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## Changes in the biochemical profile of *Anabas testudineus* Bloch 1792 on exposure to aquatic toxicants of Buckingham canal, Chennai, Tamil Nadu, India

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### Abstract

Toxicants affect biologically active molecules, viz., carbohydrates, proteins and lipids. Carbohydrate, protein and lipid metabolism of fishes are disturbed under the condition of toxic stress and are known to cause increase or decrease in the biochemical constituents of the tissues. In the present study, the biochemical effects caused to *Anabas testudineus* on exposure to aquatic toxicants present in the Buckingham canal, Chennai, Tamil Nadu, India were investigated. The per cent change recorded in the tissues of experimental *Anabas testudineus* was found to be in the order of liver > kidney > muscle > intestine > gill; muscle > gill > kidney > liver > intestine; intestine > liver > kidney > muscle > gill in carbohydrate, protein and lipid respectively. The depletion in the metabolites indicated the fact that the whole metabolic pool of the fish gets disturbed/ altered under the toxic stress. Further, the change in the biochemical profile indicates their rapid utilization to provide excess energy to cellular biochemical process in order to cope with the stressed condition.

**Keywords:** *Anabas testudineus*, biochemical parameters, carbohydrate, protein, lipid

### 1. Introduction

The interaction between the contaminants and biomolecules is the first step in the generation of toxic effects. Understanding the alterations induced by the exposure to pollutants may contribute to the prediction of toxic effects that may occur at higher level of biochemical organization<sup>[1]</sup>. Toxicants affect biologically active molecules, viz., carbohydrates, proteins and lipids<sup>[2]</sup>. Carbohydrate, protein and lipid metabolism of fishes are disturbed under the condition of toxic stress<sup>[3]</sup>. Bacterial pathogens, pesticides and heavy metal pollutants are known to cause increase or decrease in the biochemical constituents of the tissues. Therefore, in the present study, the biochemical effects caused to *Anabas testudineus* on exposure to aquatic toxicants present in the Buckingham canal, Chennai, Tamil Nadu, India were investigated.

### 2. Materials and methods

#### 2.1 Study area

Chennai situated on the eastern coast of India, on 13 0 4' north latitude and 8 0 15' east longitude has three water ways that flows through the city, viz., Cooum river, Adayar river and Buckingham canal. The Buckingham canal is a man-made water canal linking the above mentioned two rivers. The canal extends from Nellore in Andhra Pradesh to Marakkanam near Puducherry. The length of this canal in Andhra Pradesh is 257km, and 163km is in Tamil Nadu. Within the city of Chennai, the canal is badly polluted from sewage and industrial effluents, and the silting up of the canal has left the water stagnant, creating an attractive habitat for mosquitoes.

#### 2.2 Test organism

Taxonomic position

Kingdom : Animalia

Phylum : Chordata

Sub Phylum : Vertebrata  
 Class : Pisces  
 Order : Anabantiformes  
 Family : Anabantidae  
 Genus : *Anabas*  
 Species : *testudineus*

*Anabas testudineus* commonly called climbing perch is an extremely hardy, small, brown or dark greenish-brown fish, native to Southeast Asia. It inhabits the majority of drainage systems across its native range and has been recorded in many different habitat-types including swamps, marshes, lakes, canals, pools, small pits, rice paddies, puddles, tributaries and main river channels. It is highly adapted to life in a seasonal tropical environment. It can tolerate very turbid and brackish water conditions and under extreme circumstances it can aestivate for several weeks by burying itself into moist ground [4].

**2.3 Biochemical studies**

Biochemical studies, viz., total carbohydrates, total proteins and total lipids were analyzed in control and experimental fish groups. Tissues of *Anabas testudineus*, viz., gill, muscle, liver, intestine and kidney from the control and experimental groups

were selected for the present study.

**3. Results**

The values of total carbohydrate, protein and lipid in the control and experimental tissues of *Anabas testudineus*, viz., gill, muscle, liver, intestine and kidney are presented in Table 1 and Figure 2. The order of carbohydrate per cent change recorded in the tissues of experimental *Anabas testudineus* was found to be liver > kidney > muscle > intestine > gill. In the tissues of control fish, the total protein content was in the order of muscle > gill > liver > intestine > kidney. The variation in distribution suggests the gradual difference in metabolic caliber of various tissues under the influence of aquatic pollutants and the total protein contents was found to decrease in all the tissues of the experimental fish. Maximum decrease was observed in muscle and gill and minimum in kidney and intestine. The per cent decrease of total protein content in pollutant affected *Anabas testudineus* was found to be in the order of muscle > gill > kidney > liver > intestine. The decreased total lipid content of tissues in pollutant affected *Anabas testudineus* was found to be in the order of intestine > liver > kidney > muscle > gill whereas the per cent change in the lipid content over the control fish was found to be in the order of gill > liver > kidney > intestine > muscle.

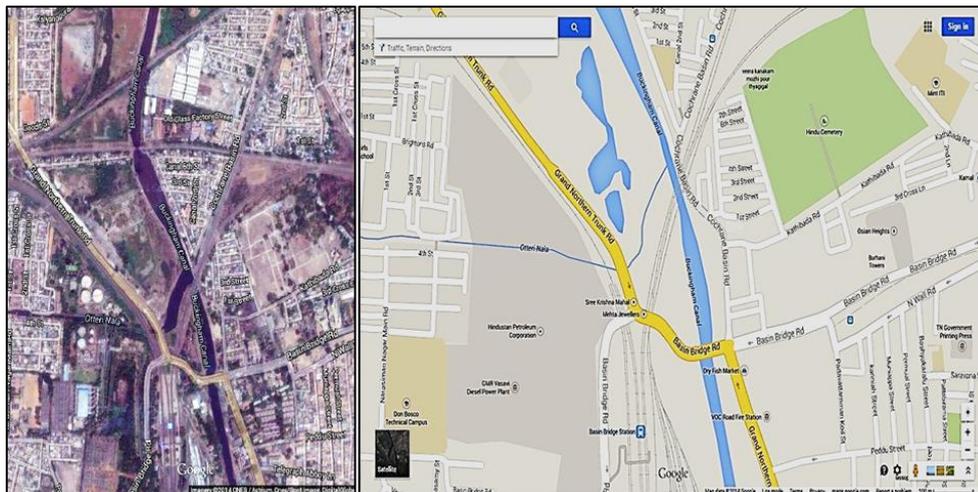
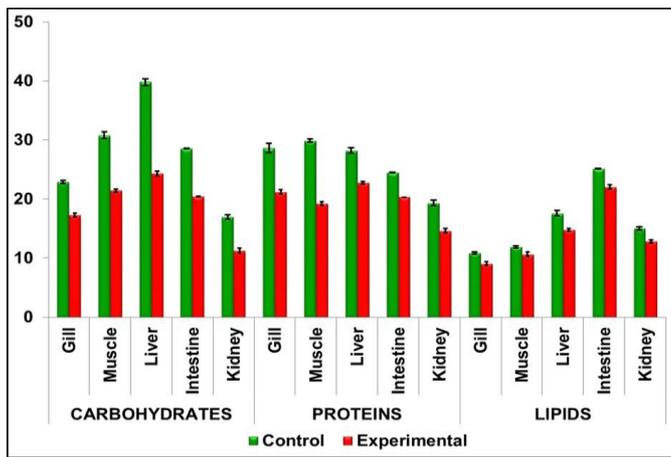


Fig 1: Study area - Buckingham canal

Table 1: Effect of aquatic toxicants on the biochemical parameters of different tissues of *Anabas testudineus*

Total Carbohydrate Content		
Tissue	Control fish	Experimental fish
Gill	22.90 ±0.30	17.29 ±0.29 (24.50%)*
Muscle	30.83 ±0.52	21.43 ±0.27 (30.48%)*
Liver	39.81 ±0.54	24.35 ±0.42 (38.28%)*
Intestine	28.56 ±0.02	20.45 ±0.04 (28.39%)*
Kidney	16.98 ±0.34	11.27 ±0.39 (33.63%)*
Total Protein Content		
Gill	28.67 ±0.82	21.20 ±0.36 (26.10%)
Muscle	29.90 ±0.31	19.25 ±0.31 (35.62%)*
Liver	28.23 ±0.46	22.76 ±0.26 (19.40%)*
Intestine	24.47 ±0.05	20.32 ±0.01 (16.95%)
Kidney	19.37 ±0.44	14.65 ±0.34 (24.37%)*
Total Lipid Content		
Gill	10.88 ±0.21	9.09 ±0.34 (16.45%)*
Muscle	11.91 ±0.18	10.64 ±0.37 (10.66%)
Liver	17.59 ±0.45	14.81 ±0.22 (15.80%)*
Intestine	25.15 ±0.06	22.05 ±0.35 (13.71%)
Kidney	15.05 ±0.28	12.84 ±0.27 (14.68%)*

Values expressed as mg/g wet weight of tissue; each value is mean ±standard error of five individual; Values in parenthesis denote per cent change; \*denote P<0.05 level of statistical significance.



**Fig 2:** Effect of aquatic toxicants on the biochemical parameters of *Anabas testudineus*. Values expressed as mg/g wet weight of tissue.

#### 4. Discussion

Carbohydrates are the chief sources of energy for any organism<sup>[5]</sup> and are found in large amounts in the liver and muscle tissues and in small amounts in kidney and heart. The decrease in carbohydrate content in the muscle and brain may be due to glucose utilization to meet excess energy demand imposed by severe anaerobic stress of mercury intoxication. A decrease in the glucose content of the liver, muscle and kidney tissue of *Clarias batrachus* was observed on exposure to sodium arsenite<sup>[6]</sup>. Neethirajan and Madhavan<sup>[7]</sup> also reported changes in the carbohydrate of liver, muscle, and gill tissue of *Mystus vittatus* when exposed to sumidon. Agarwal<sup>[8]</sup> noticed that mercuric chloride declined blood glucose of *Channa punctatus*. Srivastav *et al.*<sup>[9]</sup> investigated the effect of aldrin and carbamate on *Heteropneustes fossilis* and found a significant decline in blood glucose levels along with hyperchloremia and hypochloremia. Hypoglycemic condition may be due to increased muscular activity in the fish, which require more oxygen to meet the energy demand and consequently more amount of glucose rapidly utilized. Probably, the fish tries to meet this condition by increasing respiratory rate by drawing more amount of oxygen from contaminated water which may be the reason why low amount of blood glucose level are observed in pollution affected fish.

Vutukuru<sup>[10]</sup> reported an appreciable decline in different biochemical constituents (glycogen, total lipid and total protein levels) in liver, muscle and gills of *Labeo rohita* in chromium stressed condition when compared to that of control fish. The decrease in glycogen content of the tissues of *Labeo rohita* can be due to its enhanced utilization as an immediate source to meet energy demands under metallic stress. It could also be due to the prevalence of hypoxic or anoxic conditions, which normally enhances glycogen utilization<sup>[11]</sup>. The decrease in the glycogen content during the present study indicates its lipid utilization by the respective tissue as a consequence of pollutant stress. The results of the present study were comparable with the result of a study on *Channa punctatus*<sup>[12]</sup>; *Sarotherodon mossambicus*<sup>[13]</sup> exposed to chromium, and *Puntius conchoniis* to nickel<sup>[14]</sup>. Further, the observations made by Murty and Devi<sup>[15]</sup> in *Channa punctatus*, exposed to bleached kraft pulp mill effluent; and Bengeri *et al.*<sup>[16]</sup> in *Labeo rohita*, exposed to parathion supports the present findings in *Anabas testudineus*. Metals intoxication generally leads to glycogen depletion due to generalized disturbance of carbohydrate metabolism.

Tissue hypoxia and changes in the buffering system also leads to glycogen depletion in tissues<sup>[3]</sup>. The mechanism involved in the glycogen depletion may be due to glycogenolysis and inhibition of glycogenesis through stress induced release of adrenal catecholamine<sup>[17]</sup>. Glucose level significantly decreased in pollution affected *Anabas testudineus* from Buckingham canal in the present study. Similarly, a significant fall in blood glucose at 8ppm in *Clarias batrachus*<sup>[18]</sup> was attributed to the utilization of glucose by the tissues of extremely active fish to the enhanced production of insulin by the beta cells of the islet of Langerhans or of somatostatin by D cells of the pancreatic islet of Langerhans which has an inhibitory action<sup>[19]</sup>. In the present study, aquatic toxicant caused a significant fall in the level of total carbohydrates and the results were observed to be in consistent with previous reports.

Proteins are involved in major physiological events and therefore the assessment of the protein content can be considered as a diagnostic tool to determine the physiological phases of organism. Depletion of protein content has been observed in the muscle, intestine and brain of the fish *Catla catla* as a result of mercuric chloride toxicity<sup>[20]</sup>. Freshwater fish, *Cyprinus carpio* exposed to sub lethal concentration of cypermethrin showed marked changes in protein fractions of liver, brain and muscle tissue<sup>[21]</sup>. Similar results were observed in *Anabas testudineus* and *Anabas scandens*, when exposed to copper, lead nitrate and mercuric chloride<sup>[22]</sup>. De Smet and Blust<sup>[23]</sup> reported that proteolysis was intended to increase the role of proteins in the energy production during cadmium stress. A decrease in the total protein of the liver of *Channa punctatus* on tannery effluents exposure was observed<sup>[24]</sup>. A good number of references are available regarding decrease in the level of blood protein on the administration of sublethal dose of agrofane in *Oreochromis mossambicus*<sup>[25]</sup>; and heavy metals in *Perna viridis*<sup>[26]</sup>.

Proteins are the most fundamental and abundant biochemical constituent present in fishes. Proteins are the most important energy source to spare during chronic period of stress. Animal exposed to toxicants even at the sub-lethal levels experience great stress at the metabolic level during the period of detoxification of the toxicant<sup>[27]</sup>. Namrata *et al.*<sup>[28]</sup> observed depletion of proteins under exposure to kelthane, an organochlorine insecticide and in starvation of *Channa punctatus*. Depletion in the protein level in different tissues/organs of experimental animals was found under the stress of various metals in traces. Appreciable decrease in the protein level of tissue of *Channa punctatus* was noticed after the fish were exposed to heavy metals<sup>[29,30]</sup>. Protein depletion in liver and gonads of *Anabas testudineus* under the stress of nickel chloride was observed by Jha and Jha<sup>[31]</sup>. The present observation on total protein and free amino acid levels are in agreement with that of Jha<sup>[32]</sup>. The reduction in total protein content in the present study was further comparable with the findings of Gupta and Sastry<sup>[33]</sup> on *Heteropneustes fossilis*, exposed to mercuric chloride by Ramos and Herrera<sup>[34]</sup> on fish exposed to pesticides. The decreased protein level recorded during the present study was an indicative of increased proteolysis<sup>[35]</sup> resulting in to a shift in nitrogen metabolism<sup>[36]</sup>. Inhibition of ribosomal activity<sup>[37]</sup> resulting in protein degradation<sup>[38]</sup> is also one of the possible reasons for protein depletion.

Lipids are important constituent of cellular structures. Lipids are also essential for maintenance of normal cell permeability and structural integrity of cell membranes. Synthetic

pyrethroid and cypermethrin was found to induce a decrease in lipid content of liver and muscle tissue of *Labeo rohita* [39]. Govindan *et al.* [40] reported a significant decline in the level of total lipids in the muscle, liver and brain of *Gambusia affinis* exposed to a pesticide, phosphamidon. A decrease in total lipid content during the present study in gill, muscle, liver, intestine and kidney tissue denoted the effect of metals present in the Buckingham canal effluent on lipid as tissue specific of the species. The findings of Ram and Sathyanesan [41] on *Channa punctatus* intoxicated with mercuric chloride and Jha [32] on *Channa punctatus* under lead exposure support the present study. Loss of lipid may be a consequence of inhibition of lipid synthesis and mobilization of stored lipids [24, 42]. The decrease in tissue lipids and proteins might be partly due to their cell repair and tissue organization with the formation of lipoproteins, which are important cellular constituents of cell membranes and cell organelles present in cytoplasm [43]. Decrease in the lipid concentration observed in the pollution affected *Anabas testudineus* from Buckingham canal can also be attributed to its utilization in cell repair and cell organelles. The depletion in tissue proteins of *Channa punctatus* and *Oreochromis mossambicus* from Arakkonam lake may be due to impaired or nil protein synthesis and their pollution are due to their utilization in the formation of macroproteins, which are eliminated in the form of mucus by the fish for the direct and/or indirect utilization of proteins and lipids for energy needs were also reported [44, 45]. Normally carbohydrates, proteins and lipids which constitute the major components of the body play an important role in growth and energy metabolism. A reduction of carbohydrates, proteins and lipids were observed in *Labeo rohita* under the influence of heavy metal, lead [46]. The effluent from dye [47], tannery [48] and distillery [49] reported a severe reduction in the carbohydrate, protein and lipid contents among the fish population. Sujatha [50] observed that the total sugars and protein content in muscle, liver, brain and kidney revealed a mixed trend in *Catla catla* in the different experimental groups and different days of the exposure. It is clear from the results of the present study that there is an appreciable decline in different biochemical constituents of fish under pollution stress. Fishes are sensitive to contaminations of water, and pollutants that may significantly damage certain physiological and biochemical processes, when they enter the organs of these animals [51]. Several cases have been reported in which toxic effects of pollutants may be altered due to changes in pH, temperature, hardness and dissolved oxygen content of water [52]. The harmful effects especially sub-lethal effect of the pollutants retard growth and adversely affects the metabolic activity of surviving individual [53]. The biochemical profiles of glycogen, total protein and total lipid contents underwent a significant depletion of varying degree in the tissues of the pollution affected fish. During the present study, it was observed that metabolic levels were organs specific, indicating differential sensitivity of the tissues against the toxicant toxicities. Further, it was difficult to explain the changes under metallic stress in an organism since such alterations differ not only from metal and from species to species, but also from one experimental period to another [54]. Nair *et al.* [55] while studying biochemistry and haematology of *Anabas testudineus* found serious alterations in these factors when the fishes were exposed to titanium dioxide effluent. According to them, glycogen, being the chief source of energy for the fish is the first metabolite to be affected by any stress. It was further pointed out that the increased liver

glycogen suggests a stimulated glycogen synthesis contemporaneous with the increased liver glycogenolysis and subsequent blood glucose elevation. The dose-dependent hyperglycemic state indicated a typical stress response. Such a response might be due to general stress or it could also be a result of hypoxic state caused by a strictly mechanical and/or heavy metal action on the functioning of the gill. In general, more energy is required to cope up for the stressful condition. This energy may be obtained from the organic constituents as carbohydrates, proteins and/or lipids [56, 57]. Therefore during toxicant exposure, the fish tries to detoxify the toxicant by spending more energy and thereby showing a reduction in glycogen, protein and lipid contents.

A severe dysfunction was noticed in the liver and muscle tissue of *Anabas testudineus* exposed to aquatic pollutants. Significant depletion in glycogen, protein and lipid contents were also observed. The depletion in the metabolites indicated the fact that the whole metabolic pool of the fish gets disturbed/altered under the toxic stress. Further, the change in the biochemical profile indicates their rapid utilization to provide excess energy to cellular biochemical process in order to cope with the stressed condition. The results of the present study further confirm that aquatic pollution is one of the major reasons for decline in *Anabas testudineus* population and most probably in the population of the other freshwater fish residing in Buckingham canal.

## 5. Conclusion

The present study showed that aquatic pollutant induced alteration at the biochemical level, more pronounced changes acquiring or also time dependent this change may probably affect the enzyme mediated bio-defense mechanism of these fishes. Further research should focus on the effect of pollutant toxicity on *Anabas testudineus* at cellular and sub cellular levels.

## 6. References

1. Rosety-Rodriguez M, Rosety I, Jesus M. Erythrocyte antioxidant enzymes of gilthead as early-warning bio indicators of oxidative stress induced by malathion. *Haema*. 2005; 8(2):237-240.
2. Ghosh TK, Chatterjee SK. Effect of chromium on tissue energy reserve in freshwater fish, *Sarotherodon mossambicus*. *Environment and Ecology*. 1985; 3(2):178-179.
3. Shaffi SA. Cadmium intoxication on tissue glycogen content in three fresh water fishes. *Current Science*. 1978; 47:668-670.
4. Biswas B, Shah MS. Taxonomic comparison of local and Thai Koi (*Anabas testudineus*, Bloch) from Khulna, Bangladesh. *SAARC Journal of Agriculture*. 2009; 7(1):19-28.
5. Saravanan TS, Mohammed AM, Harikrishnan R. Studies on the chronic effects of endosulfan on blood and liver of *Oreochromis mossambicus*. *Journal of Ecological Research and Bioconservation*. 2000; 19(2):24-27.
6. Nimaichandra S, Trilochan M, Kumar NS. A Study on arsenic effects on blood and glucose concentration in a fish model. *Indian Journal of Environment and Ecoplaning*. 2005; 10(2):499-503.
7. Neethirajan K, Madhavan S. Sumidon induced changes on some aspects of metabolism in the chosen tissues of fresh water cat fish, *Mustus vittatus*. *Journal of Experimental Zoology, India*. 2004; 7(2):279-283.

8. Agarwal SK. Sub lethal effect of mercury chloride on some biochemical parameters of the blood in an air breathing fish *Channa punctatus* (Bloch.). *Journal of Environmental Biology*. 1992; 13(2):127-133.
9. Srivastav, Ajai K, Singh S, Sasayama Y. Vitamin D<sub>3</sub> induced changes in the prolactin cells of the fish, *Heteropneustes fossilis* reared in artificial freshwater, calcium-rich fresh water or calcium-deficient freshwater. *Journal of Reproductive Biology and Comparative Endocrinology*. 1995; 7:72-82.
9. Vutukuru SS. Acute effects of Hexavalent chromium on survival, oxygen consumption, hematological parameters and some biochemical profiles of the Indian major carp, *Labeo rohita*. *International Journal of Environmental Research and Public Health*. 2005; 2(3):456-462.
10. De Zwaan A, Zandee DI. Body distribution and seasonal changes in glycogen content of the common sea mussel *Mytilus edulis*. *Comparative Biochemistry and Physiology Part A: Physiology*. 1972; 43(1):53-58.
11. Sastry KV, Sunita K. Enzymological and biochemical changes produced by chronic exposure in teleost fish, *Channa punctatus*. *Toxicology Letters*. 1983; 16:9-15.
12. Jha BS, Pandey S. Alteration in the total carbohydrate level of intestine, liver and gonad induced by lead nitrate in the fish *Channa punctatus*. In: *Environmental risk assessment* (Eds.: Sahai YN, Deshmukh PN, Mathai TA, Patil KS). The Academy of Environmental Biology, 1989, 207-211.
13. Gill TS, Pant JC. Effect of sub-lethal concentration of mercury in a teleost, *Puntius conchonius*. *Biochemical and haematological responses*. *Journal of Experimental Zoology, India*. 1981; 19:571-573.
14. Murty AS, Devi AP. The effect of endosulfan and its isomers on tissue protein, glycogen and total lipids in the fish *Channa punctatus*. *Pesticide Biochemistry and Physiology*. 1982; 17(3):280-286.
15. Bengeri KV, Shivraj KM, Patil HS. Toxicity of dimethyl-parathion to freshwater fish *Labeo rohita* and oxygen uptake rate of exposed fish. *Environment and Ecology*. 1984; 2:1-4.
16. Jha V, Jayachandran C, Singh M, Singh S. Pharmacokinetic data on the oxytetracycline and its distribution in different biological fluids in female goats. *Veterinary Research Communications*. 1989; 13:11-16.
17. Paul DS, Walton FS, Saunders RJ, Styblo M. Characterization of the impaired glucose homeostasis produced in C57BL/6 mice by chronic exposure to arsenic and high-fat diet. *Environmental Health Perspectives*. 2011; 119:1104-1109.
18. Gorbman A, Dickhoff W, Vigna S, Ralph CL, Clark NB. *Comparative endocrinology*. New York: John Wiley & Sons, 1983.
19. Neff JM. *Polycyclic aromatic hydrocarbons in the aquatic environment: Sources, fates and biological effects*. Applied Science Publishers, London, UK, 1979.
20. Asztalos B, Nemcsok J, Rafeaie OJ. Effects of pesticide on some biochemical parameters of carp (*Cyprinus carpio*). *Archives of Environmental Contamination and Toxicology*. 1990; 19:275-282.
21. Chandravathy MV, Reddy SCN. *In vivo* recovery of protein metabolism in gill and brain of a freshwater fish, *Anabas scandens* after exposure to lead nitrate *Journal of Environmental Biology*. 1994; 15(1):75-82.
22. De Smet H, Blust R. Stress responses and changes in protein metabolism in carp, *Cyprinus carpio* during cadmium exposure. *Ecotoxicology and Environmental Safety*. 2007; 48:255-260.
23. Sivachandran R, Sultana M, Saravanan R, Malathy S. Tannery effluents induced DNA damage in liver and testicular tissues of fresh water fish, *Channa striatus*. *Asian Journal of Science and Technology*. 2014; 5(7):427-430.
24. Verma GP, Panigrahi P. Effect of agrofen on blood parameters of *Oreochromis mossambicus* (P). *Proceedings of the National Academy of Sciences, India*. 1998; 68B(1):29-36.
25. Vasanthi. Studies on the heavy metal induced toxicity in *Perna viridis* (L). Ph.D. thesis, University of Madras, Chennai, Tamil Nadu, India, 2012.
26. Yadav V, Sharma S, Harjai K, Mohan H, Chhibber S. Induction and resolution of lobar pneumonia following intranasal instillation with *Klebsiella pneumoniae* in mice. *Indian Journal of Medical Research*. 2003; 118:47-52.
27. Namrata S, Sanjay N, Pallavi C. Effects of starvation on the biochemical composition of freshwater fish, *Channa punctatus*. *Recent Research in Science and Technology*. 2011; 3(9):17-19.
28. Sen GM, Behera MK, Patel PN. Toxic effects of zinc on liver and brain of fish, *Channa punctatus* (Bloch). *Environment and Ecology*. 1992; 10(3):742-744.
29. Hota S. Arsenic toxicity to the brain, liver and intestine of freshwater fish, *Channa punctatus* (Bloch). *Geobios*. 1996; 23:154-156.
30. Jha BS, Jha MM. Biochemical effects of nickel chloride on the liver and gonads of the fresh water climbing perch, *Anabas testudineus* (Bloch). *Proceedings of the National Academy of Sciences, India. Section B (Biological sciences)*: 1995; 65(1):39-46.
31. Jha BS. Alteration in the protein and lipid content of intestine, liver and gonads in the lead exposed freshwater fish *Channa punctatus* (Bloch). *Indian Journal of Environment and Ecoplanning*. 1991; 2(3):281-284.
32. Gupta PK, Sastry KV. Effect of mercuric chloride on enzyme activities in the digestive system and chemical composition of liver and muscles of the cat fish, *Heteropneustes fossilis*. *Ecotoxicology and Environmental Safety*. 1981; 5(4):389-400.
33. Ramos P, Herrera E. Comparative responsiveness to prolonged hyperinsulinemia between adipose-tissue and mammary-gland lipoprotein lipase activities in pregnant rats. *Early Pregnancy: Biology and Medicine*. 1996; 2:29-35.
34. Rao CS, Neeraja P. Effect of chronic ammonia stress on some aspects of protein metabolism of fish *Tilapia mossambica*. *Indian Journal of Comparative Animal Physiology*. 1990; 8:69-73.
35. Rao SK, Moorthy SK, Naidu DM, Chetty CS, Swami KS. Changes in nitrogen metabolism in tissues of fish, *Sarotheredon mossambicus* exposed to benthocarb. *Bulletin of Environmental Contamination and Toxicology*. 1983; 40:473-478.
36. De Bruin A. *Biochemical toxicology of environmental agents*. Elsevier, North Holland Biochemical press, New York, USA, 1976.
37. Shakoori AR, Zaheer SA, Ahmad MS. Effect of malathion, dieldrin and endrin on blood serum proteins and free amino acids pool of *Channa punctatus*. *Pakistan Journal of Zoology*. 1976; 8:125-134.

38. Das BK, Mukherjee SC. Toxicity of cypermethrin in *Labeo rohita*, fingerlings: Biochemical: Enzymatic and hematological consequences. *Comparative Biochemistry and Physiology*. 2003; 13(4):109-121.
39. Govindan VS, Jacob L, Devika R. Toxicity and metabolic changes in *Gambusia affinis* exposed to phosphamidon. *Journal of Ecotoxicology and Environmental Monitoring*. 1994; 4(1):1-6.
40. Ram RN, Sathyanesan AG. Mercuric chloride induced changes in protein, lipid and cholesterol levels of liver and ovary of the fish *Channa punctatus*. *Environment and Ecology*. 1984; 2:111-113.
41. Mani K, Saxena PK. Effects of safe concentration of some pesticides on ovarian recrudescence in the fresh water murrel *Channa punctatus*: A qualitative study. *Ecotoxicology and Environmental Safety*. 1985; 9:241-249.
42. Harper. *Harpers Review of Biochemistry* (Ed.) Meyes DWPA, Rowell VW. Lange Medical Publications. Maruzen Asia, 1983.
43. Nagai M, Ikeda S. Carbohydrate metabolism in fish. I. Effect of starvation and dietary composition on the blood glucose level and the hepatopancreatic glycogen and lipid content in carp. *Nippon Suisan Gakkaishi*, 1971; 37:404-409.
44. Dheenadayalamurthy. Characterization and impact of aquatic pollutants on the biochemical, histopathological and molecular changes in various tissues of a fish, *Channa punctatus* and *Oreochromis mossambicus*. Ph.D. thesis, University of Madras, Chennai, Tamil Nadu, India, 2012.
45. Neha-Bhatkar, Venkhede GN, Dhande RR. Heavy metal induced biochemical alterations in the fresh water fish *Labeo rohita*. *Journal of Ecotoxicology and Environmental Monitoring*. 2004; 14(1):1-7.
46. Baskaran P, Palanichamy S, Arunachalam S. Effects of textile dye effluent on feeding energetics body composition and oxygen consumption. *Journal of Ecobiology*. 1989; 1:203-204.
47. Ambrose T, Kumar LCA, Vincent S, Lambert R. Biochemical responses of *Cyprinus carpio communis* to toxicity of tannery effluent. *Journal of Ecobiology*. 1994; 6(3):213-216.
48. Maruthi YA, Rao SMV. Effect of distillery effluent on biochemical parameters of fish *Channa punctatus* (Bloch). *Journal of Environmental Pollution*. 2000; 7(2):111-113.
49. Sujatha LB. Studies on the physiology, haematology and histology in the Indian major carp, *Catla catla* (Hamilton) as influenced by individual and synergistic toxic effects of a pesticide and two metallic compounds. Ph.D. thesis, University of Madras, Chennai, Tamil Nadu, India, 2006.
50. Nemcsok J, Orban L, Asztalos B, Vig E. Accumulation of pesticides in the organs of carp (*Cyprinus carpio*) at 4°C and 20°C. *Bulletin of Environmental Contamination and Toxicology*. 1987; 38:370-378.
51. Pascoe D, Evans S, Woodworth J. Heavy metal toxicity to fish and the influence of water hardness. *Archives of Environmental Contamination and Toxicology*. 1986; 15:481-485.
52. Weis JS, Weis P. Effects of heavy metals on development of the killifish *Fundulus heteroclitus*. *Journal of Fish Biology*. 1977; 11:49-54.
53. Van der Oost R, Beyer J, Vermeulen NPE. Fish bioaccumulation and biomarkers in environmental risk assessment: a review. *Environmental Toxicology and Pharmacology*. 2003; 13:57-149.
54. Nair GA, Nair VNB, Suryanarayanan H, Radhakrishnan S. Effects of titanium effluents on the peripheral haematology of *Anabas testudineus* (Bloch). *Proceedings of the National Academy of Sciences, India*. 1984; 50(9):555-558.
55. Ganesan S. Hydrobiology, biodiversity and ecotoxicological impact on the biochemical, histopathological and molecular changes in a fish inhabiting the Chrompet lake, Chennai, Tamil Nadu, India. Ph.D. thesis, Bharathiar University, Coimbatore, Tamil Nadu, India, 2010.
56. Tulasi G, Rao JK. Effect of chromium on protein metabolism in different tissues of fish, *Cyprinus carpio*. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. 2013; 4(1):143-148.