



# International Journal of Fisheries and Aquatic Studies

E-ISSN: 2347-5129

P-ISSN: 2394-0506

(ICV-Poland) Impact Value: 5.62

(GIF) Impact Factor: 0.549

IJFAS 2018; 6(2): 376-380

© 2018 IJFAS

www.fisheriesjournal.com

Received: 17-01-2018

Accepted: 21-02-2018

**Théophile Godome**

(A). Laboratoire de Recherches sur les Zones Humides, Département de Zoologie, Faculté des Sciences et Techniques, Université d'Abomey-Calavi, BP: 526 Cotonou, Bénin  
(B). Laboratoire d'Hydrobiologie, UFR Biosciences, Université Félix Houphouët-Boigny, 22BP: 582 Abidjan 22, (Côte d'Ivoire)

**Ephrem Tossavi**

Laboratoire de Recherches sur les Zones Humides, Département de Zoologie, Faculté des Sciences et Techniques, Université d'Abomey-Calavi, BP: 526 Cotonou, Bénin

**Nahoua Issa Ouattara**

Laboratoire d'Hydrobiologie, UFR Biosciences, Université Félix Houphouët-Boigny, 22BP: 582 Abidjan 22, (Côte d'Ivoire)

**Emile Didier Fiogbe**

Laboratoire de Recherches sur les Zones Humides, Département de Zoologie, Faculté des Sciences et Techniques, Université d'Abomey-Calavi, BP: 526 Cotonou, Bénin

**Correspondence**

**Théophile Godome**

(A). Laboratoire de Recherches sur les Zones Humides, Département de Zoologie, Faculté des Sciences et Techniques, Université d'Abomey-Calavi, BP: 526 Cotonou, Bénin  
(B). Laboratoire d'Hydrobiologie, UFR Biosciences, Université Félix Houphouët-Boigny, 22BP: 582 Abidjan 22, (Côte d'Ivoire)

## Determination of the optimal feed ration for best growth of *Hoplobatrachus occipitalis* (Günther, 1858) tadpoles reared in controlled medium

**Théophile Godome, Ephrem Tossavi, Nahoua Issa Ouattara and Emile Didier Fiogbe**

### Abstract

The experiment was carried out for 24 days in open circuit made of 18 cemented ponds containing 50 L water each. The current study aims to estimate the optimal feed ration in *Hoplobatrachus occipitalis* tadpoles ( $0.097 \pm 0.026$  g). Six rations (5, 10, 15, 20, 25 and 30% of biomass/day) were tested in triplicate to evaluate their effect on the growth, survival and the carcass composition of these latter. Results showed Specific Growth Rate and Feed Efficiency was not significantly influenced by feed ration ( $P > 0.05$ ). Highest Specific Growth Rates were observed in tadpoles fed on rations of 15 and 20 % of biomass or  $9.059 \pm 0.209$  %/Day and  $9.027 \pm 0.142$  %/Day respectively. Besides, highest mean values of Feed Efficiency were obtained in tadpoles fed on rations of 10 and 15% of biomass or  $0.778 \pm 0.383$  and  $0.757 \pm 0.052$ . Optimal and maximum feed rations in tadpoles of *H. occipitalis* are respectively 6.02 and 21.37 % of biomass per day.

**Keywords:** *Hoplobatrachus occipitalis*, tadpoles, optimal feed ration

### 1. Introduction

Tadpoles are important in most of fresh water ecological communities, excellent indicators of environment quality and biodiversity conservation [1-6]. Due to the destruction of their habitat, overexploitation, pollution and lack of management program, many amphibians' populations are in decrease [7]. A part from their ecological importance, frog meat is an attractive food in European big restaurants since the 16<sup>th</sup> century [8]. It represents a very important source of protein that can be affiliated to chicken meat with a good taste [9]. It is so necessary to rear them not only for species conservation but to satisfy human protein demand. Feed diet is a capital part of rearing requiring an optimization even though we don't mention what tadpoles feed on in the wildlife [10]. Feed ration can influence larvae growth and survival [11]. Similarly, the density, temperature, water quality, photoperiod, feed quality, physical characteristics of the feed etc... are factors influencing the growth of frog tadpoles [12]. The optimal feeding level improves not only growth performances but enable a reduction of production cost [13] as the price of feed represents between 50% and 70% of the production cost [14]. Several studies showed the influence of artificial feed on survival and growth of tadpoles of *Rana catesbeiana* [15], *Litoria ewingii* [16], *Rana temporaria* [17], but a few studies were focused on *H. occipitalis* tadpoles. By the same way, a few studies examined how feeding level can influence growth of amphibians' tadpoles [18]. This study aims to determine the optimal ration of *H. occipitalis*.

### 2. Materials and methods

#### 2.1 Tadpoles province and experimental design

Tadpoles of *H. occipitalis* used in this study were obtained by semi-artificial reproduction on the site of the Research Laboratory on Wetlands (LRZH) of the University of Abomey-Calavi. Mean value of water temperature, pH and dissolved oxygen in rearing ponds are respectively  $28.33 \pm 0.09$  °C,  $7.01 \pm 0.01$  and  $3.98 \pm 0.02$  mg/l.

Experiment was led for 24 days (from September 25<sup>th</sup> to October 18<sup>th</sup>, 2017). Three days after eggs hatching, tadpoles were collected, counted and distributed in an open circuit made of 18 cemented circular ponds containing 70 tadpoles each in 50 L water.

## 2.2 Feeding

Experiment was preceded of a progressive acclimation of tadpoles to physico-chemical conditions where they fed only on live zooplankton and *Artemia salina* nauplii. Starving was progressive as indicated in Table 1. Coppens was the artificial feed used (diameter 0.5-0.8 mm, 56% protein rate and 15% of lipid). Six rations (5, 10, 15, 20, 25 and 30% of tadpoles biomass) were tested on tadpoles (initial mean weight =  $0.099 \pm 0.002$  g) for a 24 days period. Feed was distributed manually every three (03) hours from 8 a.m to 5 p.m.

## 2.3 Growth control

Control fishing happened every three (03) days following by ponds emptying and cleaning. Number of tadpoles and biomass were determined in order to adjust feed ration. After 24 days of rearing, tadpoles were collected from ponds and weighted individually. Zootechnical performances such as Specific Growth Rate (SGR), Survival Rate (SR) and Feed Conversion Rate (FCR) were calculated according to the following formula:

$$\text{SGR (\%)} = 100 \times \frac{\text{Ln}(\text{final body weight}) - \text{Ln}(\text{initial body weight})}{\text{Duration of rearing period (days)}}$$

$$\text{SR (\%)} = 100 \times \frac{\text{Final number of tadpoles}}{\text{Initial number of tadpoles}}$$

$$\text{FCR} = 100 \times \frac{\text{Feed intake (g)}}{\text{Body weight gain (g)}}$$

## 2.4 Water quality monitoring

Water quality monitoring was carried out by measuring temperature, dissolved oxygen and pH. These parameters were measured twice a week (morning: 7:30 a.m and evening: 04:30 p.m) in each pool at 5 cm depth of water using the portable multi-meter (Calypso ORCHIDIS SN-ODEOA 2138). Water renewing was carried out everyday (7 a.m and 6p.m) before and after feeding with 0.5 L/min water flow.

## 2.5 Chemical analyses

Samples were taken before and after experiment for chemical composition analyses (crude protein, fat, dry matter and crude ash) of tadpoles carcass according to the method of [19].

## 2.6 Statistical analysis

Data were encoded in Microsoft Excel 2013. They served to zootechnical parameters calculation. Mean and range were calculated for each parameter. For each repetition, mean data were considered as an observation.

**Table 2:** Mean  $\pm$  range values of dissolved oxygen, pH and water temperature in ponds during the tadpoles rearing period with the different feed rations.

Parameter	Rations (%)					
	5 (R <sub>1</sub> )	10 (R <sub>2</sub> )	15 (R <sub>3</sub> )	20 (R <sub>4</sub> )	25 (R <sub>5</sub> )	30 (R <sub>6</sub> )
DO, mg/l	5,280 $\pm$ 0,330 <sup>a</sup>	5,297 $\pm$ 0,149 <sup>a</sup>	5,323 $\pm$ 0,422 <sup>a</sup>	4,141 $\pm$ 0,308 <sup>b</sup>	3,938 $\pm$ 0,414 <sup>b</sup>	2,791 $\pm$ 0,208 <sup>c</sup>
pH	6,833 $\pm$ 0,104 <sup>a</sup>	6,783 $\pm$ 0,152 <sup>a</sup>	6,838 $\pm$ 0,058 <sup>a</sup>	6,900 $\pm$ 0,133 <sup>a</sup>	6,800 $\pm$ 0,086 <sup>a</sup>	6,605 $\pm$ 0,091 <sup>a</sup>
T, °C	27,538 $\pm$ 0,149 <sup>a</sup>	27,630 $\pm$ 0,186 <sup>a</sup>	27,632 $\pm$ 0,092 <sup>a</sup>	27,433 $\pm$ 0,157 <sup>a</sup>	27,518 $\pm$ 0,127 <sup>a</sup>	27,596 $\pm$ 0,224 <sup>a</sup>

<sup>abc</sup> mean values on the same ligne that are not affected by same letter are significantly different (P < 0.05).

Statistical software R (version 3.4.2) was used for comparisons with 5 % significance. A one way analysis of variance helped to compare zootechnical parameters in different treatments. The Student-Newman-Keuls test (SNK test) was used for different means pair comparisons in case of significant differences among treatments to determine ideal ration, growth and survival rate of *H. occipitalis* tadpoles. The model of Brett & Grove [20] was applied with second order polynomial regression between specific growth rate and feed ration to determine the optimal and maximum feed ration of *H. occipitalis*.

**Table 1:** Feed substitution for *H. occipitalis* tadpoles.

Duration (Days)	Feed
2	100% zooplankton
2	<i>Artemia salina</i> 50% + zooplankton 50%
1	<i>Artemia salina</i> 75% + zooplankton 25%
1	<i>Artemia salina</i> 100%
1	<i>Artemia salina</i> 50% + artificial feed 50%
1	<i>Artemia salina</i> 25% + artificial feed 75%
16	Artificial feed 100%

Artificial feed = Coppens 0.5 – 0.8 mm, 56 % protein rate and 15 % lipid

## 3. Results

Physico-chemical parameters in different ponds are mentioned in Table 2. During experiment period, the highest pH mean value was obtained with ration of 20 % biomass/day. Although, there were no significant differences among pH values (P > 0.05). Dissolved oxygen in rations R1 (5%), R2 (10%) and R3 (15%) was significantly upper than those in R4 (20%), R5 (25%) and R6 (30%).

Growth performances, rations and survival rates of tadpoles are mentioned in Table 3. Survival rate of tadpoles varied between  $79.45 \pm 7.24$  % and  $94.97 \pm 2.09$  %. There were no significant differences between these values (P > 0.05).

The *H. occipitalis* carcass composition at the end of experiment is mentioned in Table 4. Crude proteins, lipids, ash and dry matters were significantly affected by the different feeding rates (P > 0.05). The highest protein rate was observed in tadpoles fed on the R6 (30 %) ration. Concerning ash and dry matter rates, the highest values were observed in tadpoles fed on 10 % (R2) of biomass per day. The highest value of lipid was obtained in individuals fed on the R3 (15%) ration while the lower was observed in those fed on R1. Generally, dry matter, lipid and crude protein of the initial sample are similar to the values obtained after tadpoles feeding with experimental diet.

According to the model of Brett, the optimal and maximum rations of *H. occipitalis* are approximately 6.02%/day and 21.37%/day of tadpoles biomass (Figure 2).

**Table 3:** Initial Biomass (IB), Final Biomass (FB), Survival Rate (SR), Specific Growth Rate (SGR), Feed conversion Rate (FCR) of *Hoplobatrachus occipitalis* tadpoles reared for 24 days in ponds open circuit and fed on different feed rations.

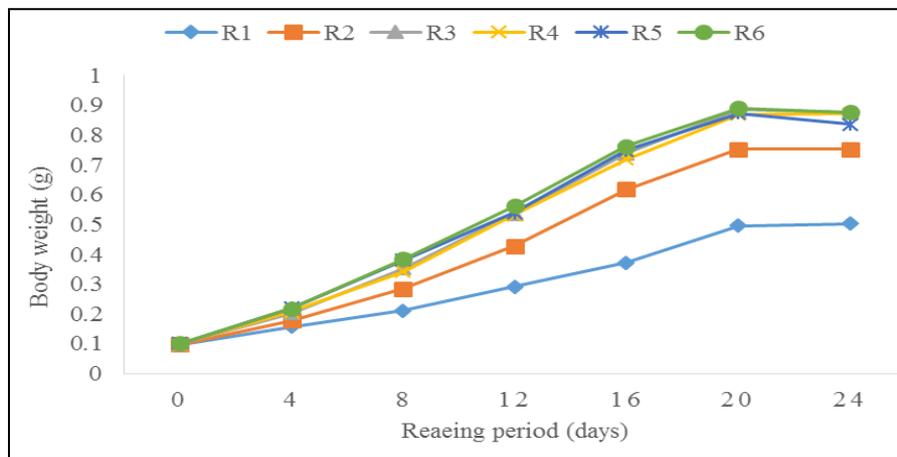
Parameter	Rations (%)					
	5 (R <sub>1</sub> )	10 (R <sub>2</sub> )	15 (R <sub>3</sub> )	20 (R <sub>4</sub> )	25 (R <sub>5</sub> )	30 (R <sub>6</sub> )
IBW (g)	7,076±0,025 <sup>a</sup>	7,180±0,147 <sup>a</sup>	7,260±0,173 <sup>a</sup>	7,283±0,185 <sup>a</sup>	7,390±0,017 <sup>a</sup>	7,377±0,040 <sup>a</sup>
FBW (g)	28,727±4,542 <sup>b</sup>	47,997±22,245 <sup>a</sup>	58,707±4,411 <sup>a</sup>	60,427±3,555 <sup>a</sup>	57,717±3,014 <sup>a</sup>	59,577±1,701 <sup>a</sup>
SR (%)	79,452±7,249 <sup>a</sup>	84,474±13,305 <sup>a</sup>	91,781±1,370 <sup>a</sup>	94,977±2,093 <sup>a</sup>	94,107±3,089 <sup>a</sup>	93,150 ±2,373 <sup>a</sup>
SGR (%/days)	6,772±1,009 <sup>b</sup>	8,253±1,704 <sup>a</sup>	9,059±0,209 <sup>a</sup>	9,027±0,142 <sup>a</sup>	8,814±0,747 <sup>a</sup>	8,999±0,076 <sup>a</sup>
FCR	1,733±0,348 <sup>a</sup>	1,657±1,139 <sup>a</sup>	1,324±0,089 <sup>a</sup>	1,536±0,088 <sup>a</sup>	1,902±0,038 <sup>a</sup>	2,098±0,071 <sup>a</sup>

<sup>abcdefg</sup> mean values on the same ligne that are not affected by same letter are significantly different (P< 0.05).

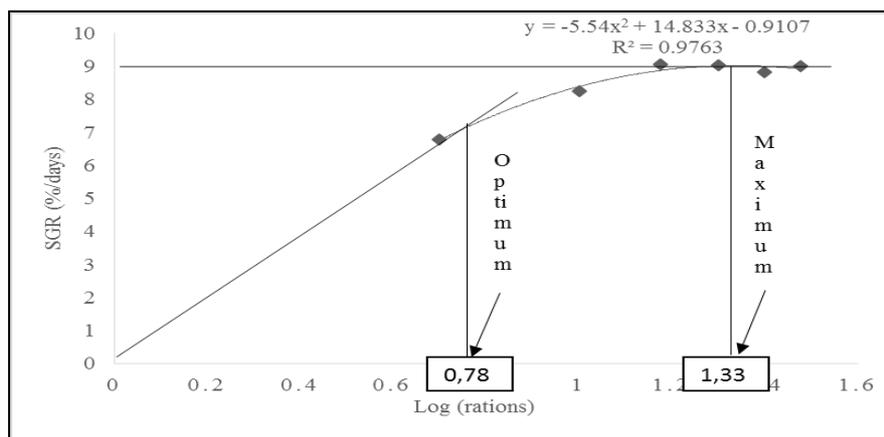
**Table 4:** The composition of *Hoplobatrachus occipitalis* tadpoles' carcass reared for 24 days and fed on six different rations (%/day) with a commercial feed (Coppens).

Analysis	Feeding rate						
	Initial	5%	10%	15%	20%	25%	30%
Dry Matter	89,754±0,001 <sup>b</sup>	87,640±0,004 <sup>f</sup>	90,388±0,001 <sup>a</sup>	89,343±0,001 <sup>c</sup>	85,190±0,004 <sup>g</sup>	89,217±0,002 <sup>d</sup>	88,357±0,005 <sup>f</sup>
Crude ash	10,885±0,003 <sup>b</sup>	7,584±0,003 <sup>d</sup>	11,066±0,002 <sup>a</sup>	7,751±0,003 <sup>c</sup>	6,356±0,007 <sup>g</sup>	7,043±0,001 <sup>e</sup>	6,857±0,006 <sup>f</sup>
Crude lipid	34,604±0,002 <sup>b</sup>	22,709±0,005 <sup>f</sup>	24,743±0,001 <sup>e</sup>	37,550±0,001 <sup>a</sup>	22,183±0,001 <sup>g</sup>	34,483±0,003 <sup>c</sup>	34,280±0,004 <sup>d</sup>
Crude protein	43,789±0,001 <sup>b</sup>	36,423±0,002 <sup>f</sup>	40,758±0,004 <sup>c</sup>	31,457±0,005 <sup>g</sup>	43,160±0,003 <sup>e</sup>	41,706±0,002 <sup>d</sup>	43,868±0,001 <sup>a</sup>

<sup>abcdefg</sup> mean values on the same ligne that are not affected by same letter are significantly different (P< 0.05).



**Fig 1:** Variation of body weight of *Hoplobatrachus occipitalis* tadpoles reared for 24 days with different feed rations.



**Fig 2:** Determination of the optimal and maximum ration of *Hoplobatrachus occipitalis* tadpoles by the model of Brett and Grove after semi-logarithmic transformation of the different feed rations.

#### 4. Discussion

Temperature is the major factor that interferes with frogs' growth due to its direct influence on the metabolism of the subject [18]. Mean value of temperature (27.632 ± 0.092) recorded during the experiment was similar to 26.52 °C obtained by Borges *et al.* [22] in Taurus frog tadpoles. Moreover, these values were upper than 24.68 °C recorded by Ferreira *et al.* [23] in *Lithobates catesbeianus* tadpoles.

Besides, temperatures recorded during this study were situated in the range of 15 – 35 °C recommended for a good frog growth [21]. Mean values of pH and dissolved oxygen recorded during the experiment were in accordance with those obtained by certain authors [23, 24, 22] in *Lithobates catesbeianus* tadpoles.

Tadpoles' growth increases with feed ration. The final biomass recorded ranged from 28.727 ± 4.54 g and 60.427 ±

3.55 g. These results are better than those observed by Pedro *et al.* [25] with the tadpoles of *Calyptocephalella gayi*. That could be explained by the different protein rates among feed. Indeed, Pedro and collaborators used a feed with a low protein level (40.11%) whereas that used in our study is Coppens which is a high protein level (56%). In addition, these results are lower to those obtained by *Rana catesbeiana* [26, 27]. That could result from the difference among experiments duration. Indeed, our experiment lasted 24 days that is very less than 15 months of theirs. Mean Specific Growth Rates (SGR) recorded in all treatments ( $6.772 \pm 1.009$ ;  $8.253 \pm 1.704$ ;  $9.059 \pm 0.209$ ;  $9.027 \pm 0.142$ ;  $8.814 \pm 0.747$  and  $8.999 \pm 0.076$  %/day) are better than those obtained in *Rana catesbeiana* [28, 29, 27] and *alyptocephalella gayi* [25]. These results could be explained by the fact that the growth of aquatic animals (fish, frogs...) depends on the quantity and quality of the feed consumed. Besides, the increase in Specific Growth Rate (SGR) is synonym of feed availability till a maximum consumption threshold [30]. The increase of tadpoles' height is subject to optimal feed consumption enabling remarkable changes in morphology, physiology and behavior, during the metamorphosis process [31, 32]. This life stage makes difference among frogs and other aquatic vertebrates and affects the nutritional experiments due to the animal weight loss during the metamorphosis [33]. In addition, metamorphosis in tadpoles could be achieved unless they satisfy nutrients needs that enable them to be transformed in a new animal totally different. This dependence on nutritive elements was also observed by Martínez *et al.* [34] who led an experiment on *Rana perezi*, though inadequate feeding elongates the larval stage. Otherwise, tadpoles in natural media are subject to food competition. If this resource is scarce, a part of animals satisfy their nutritional need that leads to food selection in tadpoles that metamorphose during a certain season. Remaining tadpoles stay in the medium until they meet *al.* the requirements for metamorphosis. In nutrition studies, evaluation of nutritive values index are important. Mean values of Feed conversion Rate in the current study are lower than those obtained by Flores-Nava *et al.* [29], and those in *R. catesbeiana* tadpoles [26]. That shows a low feed conversion by *H. occipitalis* tadpoles. These differences could result from the used feed quality (40.11% crude protein and 10.22% of lipid) that is less rich than Coppens (56% of protein and 15% of lipid) and also the difference in species. Besides, these feed conversion rates were similar to those of certain carnivorous fishes (0.93 – 3.23) in *S. intermedius* larvae [35] and *H. longifilis* larvae [36]. The influence of the different feeding rates on the tadpoles' carcass composition justify the significant differences observed among dry matter, crude ash, crude lipids and crude proteins rates of *H. occipitalis* tadpoles ( $p < 0.05$ ). Tadpoles fed on the 10 % ration showed values nearby those in the initial sample. These results are similar to those of Cagiltay *et al.* [8], who led a study on the chemical composition of *Rana ridibunda* in rearing. Nevertheless, these values are upper than those recorded by Ozogul *et al.* [37] in *Rana esculenta* and Cagiltay *et al.* [8] in *Rana ridibunda* from Adana, Bursa and Trakya regions in Turkey. That could result not only from the difference species but from the used feed quality. Moreover, Shearer *et al.* [38] explained that moisture, proteins, fats and ash rates in aquatic animals tend to vary according to species, climate, seasons, sexual maturity and food diet. Our crude proteins values are lower than those observed by Tokur *et al.* [9] in *Rana esculenta* while lipids values are upper.

Concerning crude ash values, our results are similar to those of Tokur *et al.* [9]. This difference between crude proteins and lipids values could due to the different proportions of bones and flesh in species. By the same way, the proportions of proteins in tadpoles flesh can slightly be tied to feed and climate [9].

The determination of appropriate feed quantity to animals is capital because it determines nutrients and energy availability in the subject. In the current study, the optimal ration for *H. occipitalis* tadpoles rearing is 6.02 %/day of the body weight while the maximum ration is 21.37%/day. These results are nearby those in *Rana catesbeiana* ranging from 8 to 14 % of body weight [26]. They are also in accordance with those recorded by Fontanello *et al.* [39] and Justo *et al.* [40], ranging from 3% to 13% of body weight in *Rana catesbeiana* tadpoles.

## 5. Conclusion

Globally, the current study showed that in order to obtain the best yield in *H. occipitalis* tadpoles growth, it worth an optimal ration of 6.02 % of biomass. This feeding rate will enable a rapid biomass growth, specific growth and a good feed conversion rate in tadpoles.

## 6. Acknowledgements

The authors thank World Bank through African Center of Excellence on Climate, Biodiversity and Sustainable Agriculture (CEA-CCBAD) for financial support for the scholarship granted to Théophile GODOME.

## 7. References

1. Morin PJ. Predation, competition, and the composition of larval anuran guilds. *Ecol Monogr.* 1983; 53:119-138.
2. Wilbur HM. Complex life cycles and community organization in amphibians. In: Price PW, Slobodchikoff CN, Gaed WS (eds) *A new ecology: novel approaches to interactive systems.* Wiley New York. 1984, 195-224.
3. Wilbur HM. Regulation of structure in complex systems: experimental temporary pond communities. *Ecology.* 1987; 68:1437-52.
4. Hairston NG Sr. *Ecological experiments: purpose, design, and execution.* Cambridge University Press, Cambridge, 1989.
5. Resetarits WJJr, Bernardo J. *Experimental ecology: issues and perspectives.* Oxford University Press, Oxford, 1998.
6. Bourgeois PA. L'importance des amphibiens pour la conservation des petits fragments forestiers dans les forêts tropicales humides. Mémoire de maîtrise en biologie incluant un cheminement de type cours en écologie internationale de l'Université de Sherbrooke, Québec, Canada. 2008, 153.
7. Martínez IP, Rafael À, Herràez MP. Growth and metamorphosis of *Rana perezi* larvae in culture: effects of larval density. *Aquaculture.* 1996; 142:163-170.
8. Cagiltay F, Erkan N, SelCuk A, Ozden O, Devrim TD, Ulusoy S *et al.* Chemical composition of the frog legs (*Rana ridibunda*). *Fleischwirtschaft International.* 2014; 26(5):78-81.
9. Tokur B, Gürbüz D, Özyurt G. Nutricional composition of frog (*Rana esculenta*) waste meal. *Bioresource Technology.* 2008; 99(6):1332-1338.
10. Altig R, Whiles MR, Taylor CL. What do tadpoles really eat? Assessing the trophic status of an understudied and

- imperiled group of consumers in freshwater habitats. *Freshwater Biology*. 2007; 52:386-395.
11. Al-Hafedh YS, Ali SA. Effects of feeding on survival, cannibalism, growth and feed conversion of African catfish, *Clarias gariepinus* (Burchell) in concrete tanks. *Journal of Applied Ichthyology*. 2004; 20:225-227.
  12. Culley DD, Sotiariadis PK. Progress and problems associated with bullfrog tadpole diets and nutrition. In: T. Meedham and B. Thomas (Editors), *Nutrition of Captive Wild Animals*, 3rd Annual Dr. Scholl Conf. Lincoln Park Zoological Society, Chicago. 1984, 123-148.
  13. Wu G, Saoud IP, Miller C, Davis DA. The effect of feeding regimen on mixedsize pondgrown channel catfish, *Ictalurus punctatus*. *Journal of Applied Aquaculture*. 2004; 15:115-125.
  14. Seenapa D, Devaraj KV. Effects of feeding levels on food utilization and growth of Carla fry. In: De Silva SS (ed) *Fish Nutrition Research in Asia* proceedings on the fourth. Asian Fish Nutrition workshop. Asian Fisheries Society Special Publication. 1991; 5:49-54.
  15. Rodriguez-Serna M, Flores-Nava A, Olvera-Novoa MA, Carmona-Osalde C. Growth and Production of Bullfrog *Rana catesbeiana* Shaw, 1802, at Three Stocking Densities in a Vertical Intensive Culture System. *Aquacultural Engineering*. 1995; 15(4):233-242.
  16. Chinathamby K, Reina RD, Bailey PCE, Lees BK. Effects of salinity on the survival, growth and development of tadpoles of the brown tree frog, *Litoria ewingii*. *Australian Journal of Zoology*. 2006; 54:97-105.
  17. Jon L. Growth and Development of Larval *Rana temporaria*: Local Variation and Countergradient Selection. *Journal of Herpetology*. 2003; 37(3):595-602.
  18. Castano B, Miely S, Smith GR, Rettig JE. Interactive effects of food availability and temperature on wood frog (*Rana sylvatica*) tadpoles. *Herpetological Journal*. 2010; 20:209-211.
  19. AOAC. (Association of Official Analytical Chemists). *Official methods of analysis of the Association of Official Analytical Chemists*, 19th edition. AOAC, Arlington, Virginia, USA, 2012.
  20. Brett JR, Groves TDD. *Fish physiology*. *Physiological Energetics*. 1979; 3:280-282.
  21. Petersen AM, Gleeson TT. Acclimation temperature affects the metabolic response of amphibians skeletal muscle to insulin. *Comparative Biochemistry and Physiology*. 2011; 160(3):72-80.
  22. Borges FF, Stéfani MV, Amaral LA. Quality of the effluents of bullfrog tadpole ponds, *Boletim do Instituto de Pesca*, São Paulo. 2014; 40(3):409-417.
  23. Ferreira CM, Pimenta AGC, Paivaneto JS. Introdução à ricultura. *Boletim Técnico do Instituto de Pesca*. 2002; 33:1-15.
  24. Pereira MM, Mansano CFM, Peruzzi NJ, Stéfani MV. Nutrient deposition in bullfrogs during the fattening phase. *Boletim do Instituto de Pesca*, São Paulo. 2015; 41(2):305-318.
  25. Pedro HT, Ricardo S, Maria TV. Formulated diets for giant Chilean frog *Calyptocephalella gayi* tadpoles. *Ciencia e Investigación Agraria*. 2014; 41(1):13-20.
  26. Praptee S, Pornchai L, Maleeya K, Suchart EU. Effects of diets with various protein concentrations on growth, survival and metamorphosis of *Rana tigerina* and *Rana catesbeiana*. *Journal of the Science Society of Thailand*. 1997; 23:209-224.
  27. Carmona-Osalde C, Olvera-Novoa MA, Rodriguez-Serna M, Flores-Nava A. Estimation of the protein requirement for bullfrog (*Rana catesbeiana*) tadpoles, and its effect on metamorphosis ratio. *Aquaculture*. 1996; 141:223-231.
  28. Munguia-Fragozo VP, Alatorre-Jacome O, Aguirre-Becerra H, García-Trejo FJ, Soto-Zarazúa GM, Rico-García E. Growth and Metabolic Effects of Stocking Density in Bullfrog Tadpoles (*Rana catesbeiana*) under Greenhouse Conditions. *International Journal of Agriculture and Biology*. 2015; 17(4).
  29. Flores-Nava A, Olvera-Novoa AM, Gasca-Leyva E. A comparison of the effects of three water-circulation regimes on the aquaculture of Bullfrog (*Rana catesbeiana* Shaw) tadpoles. *Aquaculture*. 1994; 128:105-114.
  30. Peres H, Oliva-Teles A. Protein and energy metabolism of European seabass (*Dicentrarchus labrax*) juveniles and estimation of maintenance requirement. *Fish Physiology and Biochemistry*. 2005; 31:23-31.
  31. Pikington JB, Simkiss K. The mobilization of the calcium carbonate deposits in the endolymphatic sacs of metamorphosing frogs. *Journal of Experimental Biology*. 1966; 45:329-341.
  32. Duellman WE, Trueb L. *Biology of Amphibians*. McGraw-Hill, NY, 670 pp. Flores-Nava, A., Olvera-Novoa, M. and Gasca-Leyva, E., 1994. A comparison on the effects of three water-circulation regimes on the aquaculture of bullfrogs (*Rana catesbeiana* Shaw) tadpoles. *Aquaculture*. 1986; 128:105-114.
  33. Wilbur HM. Interactions of food level and population density in *Rana sylvatica*. *Ecology*. 1977; 58:206-209.
  34. Martinez IP, Herriez MP, Alvarez R. Response of hatchery-reared *Rana perezii* larvae fed different diets. *Aquaculture*. 1994; 128:235-244.
  35. Tossavi CE, Ouattara NI, Fiogbe ED. First breeding trials of African catfish, *Schilbe intermedius* (Rüppel, 1832): Influence of feeding rate on growth, survival and body composition of larvae reared in circular tanks. *African Journal of Aquatic Science*. 2016; AJAS-0196.
  36. Atsé BC, Sylla S, Konan KJ, N'Dri KSA. Effets des forts taux de ration alimentaire et des fréquences de nourrissage sur la croissance et le survie des larves du silure africain *Heterobranchus longifilis* Valenciennes, 1840. *Livestock Research for Rural Development*. 2013; 25(9).
  37. Ozogul F, Ozogul Y, Olgunoglu I, Kuley EB. Comparison of fatty acid, mineral and proximate composition of body and legs of edible frog (*Rana esculenta*). *International Journal of Food Science and Nutrition*. 2008; 59(7, 8):558-565.
  38. Shearer KD, Asgard T, Andorsdottir G, Aas GH. Whole body elemental and proximate composition of Atlantic salmon (*Salmo salar*) during the life cycle. *Journal of Fish Biology*. 1994; 44:785-797.
  39. Fontanello D, Arruda-Soares H, Mandelii JRJ, Reis JM. Desenvolvimento pondérial de girinos de Ra-touro (*Rana catesbeiana*, Shaw 1802), criados com racao de diferentes niveis proteicos. *Boletim del instituto de Pesca*. 1982; 9:125-129.
  40. Justo CL, Penteado LA, Fontanello D, Arruda-Soares H, Mandelii JRJ, Campos BES. Ganho de peso de girinos de *Rana catesbeiana* Shaw, 1802, em criacao intensiva, sob diferents densidades populacionais. *Boletim del instituto de Pesca*. 1985; 12:31-37.