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Effect of chlorpyrifos on protein and carbohydrate content of *Heteropneustes fossilis* (Bloch, 1794)

Sneha Verma and Anurag Rawat

Abstract

Organophosphorus pesticides are used in the agricultural area due to their rapid biodegradability and non-persistent nature to control pests but their broad spectrum of harmful effects extends far beyond the pest. The objective of this research was to investigate the effect of sub-lethal chlorpyrifos effect on ovarian protein and carbohydrate content of the freshwater catfish "Heteropneustes fossilis". LC₁₀, LC₂₅ and LC₅₀ value of chlorpyrifos were calculated for 24, 48, 72 and 96 hr by probit analysis (Finney, 1971). The LC₁₀, LC₂₅ and LC₅₀ value for 24 hr were 9.79, 21.59 and 51.95 μ l/l, and 3.3, 7.54 and 18.91 μ l/l for 48 hr, and for 72 hr: 0.85, 2.39 and7.47 μ l/l. The 96 hr LC₁₀, LC₂₅ and LC₅₀ value were 0.12, 0.55 and 4.64 μ l/l respectively. The result showed that CPF significantly (p<0.05) decreased the ovarian protein and carbohydrate concentration. Therefore, it may be concluded that CPF is highly toxic to aquatic organism.

Keywords: Chlorpyrifos, Heteropneustes fossilis, protein, carbohydrate

1. Introduction

Pesticides are commonly used in agricultural activities. Because of drift, atmospheric transport, agricultural and residential runoff, individual misuse, and improper disposal, pesticides are also found in aquatic habitats [1, 2] where it act as a toxicant for aquatic organisms. These toxicants produce several biochemical and physiological responses. Thus, it is important that toxic effects be determined and interpreted in biochemical terms.

Chlorpyrifos (CPF), O, O-diethyl-O-(3,5,6-trichloro-2-pyridyl) phosphorothioate is an organophosphorus pesticide used extensively for the management of domestic and agricultural pests. It impairs toxic effects on aquatic non-target organisms [3]. CPF are powerful neurotoxic compound as they inhibit acetylcholinesterase (AChE) [4-9]. A number of reports are investigated the toxicity of CPF in different aquatic models viz., *Vibrio fischeri, Thamnocephalus platyurus* and *Daphnia magna* [3], *Gambusia affinis* [10], *Channa punctatus* [11], *Oreochromis mossambicus* [12], aquatic invertebrates and fish [13], yellow perch and rainbow trout [14], *Labeo rohita* [15]. Biochemical constituents are serving as potential biomarkers for a variety of different organisms [16]. The pesticide effect on biochemical properties has been studied by many workers [17, 21].

Fish acts as bio-indicators of aquatic contaminants [22]. Fish play an important and significant role in the food chain, thus, study of pesticide effect on fish has a diagnostic significance in evaluation of adverse effect of pesticides to human health [23]. *Heteropneustes* fossilis is a valuable high priced freshwater food fish and therefore often chronically exposed to insecticides. Several investigation were reported on the toxicity of various pesticides to fish, viz. Shukla *et al.* [24], Sahai *et al.* [25], Kumari [26] and Kumari [27]. The aim of the present study was to investigate the impact of chlorpyrifos on ovarian biochemical parameters of *H. fossilis*.

2. Materials and Methods

2.1 Chemicals

All the chemicals and reagents used in this study were of analytical grade and were purchased locally. The test compound chlorpyrifos was purchase from local market (Hindustan insecticide limited, India) as trade name Hilban®.

2.2 Animal collection and their maintenance

An experiment was performed in accordance with local/national guidelines of ethical committee for experimentation in animals.

The fresh water *H. fossilis* of relatively same size and weight (160±10gm) were collected from commercial fisherman of Lucknow, Uttar Pradesh, India. After formal quarantine treatment with 0.05% KMnO₄ solutions, fish were acclimatized in laboratory conditions under normal photoperiod (12:12 light: dark) and temperature (22±2°C) for three weeks in 120 L glass aquaria containing water. The physico-chemical characteristics were analyzed by APHA ^[28] (Table 1). Water was renewed daily to remove faecal matter and waste metabolite of fish during acclimatization. During this period, fish were fed regularly with commercial fish food pellets and goat liver.

 Table 1: The physico-chemical characteristics of water used for experiment

| S. No. | Water parameters | Mean Values | |
|--------|-----------------------------|-------------|--|
| 1 | pН | 7.6-7.9 | |
| 2 | Temperature (°C) | 23.5-26.5 | |
| 3 | DO (mg/l/) | 6.3-7.5 | |
| 4 | Free CO ₂ (mg/l) | 36-38 | |
| 5 | Total hardness (mg/l) | 215-220 | |
| 6 | Alkalinity (mg/l) | 184-190 | |

2.3 Acute toxicity and biochemical studies

Pilot experiments were conducted to determine the LC_{10} , LC_{25} and LC_{50} concentrations for acute toxicity. Fish were exposed to different concentrations of chlorpyrifos ranged from 0.01, 0.05, 0.1, 0.5, 1, 5, 10, 15, 20 and $25\mu l/L$ for 24, 48, 72 and 96 hr in groups of 10 individuals in five replicates. During the whole process a control group (without toxicant) was maintained side by side. The lethal concentration values were obtained and analyzed by Finney's probit analysis method [29]. The fish were not fed on the day before the beginning of the experiment.

2.4 Biochemical studies

Fish were exposed to different concentrations of CPF (0.01 to $25\mu l/l$) in batches of ten with five replicates. Control groups of fish were maintained concurrently. Water was renewed daily with CPF concentration up to 96 hr of study period. After 96 hr study, fish from each group were removed and sacrificed for ovarian tissue sampling for protein and carbohydrate analysis study. The total protein was determined by Lowry's *et al.* [30] methods and estimation of total carbohydrates by Roe [31] method and respectively.

2.5. Statistical analysis

The data was subjected to one-way analysis of variance (ANOVA) and the significance difference was set up at p<0.05. These values were expressed as Mean \pm SD.

3. Results and Discussion

The catfish *H. fossilis* showed varied degrees of mortality with different concentration of CPF. The mortality% was significantly increased in concentration and duration dependent manner of CPF (F=52.27, p<0.05). The calculated LC₁₀ LC₂₅ and LC₅₀ values were shown in table 2.

The protein and carbohydrate concentration of ovary tissue was varied with the concentrations of pesticide, chlorpyrifos. Control fish showed maximum protein and carbohydrate content in ovary. As the concentration of CPF was increased, the protein and carbohydrate content was decreased significantly (F=5415, F=19203; p<0.05). The minimum protein and carbohydrate concentration were noticed in the fish tissue exposed to highest concentration of CPF i.e., 25µl/l and maximum content were observed in lowest concentration of CPF (0.01 µl/l) after 96 h of exposure (Fig. 1). Shinde et al. [32] have also been reported the decrease of protein content in ovary of Notopterus notopteus treated with heavy metals. Same findings were observed in other fish viz., Colisa fasciatus [33], Catla catla [34], Labeo rohita [35, 36], Cyprinis carpio [37]. Tilak [38] reported reduction of protein content of liver, brain and ovary of *C. punctatus* exposed to fenvalerate. CPF cause hyper activity which results in breakdown of food reserves to meet the energy demand [39]. Total protein content decreased due to CPF exposure which suggests the possible breakdown of proteins in the fabrication of some amount of energy for organism [40]. Neff [41] stated that under toxicant exposure, animal required more energy hence protein level was depleted. Proteins are mainly involved in the architecture of the cell which is chief source of nitrogenous metabolism and during toxicant stress they are also source of energy [42]. It serves as a source of energy for fish and it will be used when insufficient energy is available from other sources like carbohydrate. It regulates the process of interaction between intra and extra cellular media. The carbohydrate comprised mainly glycogen and total free sugars and the fluctuations in this content may be due to accumulation and utilization of glycogen and total free sugars at different phases of life like growth, gametogenesis and spawning. In fish, generally the carbohydrate reserves may be rapidly utilized under unfavorable conditions like under toxicant exposure. The decrease in total carbohydrate signifies its utility possibly to meet the higher energy demands of fish reeling under pesticide stress [43].

Table 2: Estimation LC₁₀, LC₂₅ and LC₅₀ values along with 95% lower and upper confidence limits of *H. fossilis* exposed to CPF for 24, 48, 72 and 96 hr.

| Probit value µl/L) | 24 h | 48 h | 72 h | 96 h |
|--------------------|---------------|---------------|--------------|---------------|
| LC ₁₀ | 9.79 | 3.3 | 0.85 | 0.12 |
| LC10 | (5.37-14.31) | (1.13-5.84) | (0.16-1.95 | (0.02 - 0.59) |
| LC25 | 21.59 | 7.54 | 2.39 | 0.55 |
| LC25 | (14.91-27.73) | (3.74-11.27) | (0.78-4.34) | (0.05-2.84) |
| I.C., | 51.98 | 18.91 | 7.47 | 4.64 |
| LC50 | (42.79-62.62) | (13.06-25.25) | (4.04-11.69) | (1.82-9.02) |

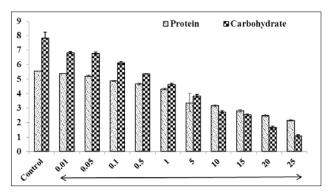


Fig 1: Effect of Chlorpyrifos on protein (mg/ml/100mg tissue wt) and carbohydrate content (mg/ml/100mg tissue wt) of freshwater catfish, *H. fossilis*.

4. Conclusion

Frequent use of this pesticide in paddy fields and its disposal in water bodies is very hazardous and unwarranted to the fresh water ecosystem. On the basis of above mentioned findings, it may be suggested that indiscriminate use of this pesticides in water bodies should not be encouraged by the farmers particularly those who are planning for farming catfishes in their paddy fields.

5. References

- McConnell LL, LeNoir JS, Datta S, Seiber JN. Wet deposition of current use pesticides in the Sierra Nevada mountain range, California, USA. Environ. Toxicol. Chem. 1998; 17:1908-16.
- Gilliom RJ, Hamilton PA. Pesticides in the nation's streams and ground water, 1992–2001. A summary fact sheet. Reston: US Geological Survey. 2006.
- 3. Palma P, Palma VL, Fernandez RM, Soares AM, Barbosa IR. Acute toxicity of atrazine, endosulfan and chlorpyrifos to *Vibrio fischeri*, *Thamnocephalus platyurus* and *Daphnia magna* relative to their concentrations in surface water from the Alentejo Region of Portugal. Bull. Environ. Contam. Toxicol. 2008; 81:485-9.
- 4. Casida JE. Esterase inhibitors as pesticides. Sci. N. Y. 1964, 146:1011.
- Coppage DL, Mathews E. Brain-acetylcholinesterase inhibition in a marine teleost during lethal and sublethal exposure to 1,2-dibromo 2,2-dimethyl phosphate (Naled) in sea water. Toxicol. Appl. Pharmacol. 1975; 31:128.
- Kabeer Ahamed SI, Rao KVR. Correlation between subacute toxicity of malathion and acetylcholinesterase inhibition in the tissues of the teleost *Tilapia mossambica*. Bull. Environ. Contam. Toxicol. 1980; 24(5):711.
- 7. Klaverkamp JF, Hobden BR. Brain-acetylcholinesterase inhibition and hepatic activation of acephate and fenitrothion in rainbow trout (*Salmo gairdneri*). Can. J. Fish. Aquat. Sci. 1980; 32:1450.
- 8. Lieske CN, Clark JH, Meyer HG, Lowe JR. Spontaneous and induced reactivation of eel acetylcholinesterase inhibited by three organophosphates. Pestic. Biochem. Physiol. 1980; 13:205.
- Rath S, Mishra BN. Toxicological effect of dichlorovos (DDVP) on brain and liver acetylcholinesterase (AChE) activity of *Tilapia mossambica*. Toxicology. 1981; 19:239
- 10. Rao JV, Begum G, Pallela R, Usman PK, Rao RN.

- Changes in behavior and brain acetylcholinesterase activity in mosquito fish, *Gambusia affinis* in response to the sub-lethal exposure to chlorpyrifos. Int J Environ Res Public Health. 2005; 2:478-83
- 11. Ali D, Nagpure NS, Kumar S, Kumar R, Kushwaha B. Genotoxicity assessment of acute exposure of chlorpyrifos to freshwater fish *Channa punctatus* (Bloch) using micronucleus assay and alkaline single-cell electrophoresis. Chemosphere. 2008; 71:1823-31.
- 12. Rao JV. Brain acetylcholinesterase activity as a potential biomarker for the rapid assessment of chlorpyrifos toxicity in a euryhaline fish *Oreochromis mossambicus*. Environ. Bioindic. 2008; 3:11-22.
- 13. Anderson RL, Defoe DL. Toxicity and bioaccumulation of endrin and methoxychlorine in aquatic invertebrates and fish. Environ. Pollut. 1980; 22:111.
- 14. Guiney PD, Peterson RE. Distribution and elimination of polychlorinated biphenyl after acute dietary exposure in yellow perch and rainbow trout. Arch. Environ. Contam. Toxicol. 1980; 9:191.
- 15. Tilak KS, Veeraiah K, Ramana Kumari GV. Biochemical changes induced in freshwater fish *Labeo rohita* (Hamilton) exposed to pesticide mixture. Asian J. of Microbiol. Biotech. & Eng. Sci. 2001; 3(4):315-319.
- 16. Agrahari S, Pandey KC, Gopal K. Biochemical alteration induced by monocrotophos in the blood plasma of fish, *Channa punctatus* (Bloch). Pestic. Biochem. Physiol. 2007; 88:268-72.
- 17. Wild D. Mutagenicity studies on organophosphorus insecticides. Mutat. Res. 1975; 32:135-150.
- Shanmugam A. Fundamentals of Biochemistry for Medical Students. College of Medical Science, Madras. 1977, 48.
- 19. Ganesan RM, Jebakumar SRD, Jayaraman. Sublethal effect of organochlorine insecticides on protein, carbohydrate and lipid content in liver tissues of *Oreochromis mossambicus*. Proc. Ind. Acad. Sci. Anim. Sci. 1980; 98(1):51-55.
- 20. Panigrahi AK, Mishra BN. Toxicological effects of a sublethal concentration of inorganic mercury on freshwater fish *Tilapia mossambica* Peters. Arch. Toxicol. 1980; 44(4):269-278.
- 21. Murty AS, Devi P. The effect of endosulfan and its isomers on tissue proteins, glycogen and lipids in the fish *Channa punctatus*. Pestic. Biochem. Physiol. 1982; 17:280-286.
- Gupta A, Rai DK, Pandey RS, Sharma B. Analysis of some heavy metals in the riverine water, sediments and fish from river Ganges at Allahabad. Environ. Monit. Assess. 2009; 157:449-58.
- 23. Begun G, Vijayaraghavan S. Alterations in protein metabolism of muscle tissue in the fish *Clarius batrachus* (Linn) by commercial grade dimethoate. Bull. Environ. Contam. Toxicol. 1996; 57:223-228.
- 24. Shukla JP, Pandey K. Aresenic induced structural changes during the ovarian cycle of a freshwater perch, *Colisa fasciatus* (Bloch & Schneider). Bull. Inst. Zool. Academic Sinicia. 1984; 23(1):69-74.
- 25. Sahai S, Singh S. Effect of lindane on gonadal function in *Rasbora daniconius* (Teleostei). Proc. 8th A.E.B. SES and Symp. 1987, 54-55.
- 26. Kumari M. Biocidal effect of Bagyon in relation to the yolk synthesis in freshwater catfish *H. fossilis*. Bull. Biol. Sci. 2005, 31-37.

- Kumari M. Biochemical changes induced by the pesticide Abate in the liver of catfish *H. fossilis*. J. Environ. Ecol. 2007; 255(4):1164-1165.
- 28. APHA, Standard Methods for the Examination of Water and Wastewater, 21st Centennial Edition, American Public Health Association (APHA), American Water Works Association (AWWA) and Water Environment Federation (WEF), Washington DC, USA. 2005.
- 29. Finney DJ. Probit Analysis. 2nd edition. Cambridge University Press, Cambridge, UK. 1953-1971, 333.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with folin phenol reagent. J. Biol. Chem. 1951; 193:265-275.
- 31. Roe JH. The determination of sugar in blood and spinal fluid with anthrone reagent. J. Biol. Chem. 1955; 153:373-380.
- Shinde VR, Veeresh MU, Kulkarni RS. Ovarian changes in response to heavy metals exposure to the fish *Notopterus notopterus* (Pallas). J. Environ. Biol. 2002; 23(2):137-141.
- Singh S, Singh SK, Yadav RP. Toxicological and Biochemical alterations of cypermethrin (synthetic pyrethroids) against freshwater teleost fish *Colisa* fasciatus at different season. World J. of Zool. 2010; 5(1):25-32.
- Martin P, Prasath D, Arivoli S. Biochemical study of freshwater fish *Catla catla* with reference to mercury chloride. Iran. J. Environ. Health Sci. Eng. 2008; 5(2):109-116.
- Waghmare SY, Wani GP. Effect of polo (an insecticide) on protein contents of liver and gonads of freshwater fish, *Labeo rohita*. Int. Res. J. Pharm. 2014; 5(10):792-794.
- 36. Patil VK, David M. Hepatotoxic potential of malathion in the freshwater teleost, *Labeo rohita* (Hamilton). Veterinarski Archiv. 2009; 79(2):179-188.
- 37. Ramesh M, Saravanan M. Haematological and biochemical responses in a freshwater fish *Cyprinis carpio* exposed to chlorpyrifos. Inter. J. Integrative. Biol. 2008; 3(1):80-83.
- 38. Tilak KS, Rao DK. Chlorpyrifos toxicity of freshwater fish. J. Aqua. Biol. 2003; 8(2):161-166.
- 39. Umminger BL. Physiological studies on super cooled Hilli fish *Fundulu heteroditus*, carbohydrate metabolism and survival at sub zero temperature. J Exp Zool. 1977; 173:159-174.
- 40. Tulsi G, Jayantha RK. Effect of chromium on protein metabolism in different tissues of fish, *Cyprinus carpio*. Res. J Pharm Biol Chem Sci. 2013; 4(1):143-148.
- Neff JM. Use of biochemical measurement to detect pollutant mediated damage to fish. In: Cardwel RD, Purdy R, Bahner RC, Eds. Aquatic Toxicology and Hazard Assessment Philadelphia: Americal Society for Testing Materials. 1985, 155-181.
- 42. Mommesen TP, Walsh PJ. Biochemical and environmental perspectives on metabolism in fishes. Experientia. 1992; 48:583-593.
- Muthukumaravel K, Sivakumar K, Kumarasamy P, Govindarajan M. Studies on the toxicity of pesticide monocrotophos on the biochemical constituents of the freshwater fish *Labeo rohita*. Int. J. Cur. Bio. Biot. 2013; 2(10):20-26.