



International Journal of Fisheries and Aquatic Studies

E-ISSN: 2347-5129
P-ISSN: 2394-0506
(ICV-Poland) Impact Value: 5.62
(GIF) Impact Factor: 0.549
IJFAS 2017; 5(5): 340-345
© 2017 IJFAS
www.fisheriesjournal.com
Received: 07-07-2017
Accepted: 08-08-2017

Richard Adande
Laboratory of Research on
Wetlands (URZH), Faculty of
Sciences and Technology,
Department of zoology,
University of Abomey-calavi
(UAC), B.P.526 Cotonou, Benin

Mouhamadou Nourou Dine Liady
Laboratory of Research on
Wetlands (URZH), Faculty of
Sciences and Technology,
Department of zoology,
University of Abomey-calavi
(UAC), B.P.526 Cotonou, Benin

Comlan Ephrem Tossavi
Laboratory of Research on
Wetlands (URZH), Faculty of
Sciences and Technology,
Department of zoology,
University of Abomey-calavi
(UAC), B.P.526 Cotonou, Benin

Emile Didier Fiogbe
Laboratory of Research on
Wetlands (URZH), Faculty of
Sciences and Technology,
Department of zoology,
University of Abomey-calavi
(UAC), B.P.526 Cotonou, Benin

Correspondence
Emile Didier Fiogbe
Laboratory of Research on
Wetlands (URZH), Faculty of
Sciences and Technology,
Department of zoology,
University of Abomey-calavi
(UAC), B.P.526 Cotonou, Benin

The effect of zooplankton produced using rabbit manure and artemia on the survival and growth of larvae of *Clarias gariepinus*, *Heterobranchus longifilis* and heteroclaris

Richard Adande, Mouhamadou Nourou Dine Liady, Comlan Ephrem Tossavi and Emile Didier Fiogbe

Abstract

Plurispesific fresh water zooplankton produced with rabbit manure, and dry artemia in powder (artemia Shell free) were used to evaluate survival and growth performances of larvae of *Clarias gariepinus*, *Heterobranchus longifilis* and heteroclaris with initial weights are respectively 2.5 ± 0.0 , 2 ± 0.14 , 2.7 ± 0.1 mg for 12 days. Six experimental diets were tested in out of ponds by triplicate as follow: Czoo (*Clarias gariepinus* fed with fresh water zooplankton), Cart (*Clarias gariepinus* fed with artemia), Hzoo (*Heterobranchus longifilis* fed with fresh water zooplankton), Hart (*Heterobranchus longifilis* fed with artemia), HCzoo (heteroclaris fed with fresh water zooplankton) and HCart (heteroclaris fed with artemia). At the end of rearing Czoo, Cart, Hzoo, Hart, HCzoo and HCart gave respectively survival rates of 74 and 73.5%, 63.5 and 60% and 70 and 62%. Final mean weights recorded were 15.54 ± 0.37 , 13.3 ± 0.97 , 34.13 ± 2.31 , 36.46 ± 1.16 , 18.14 ± 0.71 and 16.25 ± 0.52 mg for Czoo, Cart, Hzoo, Hart, HCzoo and HCart with specific growth rates respectively of 15.22 ± 0.20 , 13.91 ± 0.60 , 24 ± 1.18 , 24.56 ± 0.40 and 15.87 ± 0.11 , 14.95 ± 0.45 (%.day⁻¹). Results from the effect of local plurispesific fresh water zooplankton and artemia on the survival and growth of larvae of *Clarias gariepinus* and *Heterobranchus longifilis* and their hybrids (heteroclaris) indicate local fresh water zooplankton can successfully replace artemia in the first life stage of larvae of these two African catfish species and their hybrids.

Keywords: *Clarias gariepinus*, *Heterobranchus longifilis*, heteroclaris, Artemia, plurispesific zooplankton, survival and growth.

Introduction

Current development of aquaculture in West Africa focus essentially on certain species such as Nile Tilapia (*Oreochromis niloticus*) and African catfishes (*Clarias gariepinus* and *Heterobranchus longifilis*)^[1, 2]. In most of African countries, fish farming encounters many difficulties despite the presence of traditional production systems of catfish (claridae) though the demand is high^[3]. Indeed, the non-availability of fries and juveniles of claridae constitutes a choc to fish farming development^[4-8]. In addition, larval rearing (determinative stage) always doesn't lead to expected results due to insufficient feeding and non-adaptation of feed to first stage of development^[9, 3]. Moreover, the passing of larvae from endogenous feeding (vitellin sack) to an exogenous feeding with the appearance of digestive organs constitutes an important stage to be considered in fish life^[10, 11]. Currently, the mastery of larval rearing is necessary for the production of fries in order to satisfy demand about these two catfish species. Nowadays, the major difficulty of larvae rearing is the choice of best quality of feeding for fresh water fish larvae^[12]. It will be better to use suitable feed for larvae not only to optimize the survival and growth but to enable reduction of production cost^[13]. Certain authors^[14, 15] present artemia like the ideal feed for fresh water fish larval rearing. Although its availability, accessibility and cost constitute serious problems for fish farming in developing countries. To remedy this,^[3, 16-19] have shown in their studies the use of the live feed produced from organic fertilizers of animal origin (poultry droppings, cow dung, pig dejection) use of artemia for larviculture. By the same way, rabbit manure is rich in mineral elements able to provoke the production of fresh water zooplankton and phytoplankton for larvae feeding^[20]. Unfortunately to this day, the effect of zooplankton produced with rabbit manure on the survival and growth

of larvae of claridae has never been studied. We were glad to try larval rearing based on local plurispecific zooplankton produced with rabbit manure in order to promote catfish farming in rural areas. Therefore, in this study, we propose to compare the effect of zooplankton produced with rabbit manures and artemia on the survival and growth of larvae of *Clarias gariepinus*, *Heterobranchus longifilis* and heteroclaris.

Materials and methods

Origin of larvae and installation

Larvae of *C. gariepinus*, *H. longifilis* and heteroclaris were obtained from artificial reproduction on the station of research on the diversification of fish farming of the Laboratory of Research on Wetlands (LRZH) of the University of Abomey-Calavi (UAC). The experiments were carried out from 22 June to 4 July 2017. Two specimens of *C. gariepinus* (1 kg weight per individual) and *H. longifilis* (a female of 3 kg and a male of 1.5 kg) were used in artificial reproduction. Larvae of heteroclaris were obtained from *C. gariepinus* ♀ X *H. longifilis* ♂ weighting respectively 0.5 and 1.2 kg. To provoke spawning, an extract of ovaprim (0.5 and 0.25 mL/kg of body weight respectively for the female and the male) was used for oocytes and spermatozooids maturing [21, 22]. Ten hours (10 hours) after injection, eggs were stripped and fertilized with milt obtained from male. Fertilized eggs were hatched with baskets in hatchery tanks and in ponds at free air. Eggs hatching happened 27 hours after hatchery at $26.02 \pm 0.91^\circ\text{C}$. Two days after hatching, larvae were distributed in circular ponds (50 cm radius x 8 cm height) in triplicate and fed for 12 days [9]. An initial density of 200, 45, 100 larvae respectively for *C. gariepinus* (initial weight 2.5 ± 0.00 mg), *H. longifilis* (initial weight 2 ± 0.04 mg) and heteroclaris (initial weight 2.5 ± 0.06 mg) were constituted in six triplicate as follow:

C_{zoo}: *Clarias gariepinus* fed with fresh water zooplankton,

C_{art}: *Clarias gariepinus* fed with artemia,

H_{zoo}: *Heterobranchus longifilis* fed with fresh water zooplankton,

H_{art}: *Heterobranchus longifilis* fed with artemia,

HC_{zoo}: heteroclaris fed with fresh water zooplankton,

HC_{art}: heteroclaris fed with artemia.

Larvae were fed three times daily such 7 a.m., noon and 7 p.m. [2].

Production of fresh water zooplankton

Fresh water zooplankton was produced in plastic buckets of 80 liters capacity containing 40 liters of drilling water fertilized with 600 g/m³ of rabbit manure (optimal dose). Three days after fertilizing, 10 liters of earthen pond of polyculture (*Clarias* and *Tilapia*) were filtered carefully using plankton net of 25 µm in order to eliminate zooplankton and then completed to the 40 liters of fertilized water (phytoplankton inoculation). Seventy-two hours after phytoplankton seeding, zooplankton were seeded with an initial density of 20 ind/L. Zooplankton seeded were made of 4 ind/L of nauplii of copepods, 5 ind/L of adult's copepods, 3 ind/L of cladocerans and 8 ind/L of rotifers.

The artemia used

Dry artemia powder (artemia Shell free) is a marine crustacean canned by Aqua-Lush (China, NAFADAC, REG, N°A9-0275) and sold in Beninese markets. According to manufacturer indications, it is made of 58% of protein, 5.8% of ash, 6.0% moisture, 2.2% of fiber and 16% of fat.

Physico-chemical quality of water and rearing ponds maintenance

In order to maintain good rearing conditions, dead fishes were removed, weighted and counted every morning in each pond. Wastes were siphoned before fishes feeding. Temperature, pH, conductivity and dissolved oxygen were controlled daily and *in situ* with a multi-parameter device CALYPSO (Version Soft/2015, SN-ODEOA 2138) at $\pm 0.1^\circ\text{C}$ close to mg/L.

Larvae survival and growth

The monitoring of growth was carried out every three days on 200, 45 and 100 larvae respectively of *C. gariepinus*, *H. longifilis* and heteroclaris. At each monitoring process, half part of individuals was caught randomly with a net. The biomass of individuals was taken waterless with a scale (sensitivity 0.01 mg). Dead specimens were counted daily in each pond to evaluate larvae survival.

Recorded data were used to determine the following zootechnical parameters: Mean weight gain (MWG = (final biomass - initial biomass) / rearing duration); Specific Growth Rate (SGR = [ln (final mean weight) - ln (initial mean weight) / rearing duration] x 100); Survival Rate (SR = final fish number / initial fish number) x 100 and Production Index (mg/j) (PI = Survival Rate x [final weight - initial weight / rearing duration]) [23, 24].

Statistical analysis

Data recorded were analyzed with STATISTICA software (Statsoft inc., Tulsa, OK). Different means were compared by using a one way analysis of variance (ANOVA). All significant levels were fixed at $p < 0.05$. In case of need, significance of differences among means was tested using the test LSD of Fisher.

Results

Despite the acid pH level during experiment period, temperature, conductivity, dissolved oxygen, salinity and TDS in different rearing ponds fluctuated weakly and are mentioned in Table 1.

Table 1: physico-chemical parameters of the water.

Parameters	Variations	Mean and Range
T (°C)	27.30 - 26.02	26.02±0.91
DO (mg/L)	6.70 - 7.40	7.40±0.49
Conductivity (µs/cm)	183.60 - 184.80	184.80±0.85
pH	6.00 - 6.90	6.90±0.64
Salinity (mg/L)	0.10 - 0.11	0.11±0.01
TDS	95.34 - 94.41	94.41±0.66

Zootechnical performances of fish larvae

Zootechnical parameters of different species and hybrids are mentioned in Table 2 with a significant difference ($F_{(25,31,22)} = 14.33$, $p < 0.05$). The highest survival rates of both two species and hybrids were recorded in C_{zoo} (74%) followed by HC_{zoo} (70%) with a significant difference ($F_{(5,12)} = 191.47$, $p < 0.05$). In return, there is no significant difference between C_{zoo} (74%) and C_{art} (73.5%) on the one hand and H_{zoo} (60.3%) and H_{art} (60%) on the other hand ($p > 0.05$). Concerning hybrids, the survival rate of HC_{zoo} (70%) is higher than HC_{art} (62%) with a significant difference ($P < 0.05$). So, the survival rates (63.5-74%) of larvae of these two species and their hybrids fed with plurispecific fresh water zooplankton produced by using rabbit manure are comparable to those fed with artemia (60-73.5%). The mean weight gain (MWG) of C_{zoo} ($1.08 \pm$

0.03 mg/day) is higher than C_{art} (0.90 ± 0.08 mg/day) with a significant difference ($p < 0.05$). Besides, the mean weight gain (MWG) of H_{art} (2.87 ± 0.86 mg/day) is higher than H_{zoo} (2.28 ± 0.20 mg/day) with a significant difference ($p < 0.05$). Also, the highest PI was recorded in H_{art} with a significant difference ($F_{(5,12)} = 45.80, p < 0.05$). In return, there is no difference between the PI of H_{zoo} and H_{art} ($P > 0.05$). The same

trend was observed in hybrids. But, the PI of C_{art} was less than C_{zoo} with a significant difference ($p < 0.05$). The mean weight of C_{zoo} was higher than C_{art} at the beginning of the third day (fig 2). Those of larvae H_{art} was slightly higher than larvae H_{zoo} (fig 3). Among hybrids, the weight of HC_{art} was higher till the ninth day from which HC_{zoo} became slightly higher (fig. 4).

Table 2: Zootechnical parameters of larvae of *C. gariepinus*, *H. longifilis* and heteroclarias.

Parameters	C_{zoo}	C_{art}	H_{zoo}	H_{art}	HC_{zoo}	HC_{art}
Ni	600	600	135	135	300	300
Nf	451	441	90	82	210	187
SR (%)	74a	73,5a	63,5be	60b	70b	62ac
IMW (mg)	$2.5 \pm 0.00a$	$2.5 \pm 0.00a$	$2 \pm 0.14a$	$2 \pm 0.14a$	$2.7 \pm 0.1a$	$2.7 \pm 0.1a$
FMW (mg)	$15.54 \pm 0.37b$	$13.3 \pm 0.97a$	$34.13 \pm 2.31e$	$36.46 \pm 1.16f$	$18.14 \pm 0.71cd$	$16.25 \pm 0.52bc$
MWG/day (mg/day)	$1.08 \pm 0.03a$	$0.90 \pm 0.08b$	$2.28 \pm 0.20a$	$2.87 \pm 0.86b$	$1.28 \pm 0.05a$	$1.12 \pm 0.04b$
SGR (%/day)	$15.22 \pm 0.20a$	$13.91 \pm 0.60b$	$24 \pm 1.18c$	$24.56 \pm 0.40c$	$15.87 \pm 0.11b$	$14.95 \pm 0.45ab$
PI (mg/day)	$80.41 \pm 4.42b$	$66.15 \pm 3.75a$	$170.02 \pm 28.10c$	$172.25 \pm 10.30c$	$90.06 \pm 6.01b$	$70.00 \pm 2.19ab$

^{ab} values bearing same letters on the same line between difference species and hybrids are not significantly different at $p < 0.05$.

Ni = initial number of larvae, Nf = final number of larvae, SR = survival rate; IMW= initial mean weight, FMW = final

mean weight, Mean weight gain (MWG), SGR = specific growth rate, PI = production index.

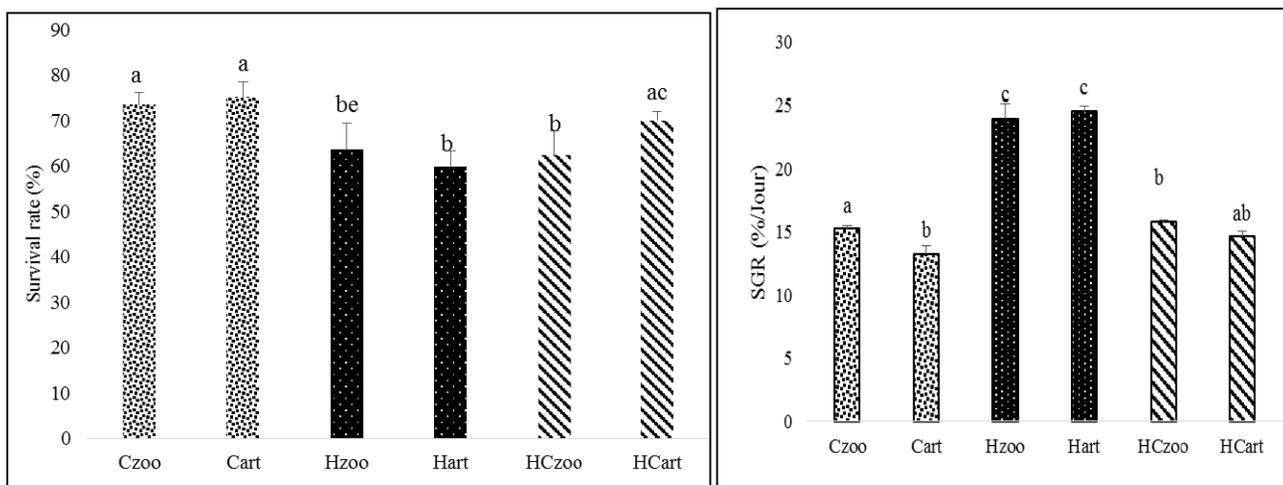


Fig 1: Effect of zooplankton and artemia on the survival and growth of larvae of *C. gariepinus*, *H. longifilis* and heteroclarias for 12 days.

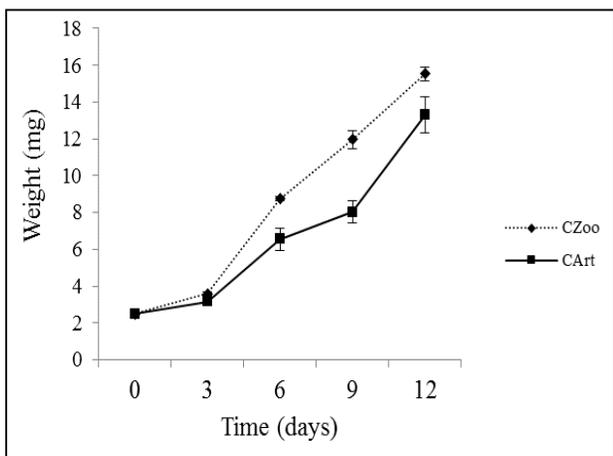


Fig 2: weight progression of *C. gariepinus* larvae fed for 12 days with zooplankton and artemia.

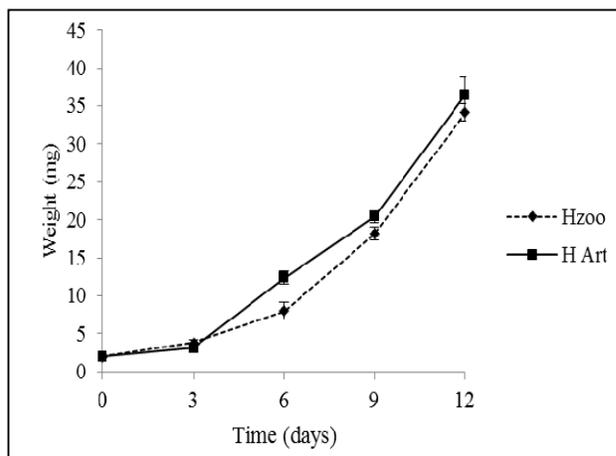


Fig 3: Weight progression of *H. longifilis* larvae fed for 12 days with zooplankton and artemia.

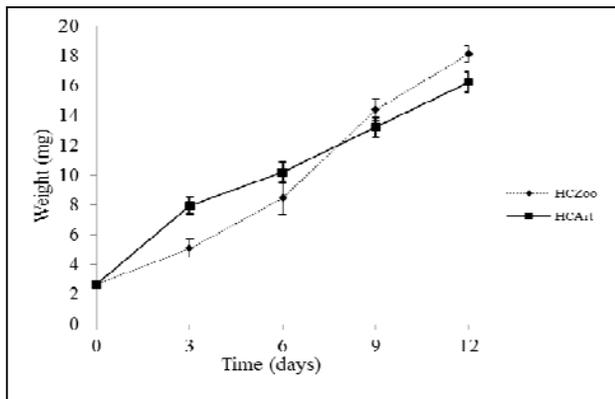


Fig 4: Weight progression of heteroclaris larvae fed for 12 days with zooplankton and artemia.

Discussion

Average values of physico-chemical parameters of the water were in range of those in larval rearing of African Catfish *C. gariepinus* and *H. longifilis* despite constant pH value (6.90 ± 0.64) [25, 26, 3, 18, 2]. The highest survival rate (74%) recorded in *C. gariepinus* larvae fed with plurispecific fresh water zooplankton in pond is less than the 97% obtained by Agadjihouèdé *et al.* [19] in larvae of *C. gariepinus* fed with fresh water zooplankton and the 99.7% recorded by Awaïss *et al.* [13] in larvae of *C. gariepinus* fed with *Brachionus calyciflorus* produced in mono-specific culture. In return, survival rates of 60-74% recorded in this study are in the range of 71-76% obtained respectively by Otémé and Gilles [5] and Atsé *et al.* [27] in larvae of these two African catfish and their hybrids fed with zooplankton. Indeed, the main reasons of survival rate decrease during larval rearing of *C. gariepinus* and *H. longifilis* are either tied to cannibalism or territorial aggression [28]. By the same way, these results are similar to those obtained by Smith *et al.*, Folkvord *et al.* and Coulibaly *et al.* [29-31] who recorded that mortality would be due to the presence of initial weak larvae. Besides, in the present study, this mortality might be also due to the acidity of drilling water pH (6 - 6.90). Similar observations were made by Tossavi *et al.* [32] during the rearing of *Schilbe intermedius* fries in the same production systems. The high mortality rate recorded in *H. longifilis* larvae fed with artemia could be explained by larvae fragility during the first stages of development contrary to *C. gariepinus* larvae. Indeed, according to Keremath *et al.* [33], that is the reason of the crossing of these two species.

For 12 days, larvae of *C. gariepinus*, *H. longifilis* and hybrids fed with zooplankton achieved respectively 15.54 ± 0.37 ; 36.46 ± 1.16 and 18.14 ± 0.71 mg as mean final weight. *H. longifilis* is a species with more rapid growth than *C. gariepinus* [5, 34]. In addition, according to Otémé *et al.* [5] and Atsé *et al.* [2], food availability (zooplankton) is a determinative factor to growth and height homogeneity of *H. longifilis* larvae thus, they would weight more than *C. gariepinus*. Specific growth rates (SGR) recorded were higher than the 8.9% obtained by Okunsebor *et al.* [15] in larvae of *C. gariepinus* fed with *Brachionus calyciflorus*, *Ceriodaphnia reticulata* and artemia. By the same way, the SGR recorded in these two species and hybrids (Table 2) are comparable to those obtained in larvae of *Clarias macrocephalus* [35], *Labeo parvus* [11], *H. longifilis* [2], *C. gariepinus* [36] and hybrids [37] fed with dry feed, artemia and live food (fresh water zooplankton). Indeed, that shows the performance of live food used (zooplankton). Although, SGR recorded in hybrids was

largely higher to the 0.78%/D obtained in hybrids (*H. longifilis* ♂ x ♀ *C. gariepinus*) by Keremath *et al.* [33]. According to Ndimele *et al.* [38], specific growth rate would be certainly tied to the type of crossing. Good growth performances and survival recorded in larvae of *H. longifilis*, *C. gariepinus* and hybrids fed with zooplankton in the present study were influenced by the quality of live food. Our results are similar to prior studies that showed the performances were due to good digestibility of live food [39, 2]. Besides, according to Lauff *et al.* [40], the live food (zooplankton) contributes in 80% to the digestion due to the quality and the quantity of protease it contains. Therefore, zooplankton remains the best live food for fish larvae contrary to artificial feed [41-44], what justify good survival performances and growth recorded in the present study. Moreover, the production index (PI) indicates the optimal production performance compared to the survival rate, density, availability and food quality. PI of H_{zoo} and H_{art} respectively 170.02 ± 28.10 and 172.25 ± 10.30 corroborate those 136.28 and 166.28 mg/day recorded by Atsé *et al.* [2] in larvae of *H. longifilis* fed three times daily with artemia. These could be due to good growth performances of *H. longifilis* larvae.

Conclusion

Results from the present study showed that larvae of *C. gariepinus*, *H. longifilis*, and hybrids fed with plurispecific fresh water zooplankton produced using rabbit manure gave good survival rate and growth. Larvae fed with zooplankton gave survival rate ranging 63.5 - 74% and growth rate 15.22 ± 0.20 ; 24 ± 1.18 and $15.87 \pm 0.11\% \cdot \text{day}^{-1}$ respectively for larvae of *C. gariepinus*, *H. longifilis*, and hybrids. Thus, plurispecific fresh water zooplankton can freely replace artemia during larval rearing of these catfish and their hybrids. It would be important to future studies to determine the influence of the biochemical composition of zooplankton on survival and growth parameters of these larvae and to evaluate these performances with fresh water macroinvertebrates produced by using the same organic fertilizer.

References

- Omitogun OG, Olaniyan OF, Oyeleye OO, Ojiokpota C, Aladele SE, Odofin WT. Potentials of short term and long term cryopreserved sperm of African giant catfish (*Clarias gariepinus* Burchell 1822) for aquaculture. African Journal of Biotechnology. 2010; 9(41):6973-6982.
- Atsé BC, Sylla S, Konan KJ, N'Dri KSA. Effets des forts taux de ration alimentaire et des fréquences de nourrissage sur la croissance et la survie des larves du silure africain *Heterobranchus longifilis* Valenciennes, 1840. Livestock Research for Rural Development. 2013; 25(9):201.
- Rukéra Tabaro S, Micha JC, Ducarme C. Adaptation Experiment of Massive Production of *Clarias Gariepinus* Juveniles in Rural Conditions. Tropicicultura. 2005; 23(4):231-4.
- Uys W, Hecht T. Evaluation and preparation of an optimal dry feed for the primary nursing of *Clarias gariepinus* larvae (Pisces, Clariidae). Aquaculture. 1985; 47:173-184.
- Otémé ZJ, Gilles S. Elevage larvaire du silure africain *Heterobranchus longifilis* : évaluation quantitative des besoins en proies vivantes des larves. Aquatic Living

- Resources. 1995; 8:351-354.
6. Legendre M, Teugels GG. Développement et tolérance à la température des oeufs de *Heterobranchus longifilis*, et comparaison des développements larvaires de *H. longifilis* et de *Clarias gariepinus* (Teleostei, Clariidae). Aquatic Living Resources 1991; 4:227-240.
 7. Hecht T, Oellennann L, Verheust L. Perspectives on clariid catfish culture in Africa. Aquatic Living Resources, 9 Hors-série, 1996, 197-206.
 8. Nguenga D. A comparison of the reproductive performance and aquaculture potential of two strains of an African catfish, *Heterobranchus longifilis* Valenciennes, 1840 (Teleostei, Clariidae) in Cameroon. PhD thesis, Catholic University of Leuven (KUL), The Netherlands, 1997, 155.
 9. Ducarme Ch, Micha JC. Technique de production intensive du poisson chat africain, *Clarias gariepinus* Tropicultura. 2003; 21(4):189-198.
 10. Wang C, Xie S, Zheng K, Zhu X, Lei W. Effects of live food and formulated diets on survival, growth and protein content of first feeding larvae of *Pelteobagrus fulvidraco*. Journal of Applied Ichthyology. 2005; 21:210-214.
 11. Montchowui E. Study of the biology of reproduction and the artificial reproduction of one species of fish cyprinidae of Oueme River of Benin: *Labeo parvus* (Boulenger, 1902), PhD thesis, University of Liège, 2009, 152.
 12. Awaïss A, Kestemont P. Feeding sequences (rotifère and dry diet), larval, growth and biochemical composition of African catfish *Clarias gariepinus* (pisces : claridae) larvae. Aquaculture research. 1998; 29:731-741.
 13. Wu G, Saoud IP, Miller C, Davis DA. The effect of feeding regimen on mixed-size pond-grown channel catfish, *Ictalurus punctatus*. Journal of Applied Aquaculture. 2004; 15:115-125.
 14. Awaïss A, Kestemont P, Micha JC. Etude du premier alevinage du poisson-chat africain, *Clarias gariepinus* (Birchell, 1822), avec le rotifère d'eau douce, *Brachionus calyciflorus* (Pallas). EAS Spec Publ. 1993; 18:443-453.
 15. Okunsebor SA, Sotolu AO. Growth performance and survival rate of *clarias gariepinus* fry fed on live feeds *brachionus calyciflorus*, *ceriodaphnia reticulata* and shell free artemia. www.patnsukjournal.net/currentissue. 2011; 7(2):108-115.
 16. Ekelemu JK, Nwabueze AA. Comparative studies on zooplankton production using different types of organic manure. International Journal of Sciences and Nature. 2011; 2(1):140-143.
 17. Akodogbo HH, Bonou CA, Adandé R, Sossou DS, Fiogbé ED. Optimization of zooplankton production from pig dung optimal dose : renewed medium. Agricultural Advances. 2015; 4(2):15-21.
 18. Adewumi AA. Growth performance and survival of *Clarias gariepinus* hatchlings fed different starter diets. European Journal of Experimental Biology. 2015; 5(3):1-5.
 19. Agadjihouédé H, Bonou CA, Avocèvou-Ayisso C, Lalèyè PA. Production Cost of *Heterobranchus longifilis* (Valenciennes, 1840) Fingerlings in the Ponds Fertilized with Poultry Droppings. Journal of Food Science and Engineering. 2017; 7:93-98.
 20. Adande R, Bokossa HKJ, Liady MND, Fiogbe ED. Valorization of various sources of rabbit manure in agropiscicultural system in Benin (West Africa): dynamics and effect of mineralization upon quality of fresh water. International Journal of Recycling Organic Agriculture in Waste. 2017; 6:233-243.
 21. Micha JC. Synthèse des essais de reproduction, d'alevinage et de production chez un silure africain : *clarias lazera* val. Bulletin Français de Pisciculture. 1975, 256.
 22. Legendre M, Teugels GG, Cauty C, Jalabert B. Comparative study on morphology, growth rate and reproduction of *Clarias gariepinus* (Burchell, 1822), *Heterobranchus longifilis Valenciennes, 1840*, and their reciprocal hybrids (Pisces, Clariidae). Journal of Fish Biology. 1992; 40:59-79.
 23. Zacharia S, Kakati VS. Growth and survival of *Penaeus merguensis* post larvae at different salinities. Israeli Journal of Aquaculture Bamidgeh. 2002; 54:157-162.
 24. Mohanty RK. Density-dependent growth performance of Indian major carps in rainwater reservoirs. Journal of Applied Ichthyology. 2004; 20:123-127.
 25. Hem S, Legendre M, Trebaol L, Cissé A, Otémé Z, Moreau Y. L'aquaculture lagunaire. In : Les milieux lagunaires, Tome II : Environnement et ressources aquatiques de Côte d'Ivoire. (Eds, J R Durand, P Dufour, D Guiral, and S G F Zabi). ORSTOM, Paris, France, 1994, 455-505.
 26. Gilles S, Dugue R, Slembrouk J. Manuel de production du silure africain *Heterobranchus longifilis*. Moissonneuve et Larose Paris, 2001.
 27. Atsé BC, Konan KJ, Kouassi NJ. Biologie de la reproduction du Cichlidae *Tylochromis jentinki* dans la lagune Ebrié (Côte d'Ivoire). Cybium. 2009; 33(1):11-19.
 28. Hecht T, Appelbaum A. Notes on the growth of the Israeli sharptooth catfish *Clarias gariepinus* during the primary nursing phase. Aquaculture. 1987; 63:195-204.
 29. Smith C, Reay P. Cannibalism in teleost fish. Rev. Fish Biol. 1991; 1(1):41-64.
 30. Folkvord A, Ottera H. Effects of initial size distribution, day length, and feeding frequency on growth, survival, and cannibalism in juvenile Atlantic cod (*Gadus morhua* L.). Aquaculture. 1993; 114(3-4):243-260.
 31. Coulibaly A, Ouattara IN, Koné T, N'Douba V, Snoeks J, Gooré Bi G *et al.* First results of floating cage culture of the African catfish *Heterobranchus longifilis Valenciennes, 1840*: Effect of stocking density on survival and growth rates. Aquaculture. 2007; 263:61-67.
 32. Tossavi EC, N'tcha A, Djissou A, Kpogue D, Ouattara IN, Fiogbe ED. Feeding rate requirements for *Schilbe intermedius* (Rüppel, 1832) fingerlings reared in captivity. International Journal of Agronomy and Agricultural Research. 2015; 7(6):34-41.
 33. Keremah RI, Deekae SN, Akalokwu UA. Growth and Feed utilization of Catfish Hybrid (*Heterobranchus longifilis* ♂ x ♀ *Clarias gariepinus*) Fingerlings fed practical diet Greener Journal of Agricultural Sciences. 2013; 3(4):286-290.
 34. Agadjihouédé H, Chikou A, Bonou CA, Lalèyè PA. survival and growth of *Clarias gariepinus* and *Heterobranchus longifilis* larvae fed with freshwater zooplankton. Journal of agricultural science and technology. 2012; 2:192-197.
 35. Evangelista AD, Fortes NR, Santiago CB. Comparison of some live organisms and artificial diet as feed for Asian

- catfish *Clarias macrocephalus* (Günther) larvae. Journal of Applied Ichthyology. 2005; 21:437-443.
36. Okunsebor SA, Shima KG, Sunnuvu TF. Effect of Water Volume on Growth, Survival Rate and Condition Factor of *Clarias gariepinus* Hatchlings Cultured in Different Enclosures. American Journal of Experimental Agriculture. 2015; 7(3):150-154.
 37. Adewolu MA, Akintola SL, Akinwunmi OO. Growth performance and survival of hybrid African catfish larvae (*Clarias gariepinus* x *Heterobranchus bidorsalis*) fed on different diets. The zoologist. 2009; 7:45-51
 38. Ndimele PE, Owodeinde FG. Comparative Reproductive and Growth Performance of *Clarias gariepinus* (Burchell, 1822) and Its Hybrid Induced with Synthetic Hormone and Pituitary Gland of *Clarias gariepinus*. Turkish Journal of Fisheries and Aquatic Sciences. 2012; 12:619-626.
 39. Kumar D, Marimuthu K, Haniffa MA, Sethuramalingam TA. Effect of Different Live Feed on Growth and Survival of Striped Murrel *Channa striatus* larvae. Journal of Fisheries & Aquatic Sciences. 2008; 25(2):105-110
 40. Lauff M, Hofer R. Development of proteolytic enzymes in fish and the importance of dietary enzymes. Aquaculture. 1984; 37:335-346.
 41. Dabrowski K, Charlon N, Bergot Pand Kaushik S. Rearing coregonid (*Coregonis schinzi* palea Cuv et Val) larvae using dry and live food. I. Preliminary data. Aquaculture. 1984; 411:11-20.
 42. Dave G. Experiences with waste water cultured *Daphnia* in the start feeding of rainbow trout (*Salmo gairdneri*). Aquaculture. 1989; 79:337-343.
 43. Villegas CT, Lumasag GL. Biological evaluation of frozen zooplankton as food for milkfish. Journal Applied Ichthyology. 1991; 7:65-71.
 44. Adeyemo AA, Oladosu GA, Ayinla AO. Growth and survival of fry of African catfish species, *Clarias gariepinus* Burchell, *Heterobranchus bidorsalis* Geoffery and *Heteroclarias* reared on *Moina dubia* in comparison with other first feed sources. Elsevier Science B.V., Amsterdam Aquaculture. 1994; 119:41-45.