 1973;5:571-604.
9. Bridges DW, Cech Jr J, Pedro DN. Seasonal hematological changes in winter flounder, *Pseudopleuronectes americanus*. *Transactions of the American Fisheries Society* 1976;105:596600.
10. Burrows AS, Fletcher TC. Blood leucocytes of the turbot *Scophthalmus maximus* L. *Aquaculture* 1987;67(2):14-2. 10.
11. Catton J. Blood cell formation in certain teleost fishes. *Blood* 1951, 39-60.
12. Cengizler I, Sahan (Azizoglu) A. Determination of some blood parameters in mirror carps (*Cyprinus carpio*, Linnaeus, 1758) living in Seyhan Dam Lake and Seyhan River. *Turk J Vet Anim Sci* 2000;24:205-214. (in Turkish).
13. Conroy DA. Studies on the haematology of Atlantic salmon (*Salmo salar* L.). *Symposia of the Zoological Society of London* 1972;30:101-127.
14. Denton JE, Yousef MK. Seasonal changes in haematology of rainbow trout, *Salmo gairdneri*. *Comparative Biochemistry and Physiology SIA* 1975;10:1-1 53.
15. Dunn SE, Murad A, Houston AH. Leucocytes and leucopoietic capacity in thermally acclimated goldfish, *Carassius auratus* L. *Journal of Fish Biology* 1989;34:223-230.
16. Wilhelm Filho D, *et al.* Influence of season and pollution on the antioxidant defenses of the Cichlid fish acara (*Geophagus brasiliensis*). *Brazilian Journal of Medical and biological research* 2001.
17. Ellis AE. Leukocytes and related cells in the plaice *Pleuronectes platessa*. *Journal of Fish Biology* 1976;8:143-106.
18. Ezzat AA, *et al.* Studies on the blood characteristics of *Tilapia zilli* (Gervais). *Journal of Fish Biology* 1974;6:1-12.
19. Enomato Y. On some notes about the fluctuations of the blood leucocyte numbers of the cultured fish. *Bull. Tokai reg. Fish. Research Lab* 2020;57:137-177.
20. Ezzat AA, Mikhail MY, Wadie WF, Hashem MT. Length weight relationship and condition factor of *Epinephelus aeneus* and *Epinephelus alexandrinus* in the Egyptian Mediterranean waters. *Bull. Inst. Oceanogr. Fish* 1982;8:173-185.
21. Goel C, *et al.* Length weight relationship of snow trout (*Schizothorax richardsonii*) based on linear and nonlinear models from hill stream of Uttarakhand, India. *World J Fish and Marine Sci* 2011;3(6):485-488
22. Jewet MGD, Behmen J, Johnson GH. Effects of hyperoxic rearing water on blood haemoglobin and haematocrit level of Rainbow trout. *J Aquatic Anim Health* 1991;3:153-160

23. Kekic H, Ivanc A. A new direct method for counting fish blood cells. *Ichthyologia* 1982;14:55-58.
24. Kori-Siakpere. Haematological characteristics of *Clarias iberiensis* Sydenham. *Journal of Fish Biology* 1985;27:259-263.
25. Korzhuev PA, Alyorkrinskaya C, Dgora SN. Characteristics of the blood in young and adult *Salmo salar* (Salmonidae). *Journal of Ichthyology* 1982;22:112-120.
26. Lehmann J, Stiirenberg FJ. *Haematologisch serologisch Suhstratunter-suchungen an der Regenbogenforelle (Salmo gairdneri Richardson)*. Krefeld-Miils: Kaltenmeier 1975, 901-911.
27. Lumsden JS. Use of Hydrogen Peroxide to Treat Experimentally Induced Bacterial Gill Disease in Rainbow Trout. *Journal of Aquatic Animal Health* 2011.
28. Murray SA. Hematological study of the bluegill, *Lepomis macrochirus* Raf. *Comparative Biochemistry and Physiology* 1984;78A:787-791.
29. Peters G, Schwarzer R. Changes in hemopoietic tissue of rainbow trout under influence of stress. *Diseases of Aquatic Organisms* 1985;1:1-10.
30. Pickering AD. Changes in blood cell composition of the brown trout *Salmo trutta* L., during the spawning season. *Journal of Fish Biology* 1986;29:332-347.
31. Preston A. Red blood values in the plaice (*Pleuronectes platessa* L.). *Journal of the Marine Biological Association of the United Kingdom* 1960;39(68):1-687.
32. Ramesh M, Saravanan M. Hematological and biochemical responses in a freshwater fish *Cyprinus Carpio* exposed to chlorpyrifos. *International Journal of Integrative Biology* 2008;3(1):80-83.
33. Soivio A, Oikari A. Hematological effects of stress on a teleost. *Esox lucius* L. *Journal of Fish Biology* 1976;8(3):97-411.
34. Terao Toshiro *et al.* Changes in Fatty Acid Composition of Masu Salmon, *Oncorhynchus masou*, Reared in Sea Water. *Bull. Fac. Fish. Hokkaido Univ* 1984;29(2):155-163.