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Benzo [a] Pyrene-carcinogenic and mutagenic equivalents of *Clarias gariepinus* (Burchell, 1822) to Nigerian-crude and diesel oils

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Abstract

The toxicity and PAH of *C. gariepinus* (0.9±0.01g) to 1, 3, 6, 9, 12 and 0.0 ml/L of crude oil and diesel was determined. There was no mortality in the control but 10, 20, 50 and 60% occurred in crude oil and 10, 20, 30, 40 and 60% in diesel after 96 h exposure. The logarithmic probit lines and R^2 : $y = 1.372x + 3.724$, $R^2 = 0.997$ and $y = 1.665x + 3.694$, $R^2 = 0.998$ and 96-h LC_{50} : 9.6 and 11.0 ml/L represented toxicity values for crude oil and diesel. Total PAH in exposed fish ranged from the highest value of 99.68±4.81 crude oil > 97.30± 14.57 diesel oil ng/μL the lowest value of 0.061 Nap to the highest value of 384.68 in DahA for crude oil and Diesel oil indicated lowest value of 2.22 Ace to highest value of 389.05 ng/μL DahA. Mean Σ 16PAH of lighter product showed greater toxicity than heavier products require further investigation to the pharmacokinetics of the products on fishes.

Keywords: Toxic equivalent factor, carcinogenicity, mutagen, PAH, petroleum

1. Introduction

Due to the lipophilic nature and high chemical stability of PAHs, they accumulate in the fatty tissues of fish following their uptake. Fishes are therefore good indicators of pollution in inland and coastal waters [11, 10, 6]. Two broad groups exist based on their physical and biological properties including, high molecular weight (HMW) and low molecular weight (LMW) PAHs. The HMW PAHs consists of 4-6 aromatic rings and are less readily biodegraded by indigenous microorganisms, hence can persist in the aqueous environment by bioaccumulating in aquatic organisms like fish and mussels and are more carcinogenic. The LMW PAHs consists of 2-3 aromatic rings and although less carcinogenic, also pose toxic effect to many aquatic organisms [2]. The concentrations of petroleum products toxic to aquatic organisms depend on the type and hydrocarbon constituents, as well as the species involved [14, 13]. Estimated concentrations of petroleum toxic to fish eggs and fingerlings to be 0.5-10 mg/L. Benzo (a) pyrene binds to DNA to cause cancer and is frequently used as a marker for carcinogenic disorders and may provide the basis for predicting the impact of exposures of PAH to *C. gariepinus* fingerlings [18]. BaP-TEQ (carcinogenic equivalents and BaP- MEQ (mutagenic equivalents are measure for sum of total 8 number of particulate PAHs (Σ 8PAH), having molecular weight greater than 228 grams. Σ 8PAH includes benzo (a) pyrene (BaP), benz (a) anthracene (BaA), chrysene/iso-chrysene (CHR), benz (b) fluoranthene (BbF), benzo (k) fluoranthene (BkF), indo (123-cd) pyrene (IP), Dibenz (a, h) anthracene (DahA) and benzo (ghi) pyrene (BghiP) [17, 4, 5]. The hardy nature and possession of accessory air-breathing organs enable *C. gariepinus* to tolerate adverse aquatic conditions [21]. Nonetheless, *Clarias gariepinus* fingerlings are very delicate and sensitive to aquatic pollutants including crude oil and other petroleum products. This study was undertaken to determine the comparative toxicity and levels of PAH of various Nigerian- petroleum products on *C. gariepinus* fingerlings.

2. Materials and Methods

2.1 Experimental fish and petroleum

A total of seven hundred and twenty (720) fingerlings of African catfish (mean weight 0.96 ± 0.1g) were obtained from local outskirts in Enugu Nigeria and transported to Fisheries

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Laboratory of the Department of Animal/Fisheries Science and Management, Enugu State University of Science and Technology ESUT, Enugu Nigeria. They were held in four fiber reinforced plastic (FRP) tanks, containing 320 L of de-chlorinated tap water. Aeration was provided to all tanks round the clock in order to maintain dissolved oxygen contents. Before the commencement of the study, the fish were acclimatized for two weeks and were fed with commercial fish diet composed of 40% crude protein. The faecal matter and other waste materials were siphoned off daily to reduce ammonia content in water. Petroleum (crude oil, petrol, kerosene and diesel) was obtained from Nigerian National Petroleum Cooperation Enugu. The water-soluble fraction WSF was prepared following the method of [25], which involved 20-h mixing of 10:1 clean water to petroleum with a rotator magnetic stirring rod, separated layers after resting for 12-hrs with separating flask before storing as stock solution in corked 50L plastic gallons. Ethical clearance from Enugu State University of Science and Technology Committee on Experimental Animal Care was obtained and followed.

2.2 Acute toxicity test

Toxicity of petroleum to *C. gariepinus* was carried out according to the OECD guideline for testing of chemicals No. 203 in a semi-static renewal system by using 200 L capacity glass aquaria. Thirty (30) fish per treatment were randomly exposed to 6 experimental treatments (1, 3, 6, 9, 12 and 0 ml/L of water soluble fractions WSF which served as the control of each triplicate group of petroleum product (crude oil and diesel), to determine 96h LC₅₀ using the probit analysis proposed by [7, 26] and polycyclic aromatic hydrocarbons (PAH) in exposed fish [14]. The exposure pollutant was renewed each day and was analyzed using LC-MS/MS to ensure the agreement between nominal and actual concentrations of the petroleum in the aquaria. The experiment was conducted under the natural photoperiod of

12:12 light-dark cycle. The physico-chemical parameters of the test water were analyzed daily, using standard methods [1] and were recorded (dissolved oxygen 7.50 ± 0.45 mg L⁻¹, temperature 27.75 ± 0.5 °C, pH 7.8 ± 0.13 and free carbon dioxide 4.28 ± 0.6 mg L⁻¹). The test fish of 9 and 12 ml/L in each product were sampled to determine Σ 16PAH, Σ 8PAH, TEQ and MEQ of each product. A portion of each sample using the GC-MC was taken for extraction and analysis of PAH [24, 16].

2.3 PAH extraction

The method described by [24, 12, 15] with slight modification for extraction and dosing of PAHs was employed.

The toxic equivalent factors (BaP_{TEF}) and mutagenic equivalent factors (BaP_{MEF}) relating the carcinogenic mutagenic potency of individual PAH to BaP have been used [17, 23, 5]. The BaP carcinogenic equivalent (BaP_{TEF}) and BaP mutagenic equivalent (BaP_{MEF}) for the individual PAHs was calculated: $BaP_{TEQ} = \Sigma C_i \times BaP_{TEF}$; $BaP_{MEQ} = \Sigma C_i \times BaP_{MEF}$, where C_i = concentration of PAHs.

2.4 Statistical analysis

Data obtained were expressed as standard mean \pm standard error of mean and analyzed using the statistical package SPSS 20.0 computer program (SPSS Inc. Chicago Illinois, USA).

3. Results

3.1 Mortality and Logarithmic probit line for 96-h LC₅₀ of petroleum products to *C. gariepinus*

Figure 1 gave mortality values of fish to various concentrations (0, 1, 3, 6, 9 and 12 ml/L. There was no mortality in the control but 10, 20, 50 and 60% for crude oil and 10, 20, 30, 40 and 60% for diesel occurred respectively in 1, 3, 6, 9 and 12 ml/L after 96 hour exposure. Figure 1 gave the logarithmic probit lines and R^2 : $y = 1.372x + 3.724$, $R^2 = 0.997$ and $y = 1.225x + 3.712$, $R^2 = 0.925$ and 96-h LC₅₀: 9.6 and 11.0ml/L for crude oil and diesel to *C. gariepinus*

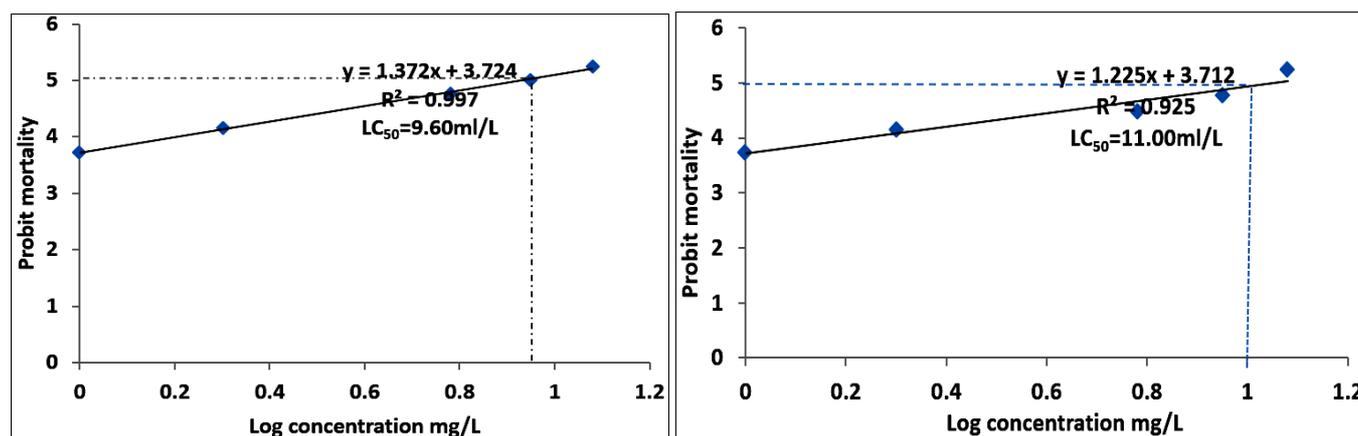


Fig 1: Logarithmic probit line for determination of 96-h LC₅₀ crude oil and Diesel to *C. gariepinus*

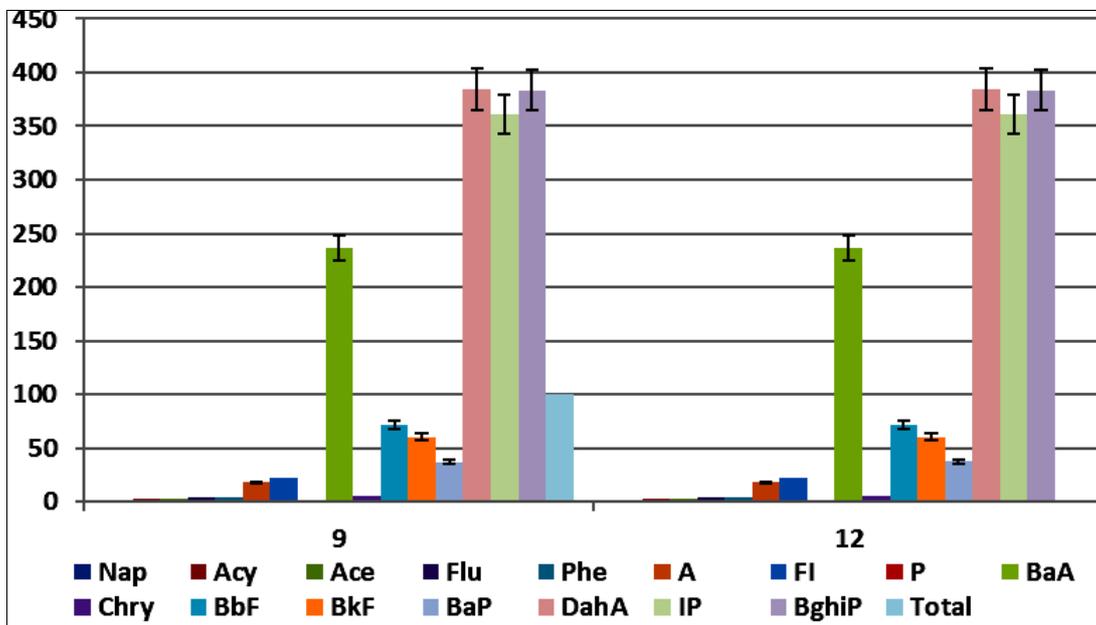
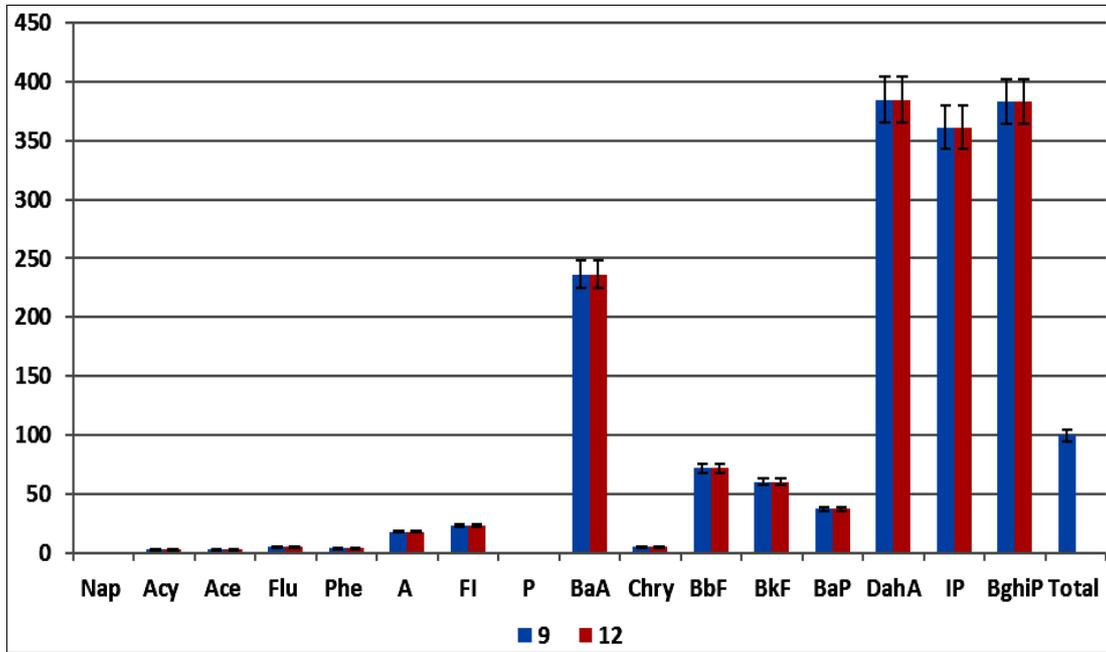
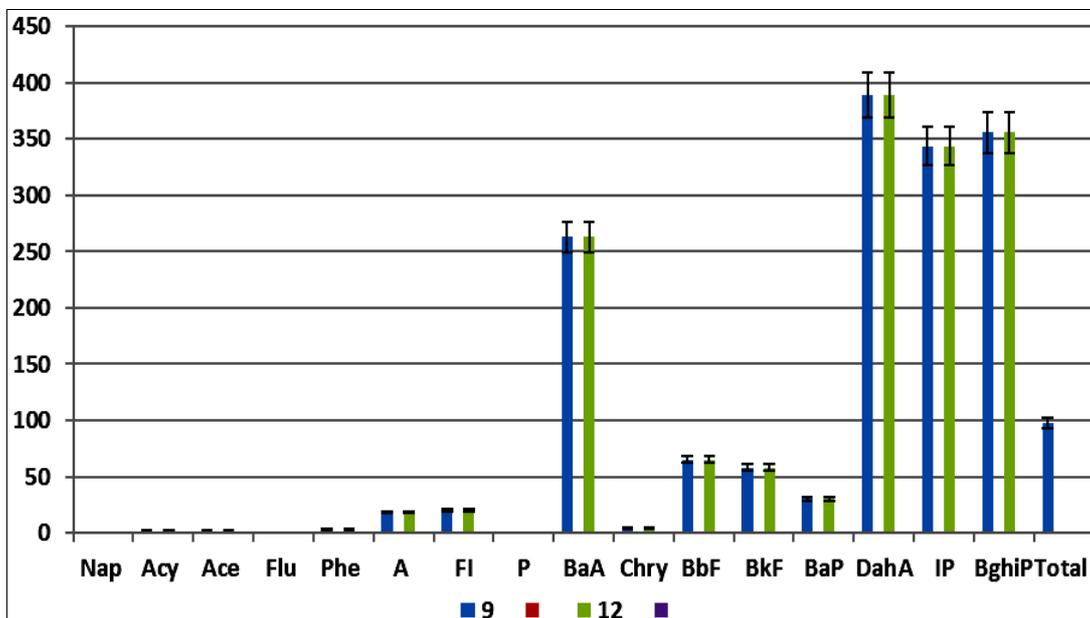


Fig 2: PAHs in fish exposed to concentrations of crude oil



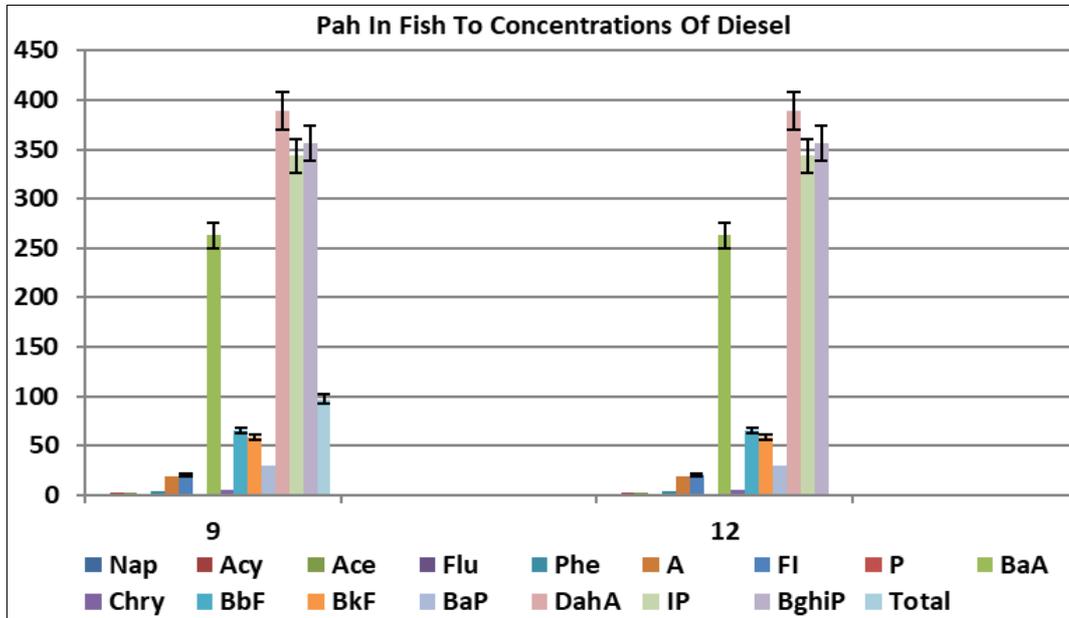


Fig 3: PAHs in crude oil and diesel

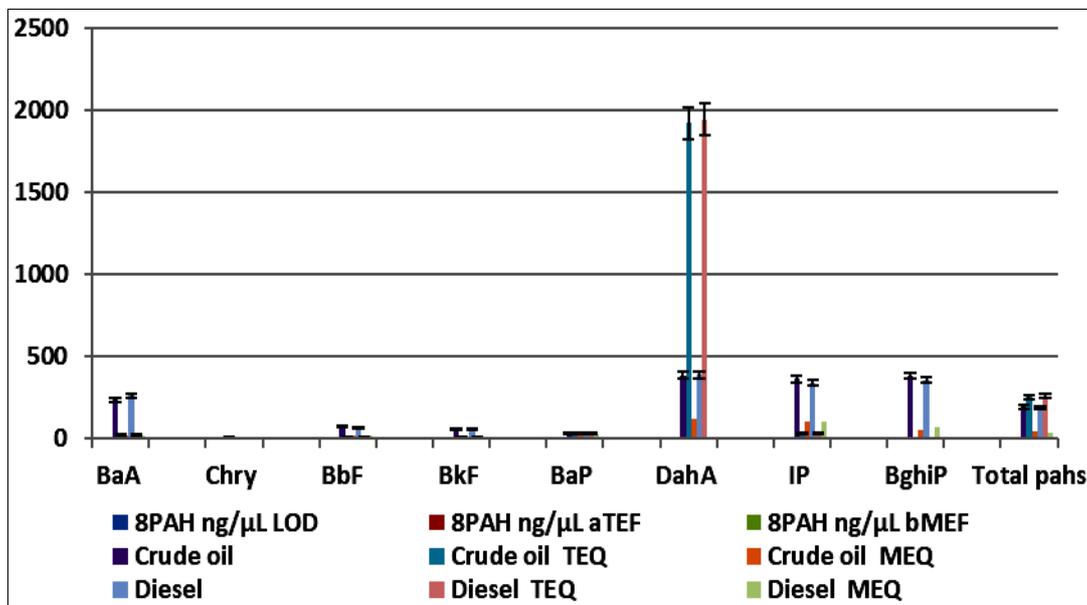
Different superscripts in a row indicate significant difference between means (ANOVA, $P < 0.05$)

KEY

cNap = Naphthalene, Acy= Acenaphthylene, Ace= Acenaphthene Flu= Fluorene, Phe=Phenathrene, A =Anthracene, FI =Fluoranthene P=Pyrene BaA=Benz {a} anthracene, Chry=Chrysene, BbF= Benzo [b] fluoranthene, BkF=Benzo [k] fluoranthene, BaP = Benzo [a] pyreneDahA = Dibenz [ah] anthracene IP = Indeno [123-cd] pyrene,, B[ghi]P = Benzo [ghi] pyrene, CO=Crude oil, P= Petrol, K= kerosene, D= Diesel, pp=petroleum products, c= concentration in ml/L, nd= not detected.

3.2 PAHs in exposed fish to petroleum

Total PAHs values in ng/μL (Figure 2) of petroleum products in exposed fish ranged from highest value of 99.68 ± 4.81 crude oil > 97.30 ± 14.57 diesel oil ng/μL The lowest value of 0.061 was shown in Nap while highest value was 384.68 in DahA >383.47 B[ghi]P > 361.38 IP >236.41BaA >71.59 BbF>60.17 BkF >37.17 BaP> 22.82FI> 17.72A >5.33Chry >4.56 Flu> 4.35 Phe>2.79 Acy>2.40Ace >0.06Nap for crude oil.but P was not detected.. Diesel oil however indicated highest value of 389.05DahA> 355.70 B[ghi]P >343.48 IP>262.80 BaA>65.35 BbF>58.16 BkF>30.38 BaP>20.09 FI>18.48 A>4.79 Chry>3.79 Phe>2.51 Acy>2.22 Ace. P, Flu and Nap were not detected.



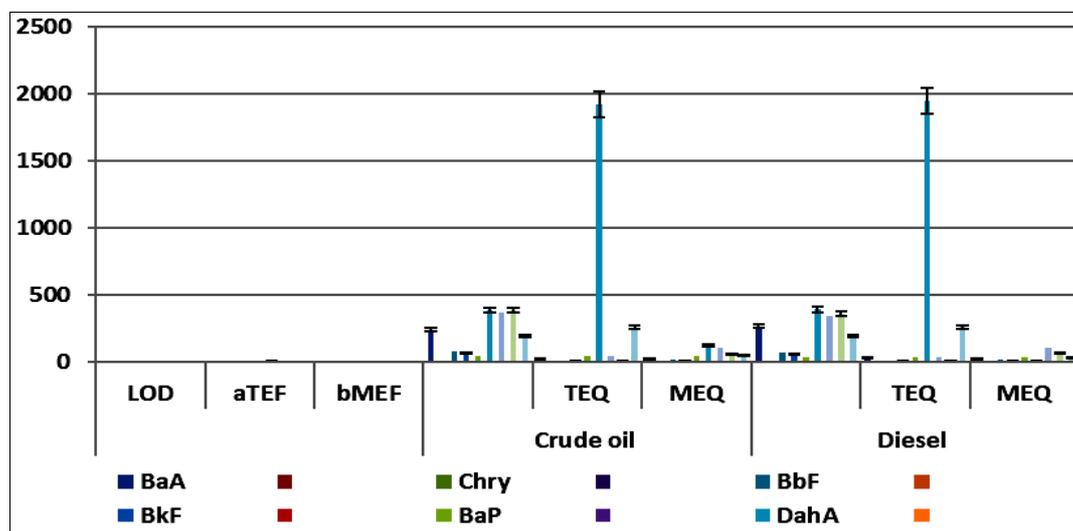


Fig 4: Bap-TEQ and Bap- MEQ of crude oil and diesel

TEF*: Toxic equivalency factors for cancer potency relative to BaP (Nisbet and LaGoy *et al.* 1992) [17].

MEF*: Mutagenic potency factor relative to BaP (Durant *et al.* 1996 and 1999) [5].

3.3 Carcinogenicity and mutagenicity equivalents

The least observed difference LOD of PAH among 8PAHs was 0.02ng/ μ L. Total TEQ and MEQ gave 254.67 and 44.84; 256.58 and 31.24 respectively for crude oil and diesel. BaP-TEQ in CO ranged from 0.05 in Chry to 1923 in DahA and 0.48 Chry-1.945 DahA respectively in Diesel. Similarly, BaP-MEQ ranged 0.09 Chry -119.25 DahA and 0.08 Chry-67.58 to BghiP in diesel.

4. Discussion

The result showed that diesel contained greater number of PAH than crude oil as well as being more carcinogenic and mutagenic, therefore was expected to be more toxic to the fish than petrol-treated ones. The contrary played out in our findings which gave the lowest value of toxicity level to be crude oil and therefore was more toxic compared to diesel. The indication that lighter petroleum products with lower total mean Σ 16PAH showed greater toxicity on exposed fish than heavier ones having higher total mean Σ 16PAH, probably due to greater level of more toxic BaA to BaP compared to less toxic but carcinogenic DahA- BaP in crude oil and DahA-BaP in diesel [2]. The impact of petroleum water soluble fraction previously under-reported has in recent times posed critical health concerns to aquatic biota, especially fish [8, 22]. The foregoing gave an indication that petroleum products with high molar mass and greater mean Σ 8PAH were more carcinogenic and mutagenic compared to lighter petroleum with lower mean Σ 8PAH. Crude and diesel oils have shown greater ability than crude oil to cause cancer and changes in the genetic makeup and may damage the genome materials or disrupt cellular metabolic processes of exposed fish to humans that consume them [9]. There is greater need for further investigation of the biochemistry exposed fish to these products since this approach may provide overestimation of cancer and mutagen potency of individual PAH, as most PAH indicated less comparative carcinogen than BaP.

Recent approaches have centered to identify and quantify PAHs in water, soil and air environment, their emission sources through various methods in order to evaluate their carcinogenic and mutagenic effects to human health [7, 19, 3].

The approaches distinguish anthropogenic multiple releases chiefly from petroleum and other sources [18, 9]. BaP is widely accepted as the indicator for measurement of carcinogenicity, thus BaP-equivalent toxicity for other carcinogenic PAHs has been recommended [28] and evaluated for cancer risk assessment [17, 5, 20, 27].

5. Conclusion

Crude oil was shown to be more toxic to the experimental fish but was less carcinogenic and mutagenic and had less number of pah. Further investigations on the biochemistry effects are required since over estimation of cancer and mutagen potencies may not be entirely ruled out.

6. References

1. APHA. American Public Health Association, American Water works Association and Water Environmental Federation). Standard Methods of Examination of water and Wastewater. 21st ed. APHA Washington DC, 2005, 20001-23710.
2. Brown J, Peake B. Sources of heavy metals and polycyclic aromatic hydrocarbons in urban storm water runoff. *Sci. Total Environ.* 2006; 359:145-155.
3. Changsheng Q, Bing L, Haisui W. Multi-pathway assessment of human health risk posed polycyclic aromatic hydrocarbon, *Environ Geochem Health.* 2015; 37:1-15.
4. Choi H, Harrison R, Komulainen H, Delgado SJ. Polycyclic aromatic hydrocarbons. WHO Guidelines for Indoor Air Quality: Selected Pollutants. Geneva: World Health Organization, 2010. Retrieved 2014-12-12.
5. Durant J, Lafleur A, Busby W *et al.* Human cell mutagenicity of oxygenated, nitrated and unsubstituted polycyclic aromatic hydrocarbons associated with urban aerosols, *Mutation Research/Genetic Toxicology and Environmental Mutagenesis.* 1996; 371:123-157
6. Ezike CO, Echor FO, Malachy NOA, Vera LM. Butrylacetlycholinesterase activities in liver and plasma, liver glycogen and plasma glucose content, haematology and behaviour of Clariid Catfish *Clarias gariepinus* to Dichlorvos *International Journal of Advanced Fisheries and Aquatic Sciences.* 2017; 3(1):90-105. Doi: <https://doi.org/10.23953/cloud/ijafas.332.crossref:23953/cloud.ijafas.332>
7. Finney DJ. Probit Analysis Canbrige University Press

- London, 1971, 23-125.
8. Hylland K. Polycyclic aromatic hydrocarbon (PAH) ecotoxicology in marine systems. *Journal of Toxicology and Environmental Health, Part A*. 2006; 69(1-2):109-123.
 9. Isioma T, Ozekeke O, Lawrence E. Human health risk assessment of polycyclic aromatic hydrocarbons (PAHs) in smoked fish species from markets in Southern Nigeria. *Toxicology Reports*. 2017; 4:56-61.
 10. Lawal AT. Polycyclic aromatic hydrocarbons, a review. *Environmental Science*. 2017; 3:1339841. <https://doi.org/10.1080/23311843.2017.1339841>.
 11. Lee RF. Metabolism of petroleum hydrocarbons in marine sediments. In: Sources, effects and sinks of hydrocarbons in aquatic environment. American Institute Biological Sciences, 1976, 333-344.
 12. Li N, Leu HK. Solid phase extraction of polycyclic hydrocarbons in surface water. *J Chromatogr. A*. 2001; 921:255-263.
 13. Li Z.H, Velisek J, Zlabek V. Chronic toxicity of verapamil on juvenile rainbow trout (*Oncorhynchus mykiss*): effects on morphological indices, hematological parameters and antioxidant responses. *J Hazard Mat*, 2001; 185:870-880.
 14. Lonning S. The effects of crude oil and oil products on marine fish larvae. *Astate*. 1977; 10:37-47.
 15. Martinez E, Gros M, Lacorte S, Barcel D. Simplified procedures for the analysis of polycyclic aromatic hydrocarbons in water sediments and mussels. *J Chromatogr A*. 2004; 1047:18-188.
 16. Neff JM. Polycyclic aromatic hydrocarbons. In: Rand G.M. and Petrocelli S.R. (Eds.). *Fundamentals of aquatic toxicology*. Hemisphere Publ. Corp. New York, 1985, 416-454.
 17. Nisbet IC, LaGoy PK. Toxic equivalency factors (TEFs) for polycyclic aromatic hydrocarbons (PAHs). *Regul. Toxicol. Pharmacol*. 1992; 16:290-300.
 18. Payne JF, Penrose WR. Induction of akyl hydrocarbon (benzo (a) Pyrene) hydroxylase in fish by petroleum. *Bull Environ. Contam. Toxicol*. 1975; 14:112-226.
 19. Peng C, Chen W, Liao X, Wang M, Zhiyun Q, Jiao W *et al*. Polycyclic aromatic hydrocarbons in urban soils of Beijing: Status, Sources, distribution and potential risk. *Environmental Pollution*. 2011; 159:802-808.
 20. Ramesh A, Archibong A, Hood DB, Guo Z, Loganathan BG. Global environmental distribution and human health effects of polycyclic aromatic hydrocarbons. *Global Contamination Trends of Persistent Organic Chemicals*. Boca Raton FL: CRC Press, 2011, 97-126.
 21. Reed WJ, Burchard Hopson AJ, Jonathan J, Ibrahim Y. *Fish and Fisheries of Northern Nigeria*. Govt. Press, London, 1967, 226.
 22. Roubal WT, Collier TK, Malins DC. Accumulation and metabolism of carbon-14 labelled benzene, naphthalene and Anthracene by young cohosalmon (*Oncorhynchus kisutch*) *Arch. Environ. Contam. Toxicol*. 1977; 5:515-529.
 23. So-Young L, Lee-Yeon I, Han-Seun S. Evaluation and chemical analysis method and determination of polycyclic aromatic hydrocarbon content in seafood and diary products. *Toxicology Reports*. 2015; 31(3):265-271.
 24. Takatsuki S, Susuki S, Sato N, Ushizawa I. Association of Official Analytica Chemists. *Toxicology*. 1985; 79(2):221-271.
 25. United Nations Environmental Programme (UNEP). Comparative toxicity of water accommodated fraction of oil and oil dispersants to marine organisms. United Nations Environmental programme. Reference Methods for Marine Pollution Studies. 1989; 43:27.
 26. United States Environmental Protection Agency (USEPA). Washington, DC, EPA/600/P-95/002F a-c Exposure Factors Handbook (1997 Final Report) Risk Assessment, 1997, <https://cfpub.epa.gov/ncea/risk/recordisplay.cfm?deid=12464>.
 27. Vo Thi LH, Nguyen Thi TH, Minora Y. Human health hazard of polycyclic aromatic hydrocarbon in road dust in Ha Noi metropolis. *Journal of Science and Technology*. 2016; 54(24):27-34.
 28. World Health organization (WHO). WHO Guidelines for indoor air quality: selected pollutants, 2010. www.euro.who.int/data/assets/pdf/0009/128169/e94535.pdf.